

Three new species of foetid *Gymnopus* in New Zealand

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Abstract

We describe three new species, *Gymnopus imbricatus*, *G. ceraceicola* and *G. hakaroa*, from New Zealand that are similar to *G. foetidus* (= *Micromphale foetidum*), growing on wood, with an insititious stipe and foetid odour. The position of these species within the /gymnopoide clade is confirmed by ITS sequence analysis.

Key words

Gymnopus, *Micromphale*, New Zealand

Introduction

The new species we describe are members of the family Omphalotaceae Matheney et al. (2006) and have the morphological characteristics of *Gymnopus* section *Vestipedes* subsection *Impudicae* (Antonín & Noordeloos 2010) which contains *Micromphale foetidum* (Sowerby) Singer, the type species of the formerly recognised genus *Micromphale* (e.g. in the sense of Singer 1986). Fruitbodies of the group often have a foetid odour when crushed, described as like rotting cabbage or garlic. The only existing records of this group in New Zealand were found to be misapplications of names applied to northern hemisphere species.

Moncalvo et al. (2002) investigated nLSU rDNA sequence data for a large number of agarics and recognised a /micromphale clade containing *M. foetidum* (AF261328). The clade also contained *Gymnopus* pro parte, *Caripia*, *Setulipes* and *Micromphale*.

Their /micromphale clade was nested within a broader /lentinuloid clade including *Rhodocollybia*, *Marasmiellus ramealis* (Bull.) Singer, *Marasmius scorodoni* (Fr.) Fr. and *Lentinula*. Mata et al. (2004), based on an LSU analysis, also identified a clade containing sequences of *M. foetidum* and material named as *Setulipes androsaceus* (L.) Antonín and *Gymnopus fusipes* (Bull.) Gray, the type species of those respective genera. They adopted a broad concept of *Gymnopus* incorporating these genera together with *Marasmiellus*. Wilson and Desjardin (2005) used LSU to examine the group and identified a /gymnopus clade containing *G. fusipes*, *M. foetidum*, *S. androsaceus* at its core with *Micromphale perforans* (Hoffm.) Gray lying on its boundary. These results were broadly supported by Mata et al. (2006) in their analysis using ITS1–5.8–ITS2 but they demonstrate clustering of *G. fusipes*, *S. androsaceus* and *Micromphale* on the periphery of a concentration of *Gymnopus*-labelled samples. On the basis of these results the currently generally accepted concept of *Gymnopus* is broad (e.g. Noordeloos 2012), and incorporates a number of previously recognised genera. Hughes et al. (2010) erected the genus *Connopus* to accommodate the *Gymnopus acervatus* group within the gymnopoid clade and presented LSU and ITS data indicating its placement close to *Rhodocollybia*. Their LSU analysis supports a core gymnopoid clade containing *G. fusipes*, *S. androsaceus*, which once again places *Micromphale foetidum* and *M. perforans* on a boundary with a sister group containing *Rhodocollybia*, *Marasmiellus juniperinus* Murrill and various *Gymnopus* species. The /gymnopus, clade as interpreted by Hughes et al., contains significant substructure. A multi-gene analysis including more representatives may indicate the recognition of further segregates at genus-level. In this paper we accept our newly described species within the current broad concept of *Gymnopus* whilst recognising their close alliance to the historical concept of the genus *Micromphale*.

For this study we analysed ITS1–5.8–ITS2 data for related New Zealand collections together with representative sequences from Genbank, many from the studies cited above. The structure of our ITS tree is consistent with these previous analyses, and once again identifies a /micromphale clade closely linked to core *Gymnopus* species. ITS data generated for a number of representative collections of our newly described taxa support species concepts based on morphology.

Materials and methods

Morphological protocols

Spore dimensions are stated as the mean \pm 1.5 SD of 20 measurements, thus covering 86% of measurements under an assumed normal distribution model. Fresh or dried material was examined mounted in 10% KOH or Melzer's reagent. Material was hand-sectioned. Some micrographs were obtained under DIC conditions. Measurements were always taken without DIC optics and an extended objective iris in order to maximise boundary contrast.

Phylogenetic protocols

DNA extraction and sequencing followed the protocols outlined in Cooper and Leonard (2012). We downloaded from Genbank selected sequences used in cited publications, together with close BLAST matches, Table 1. General sequence management was carried out using Geneious (Drummond et al. 2011). Data exchange between applications was facilitated using Alter (Glez-Peña et al. 2010). Sequence alignment was carried out using MAFFT within Geneious (Kato et al. 2002). A maximum likelihood analysis was executed using RAxML (Stamatakis 2006), with 100 bootstrap runs, launched from Topali 2.5 (Milne et al. 2004). The substitution model of GTR+G was recommended by Topali 2.5. We selected a sequence of *Anthracoephyllum archeri* (Berk.) Pegler as the outgroup.

Table 1. ITS Sequences used in the analysis. New sequences generated for this analysis are in bold.

Genbank #	Collection #	Organism	PDD Voucher#	Country
DQ444308	TENN50049	<i>Anthracoephyllum archeri</i>		New Zealand
DQ480112	TENN58672	<i>Gymnopus alkalivirens</i>		Greenland
DQ480114	TENN55834	<i>Gymnopus alpinus</i>		Scotland
AY256691	TENN57012	<i>Gymnopus aquosus</i>		Germany
DQ449971	TENN59738	<i>Gymnopus aquosus</i>		USA
KC248409	PL6304	<i>Gymnopus ceraceicola</i>	PDD 101750	New Zealand
KC248389	PL126406	<i>Gymnopus ceraceicola</i>	PDD 101754	New Zealand
KC248400	PL189402	<i>Gymnopus ceraceicola</i>	PDD 76358	New Zealand
KC248403	JAC9334	<i>Gymnopus ceraceicola</i>	PDD 80771	New Zealand
KC248405	JAC10084	<i>Gymnopus ceraceicola</i>	PDD 87181	New Zealand
KC248404	JAC10336	<i>Gymnopus ceraceicola</i>	PDD 87424	New Zealand
KC248394	JAC10395	<i>Gymnopus ceraceicola</i>	PDD 87483	New Zealand
KC248408	JAC10817	<i>Gymnopus ceraceicola</i>	PDD 87661	New Zealand
KC248392	RHP13063	<i>Gymnopus ceraceicola</i>	PDD 90101	New Zealand
KC248391	KWH12891	<i>Gymnopus ceraceicola</i>	PDD 90119	New Zealand
KC248393	RHP12871	<i>Gymnopus ceraceicola</i>	PDD 90132	New Zealand
KC248397	JAC11005	<i>Gymnopus ceraceicola</i>	PDD 95459	New Zealand
KC248395	JAC11093	<i>Gymnopus ceraceicola</i>	PDD 95544	New Zealand
AY256690	TENN57012	<i>Gymnopus dryophilus</i>		USA
DQ449974	TENN58087	<i>Gymnopus dryophilus</i>		Costa Rica
AF505778	TENN 59141	<i>Gymnopus dysodes</i>		Costa Rica
AY256694	TENN59457	<i>Gymnopus earleae</i>		USA
DQ449973	TFB10718	<i>Gymnopus exculptus</i>		Greenland
AF505780	FB11434	<i>Gymnopus foetidum</i>		USA
AY256710	TENN59217	<i>Gymnopus fusipes</i>		France
KC248407	JAC9585	<i>Gymnopus bakaroa</i>	PDD 81086	New Zealand
KC248410	JAC10225	<i>Gymnopus bakaroa</i>	PDD 87315	New Zealand
KC248411	PL25404	<i>Gymnopus imbricatus</i>	PDD 101753	New Zealand
KC248406	JAC10089	<i>Gymnopus imbricatus</i>	PDD 87186	New Zealand

Genbank #	Collection #	Organism	PDD Voucher#	Country
KC248398	JAC10310	<i>Gymnopus imbricatus</i>	PDD 87398	New Zealand
KC248401	JAC10322	<i>Gymnopus imbricatus</i>	PDD 87410	New Zealand
KC248399	JAC10815	<i>Gymnopus imbricatus</i>	PDD 87659	New Zealand
KC248402	JAC10816	<i>Gymnopus imbricatus</i>	PDD 87660	New Zealand
KC248396	JAC10495	<i>Gymnopus imbricatus</i>	PDD 87675	New Zealand
KC248390	JAC11038	<i>Gymnopus imbricatus</i>	PDD 95489	New Zealand
AF505779	TENN56658	<i>Gymnopus impudicus</i>		Costa Rica
DQ449986	Duke RV94154	<i>Gymnopus ioccephalus</i>		USA
AY256693	TENN59532	<i>Gymnopus junquilleus</i>		USA
DQ449960	TENN50620	<i>Gymnopus ocior</i>		Switzerland
DQ449972	TENN56321	<i>Gymnopus subsulphureus</i>		USA
AY263453	AWW115	<i>Gymnopus vitellinipes</i>		Java/Bali
AY256708	TENN59540	<i>Marasmiellus juniperinus</i>		USA
GU234007	JB14	<i>Marasmius androsaceus</i>		Sweden
DQ444312	TENN50482	<i>Marasmius androsaceus</i>		UK
DQ444311	TENN50704	<i>Marasmius androsaceus</i>		USA
DQ449990	TENN59293	<i>Micromphale brassicolens</i>		Austria

Results

Our analysis places the New Zealand taxa in a monophyletic clade close to *G. foetidum* and *G. brassicolens* historically recognised in the genus *Micromphale* (Fig. 1). The combination of sequence data and morphological analysis of many collections indicate two major groups which we equate with the newly described species *G. imbricatus* and *G. ceraceicola*. In addition we recognise a further species, *G. hakaroa*, which is poorly distinguished from *G. imbricatus* on the basis of ITS sequences but which is morphologically consistently different. Minor sequence variation in the *G. ceraceicola* group does not correlate with morphology and we choose to recognise these specimens as a single species. More information and images of collections may be found on the Landcare Research website (Systematics Collections Data).

Gymnopus ceraceicola J.A. Cooper & P. Leonard, sp. nov.

http://species-id.net/wiki/Gymnopus_ceraceicola

Holotype: PDD 87181. Registration identifier: IF550091

Diagnosis. *G. ceraceicola* is distinguished from related New Zealand species by the combination of pruinose, central stipe and dark pileus.

Macromorphology. Pileus 5–20 mm, generally broadly convex to applanate, but sometimes campanulate when young, brick to purplish chestnut, minutely felty, radially furrowed and striate towards the margin, margin slightly fimbriate. Lamellae cream, creamy yellow to vinaceous buff, waxy, adnate. Lamellae present, in series of

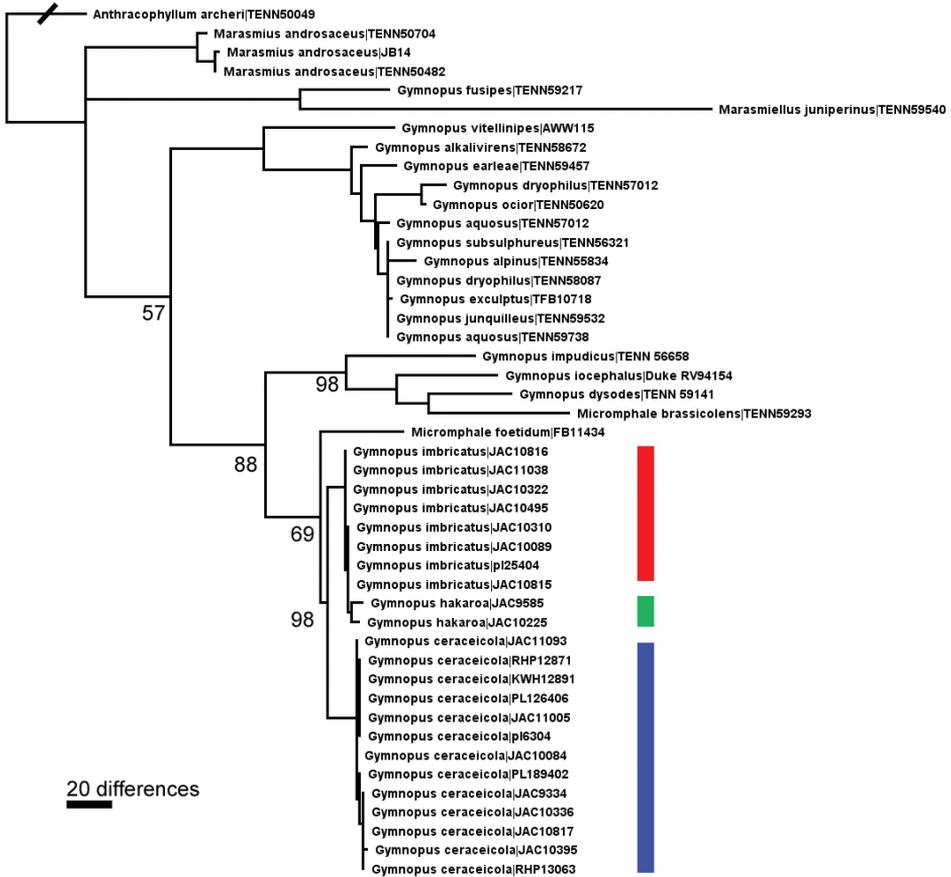


Figure 1. Maximum likelihood cladogram of selected ITS sequences, with bootstrap proportion. Red bar = *Gymnopus imbricatus*, green bar = *G. hakaoroa*, blue bar = *G. ceraceicola*

three: intercalated short/long/short. Stipe central, cartilaginous, 10–20 × 1–2 mm, equal, brown vinaceous, sometimes paler towards apex or base, always entirely finely pruinose. Stipe base insititious and always associated with a thin waxy to chalky cream layer of partially gelatinised hyphae covering the substrate. This layer is often extensive, with a distinct margin, and often green with algal cells. Fruitbodies with garlic/rotten cabbage smell, especially when crushed.

Micromorphology. Pileipellis a partially gelatinised radially arranged clamped cutis of smooth hyphae to 5 µm diameter, with brown extra-cellular encrustation. Epidermal layer to 140 µm. Subepidermis of thick glassy-walled non-gelatinised smooth hyaline hyphae, weakly dextrinoid. Basidia clavate to 40 × 8 µm. Sterigmata to 7 µm, 4-spored. Basidioles cylindrical, tapering towards apex, 40 × 4 µm. Spores hyaline, lacrymoid, $7.9 \pm 1 \times 4.5 \pm 0.6$ µm, $Q = 1.8 \pm 0.1$ including apiculus. Cheilocystidia and pleurocystidia not observed. Stipitipellis a cutis of brown parallel hyphae, to 5 µm wide. Caulocystidia smooth, hyaline, agglutinated into fascicles.



Figure 2. *Gymnopus ceraceicola* Holotype, PDD 87181. Fruitbodies.

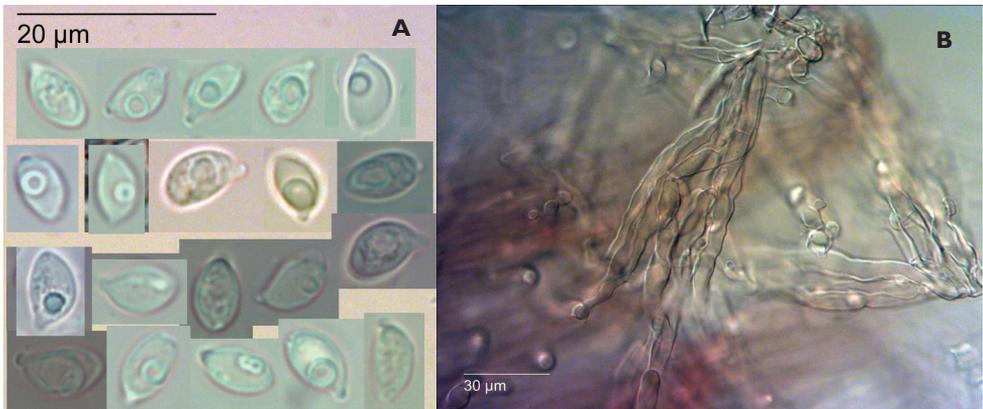


Figure 3. *Gymnopus ceraceicola* Holotype, PDD 87181. **A** Spores (KOH) **B** Agglutinated fascicles of caulocystidia on stipe (KOH).

Habitat. Colonies of a few to hundreds of fruitbodies on bark of fallen, dead branches and twigs, especially *Nothofagus*.

Distribution. Broadly distributed and common in both North and South Islands of New Zealand.

Etymology. Ceraceicola, indicating association with a basal waxy layer, although this feature is common to the three species described here.

Notes. Sequence data indicate variability in the taxon but the morphological details are constant and we choose to recognise a single species. New Zealand records of *Gymnopus (Micromphale) foetidum* and *Gymnopus (Micromphale) brassicolens* are attributable to *G. ceraceicola*. Authentic New Zealand material of these two species has not been identified. *Gymnopus brassicolens* has paler pileus colours, non-gelatinized pileipellis, cheilocystidia and pileipellis elements with lateral projections, and larger basidiospores. *Gymnopus foetidus* is macroscopically similar but does not possess the agglutinate fascicles of caulocystidia of *G. ceraceicola*.

Specimens examined. New Zealand, North Island: PDD 40852, on dead wood, Anawhata Rd., Waitakare Ranges, Collector P.R. Johnston & G. Samuels, 9 June 1981. PDD 80771, on dead wood of *Beilschmiedia tawa*, Erua Forest, Taupo, Collector J.A. Cooper (JAC9334), 4 April 2005. PDD 87382, on dead wood of *Nothofagus fusca*, Mt Holdsworth, Gentle Annie Track, Wairarapa, Collector J.A. Cooper (JAC10294), 11 May 2007. PDD 87483, on wood, Mt Holdsworth, Donnelly Flat Loop Track, Wairarapa, Collector G. Gates & D. Ratkowsky (JAC10395), 7 May 2007. PDD 87424, on dead bark of *Nothofagus*, Mt Holdsworth, Gentle Annie Track, Wairarapa, Collector J.A. Cooper (JAC10336), 11 May 2007. PDD 95544, on bark of *Nothofagus fusca*, Rimutaka Forest Park, Wellington, Collector J.A. Cooper (JAC11093), 14 May 2009. PDD 95545, on bark of dead branch of *Nothofagus fusca*, Rimutaka Forest Park, Wellington, Collector J.A. Cooper (JAC11094), 14 May 2009.

New Zealand, South Island: PDD 76357, on dead twig of *Nothofagus*, Canaan Road Track, Nelson, Collector P.L. Leonard, 30 April 2002. PDD 96730, on dead wood, Wangapeka, Nelson, collector P.L. Leonard (PL126406), 14 April, 2006. PDD 90101=TENN 061068, on bark, vicinity of Seddonville, Charming Creek Track, Nelson, Collector R.H. Petersen (RHP 13063), 11 May 2006. PDD 76358, on bark on dead branch of *Nothofagus menziesii*, Lake Daniels Track, Nelson, Collector P.L. Leonard (PL189402), 2 April 2002. PDD 95459, on bark of dead branch of *Nothofagus solandri*, Kowai Bush, Springfield, Mid Canterbury, Collector J.A. Cooper (JAC11005), 2 May 2009. PDD 95462, on bark of dead branch of *Nothofagus solandri*, Kowai Bush, Springfield, Mid Canterbury, Collector J.A. Cooper (JAC11008), 2 May 2009. **Holotype** PDD 87181, on dead branch of *Nothofagus fusca*, Hinewai Reserve, Akaroa, Mid Canterbury, Collector J.A. Cooper (JAC10084), 3 June 2006. PDD 87661, on dead twigs of *Leptospermum scoparium*, Government Track, Waipori Falls Road, Dunedin, Collector K. Soop (JAC10817), 12 May 2008. PDD 96636, on dead wood of *Nothofagus solandri*, Lake Hauroko, Fiordland, Collector P. White (JAC12522), 7 May 2012. PDD 90119 =TENN061007, on twigs, Vicinity of Te Anau, Kepler Track from Rainbow Reach, Fiordland, Collector K.W. Hughes (KWH12891), 30 April 2006. PDD 90132=TENN060986, vicinity Manapouri, Borland Lodge, Nature Track, Fiordland, Collector R.H. Petersen (RHP12871), 29 April 2006.

***Gymnopus imbricatus* J.A. Cooper & P. Leonard, sp. nov.**

http://species-id.net/wiki/Gymnopus_imbricatus

Holotype: PDD 95489. Registration identifier: IF550092

Diagnosis. *G. imbricatus* is distinguished from related New Zealand species by the smooth stipe, larger basidiospores, and imbricate habit.

Macromorphology. Pileus 3–20 mm in diameter convex, cream to fawn, minutely felty, radially furrowed and striate towards the margin, margin fimbriate. Lamellae cream to creamy yellow, adnate. Lamellae present, in series of two: short/long. Stipe mostly eccentric, cartilaginous, to 3 × 0.5 mm, equal, umber to black, sometimes paler



Figure 4. *Gymnopus imbricatus*. **A** Holotype PDD 95489. Fruitbodies, scale 1 cm **B** PDD 87186. Scale 1 mm.

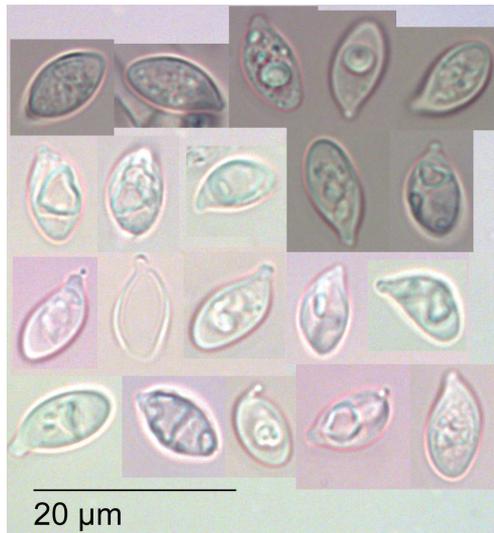


Figure 5. *Gymnopus imbricatus* Holotype PDD 95489. Spores (in KOH).

towards base, always entirely smooth. Stipe base insititious and usually associated with a thin waxy to chalky cream layer of partially gelatinised hyphae covering the substrate, usually green with algal cells. Fruitbodies with garlic/rotten cabbage smell, especially when crushed.

Micromorphology. Pileipellis a partially gelatinised irregular clamped cutis of hyphae 4 μm diameter, without intra or extracellular pigmentation, terminal layer with gelatinised coralloid elements, to 2 μm wide, and occasional small finger-like trichodermal elements to 20 μm . Epidermal layer to 25 μm . Subepidermis of thick glassy-walled non-gelatinised smooth hyaline hyphae, weakly dextrinoid. Basidia clavate to 50 \times 10 μm . Sterigmata to 5 μm , 4-spored. Basidioles to 50 \times 6 μm cylindrical and tapered towards apex. Spores hyaline, lacrymoid 9.8 \pm 1.2 \times 5.1 \pm 0.4 μm , Q = 1.9 \pm 0.3 including apiculus. Cheilocystidia and pleurocystidia not observed. Stipitipellis a cutis of parallel brown hyphae, to 6 μm wide. Caulocystidia absent.

Habitat. Forming imbricate colonies of dozens to hundreds of fruitbodies on bark and decorticate wood of dead branches and twigs, especially *Kunzea* and *Leptospermum* but occurs with other trees. Also occurs at the stem base of live trees.

Distribution. Broadly distributed and common in both North and South Islands of New Zealand.

Etymology. Imbricatus, pertaining to the often tiered and overlapping eccentrically stemmed caps.

Specimens examined. New Zealand, North Island: PDD 80766, on bark of *Beilschmiedia tawa*, Erua Forest, Taupo, collector J.A. Cooper (JAC9329), 4 April, 2005. PDD 87398, bark on dead branch of *Nothofagus*, Waiohine Gorge, Wairarapa, Collector J.A. Cooper (JAC10310), 10 May 2007. PDD 87410, dead stems of *Ripogonum scandens*, Waiohine Gorge, Wairarapa, Collector J.A. Cooper (JAC10322), 10 May 2007.

New Zealand, South Island: PDD 101753, dead branches of *Nothofagus menziesii*, Riwaka Resurgence, Nelson, Collector P.L. Leonard (PL25404), 10 April, 2006. PDD 96141, dead twigs of *Kunzea ericoides*, Mt Fyffe Track, Kaikoura, collector J.A. Cooper (JAC11734), 26 Feb. 2011. PDD 80154, dead log of *Nothofagus menziesii*, Lewis Pass, Buller, collector J.A. Cooper (JAC8287), 24 November, 2001. PDD 80157, on dead de-corticate log, Lyell Walkway, Nelson, collector J.A. Cooper (JAC80157), 25 November, 2001. PDD 87675, living stem of *Fuchsia excorticata*, Saddle Hill, Mid Canterbury, Collector J.A. Cooper (JAC10495), 22 May 2005. **Holotype** PDD 95489 (Figs 4 and 5), base of live trees of *Kunzea ericoides*, Kennedy's Bush, Mid Canterbury, Collector J.A. Cooper (JAC11038), 24 May 2009. PDD 79799, bark of dead tree, Kennedy's Bush, Mid Canterbury, Collector J.A. Cooper (JAC8921), 20 March 2004. PDD 87186, on bark of living tree of *Kunzea ericoides*, Hinewai Reserve, Akaroa, Mid Canterbury, Collector J.A. Cooper (JAC10089), 3 June 2006. PDD 87660, fallen log, Racemans Track, Silverstream Valley, Dunedin, Collector S. Dodd (JAC10816), 13 May 2008. PDD 87659, on dead twigs of *Kunzea ericoides*, Evansdale Glen, Dunedin, Collector P.R. Johnston (JAC10815), 12 May 2008.

***Gymnopus hakaroa* J.A. Cooper & P. Leonard, sp. nov.**

http://species-id.net/wiki/Gymnopus_hakaroa

Holotype: PDD 87315. Registration identifier: IF550093

Diagnosis. *G. hakaroa* is distinguished from *G. ceraceicola* by smaller stature and a pruinose stipe lacking fascicles of agglutinate caulocystidia. It is distinguished from *G. imbricatus* by non-imbricate growth, a consistently central stipe, and smaller basidiospores.

Macromorphology. Pileus 3–10 mm diam. convex, rusty tawny to umber, minutely felty, weakly radially furrowed and striate towards the margin. Lamella cream to yellow, waxy. Lamellae present, in series of three: intercalated short/long/short. Stipe central, cartilaginous, to 5 × 0.6 mm, equal, umber to black, paler towards base, smooth to minutely pruinose. Stipe base insititious and always associated with an obvious waxy to chalky cream layer of partially gelatinised hyphae covering the substrate,

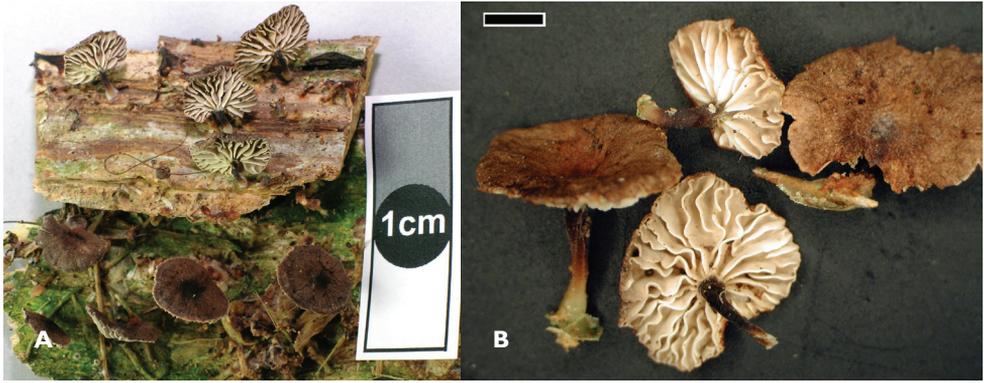


Figure 6. *Gymnopus hakaroa* **A** PDD 81086. Fruitbodies **B** scale= 2 mm

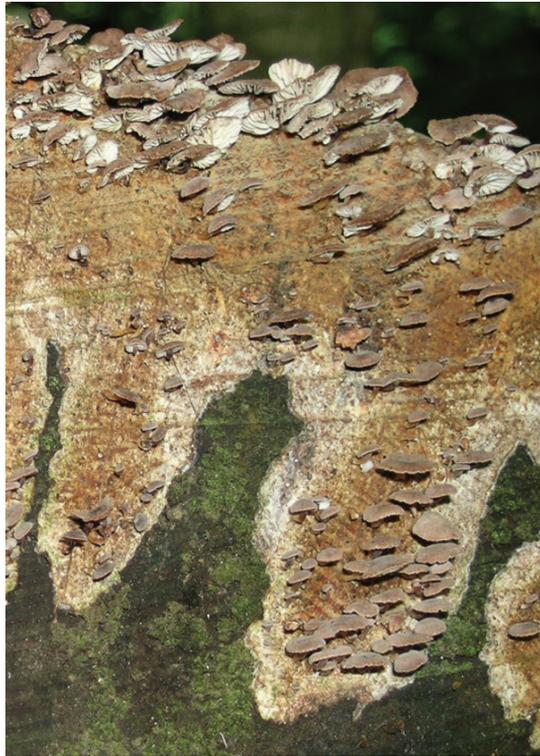


Figure 7. *Gymnopus hakaroa* Hologotype PDD 87315. Fruitbodies, showing waxy substratum.

usually green with algal cells. Fruitbodies with garlic/rotten cabbage smell, especially when crushed.

Micromorphology. Pileipellis a partially gelatinised radially arranged clamped cutis of smooth hyphae to 3 μm in diameter, with brown extra-cellular encrustation. Epidermal layer to 80 μm . Subepidermis of thick glassy-walled non-gelatinised smooth hyaline hyphae, to 3 μm in diameter, weakly dextrinoid. Basidia clavate to 40 \times 8 μm .

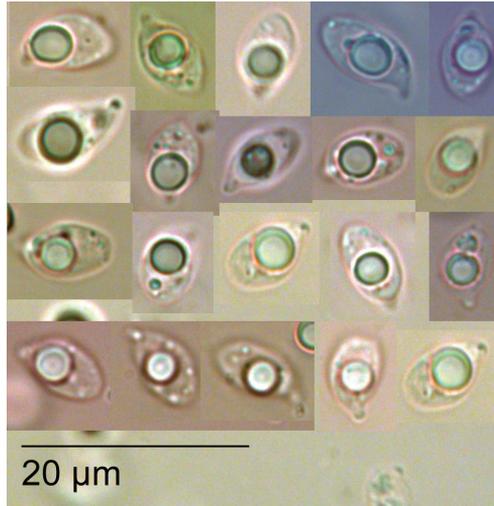


Figure 8. *Gymnopus hakaroa* Holotype PDD 87315. Spores (KOH).

Sterigmata to 7 μm , 2–4-spored. Basidioles cylindrical and tapered towards apex 40 \times 6 μm . Spores hyaline, lacrymoid $8.3 \pm 1 \times 4.8 \pm 0.3 \mu\text{m}$, including apiculus, $Q = 1.7 \pm 0.2$. Cheilocystidia and pleurocystidia not observed. Stipitipellis a cutis of hyaline to pale brown hyphae, to 5 μm wide. Stipe without caulocystidia.

Habitat. Forming imbricate colonies of dozens to hundreds of fruitbodies on decorticate dead wood.

Distribution. Currently *G. hakaroa* is only known from a single location on the Canterbury Port Hills in the South Island of New Zealand.

Etymology. Hakaroa, a Maori name for the Bank's Peninsula region of New Zealand.

Notes. Sequence data (Fig 1) indicates a close phylogenetic relationship to *G. imbricatus* but there are consistent and substantial morphological differences.

Specimens examined. New Zealand, South Island: Holotype PDD 87315 (Figs 6 and 7) on dead log, Kennedys Bush Reserve, Port Hills, Mid Canterbury, Collector J.A. Cooper (JAC10225), 11 Feb. 2007. PDD 81086 (Fig. 8), on dead wood of *Kunzea ericoides*, Kennedys Bush Reserve, Port Hills, Mid Canterbury, Collector J.A. Cooper (JAC9585), 23 July, 2007. PDD 96390, on dead decorticate log of *Melicytus ramiflorus*, Kennedys Bush Reserve, Port Hills, Mid Canterbury, Collector J.A. Cooper (JAC11301), 17 April, 2010.

Dicussion

Gymnopus imbricatus, as its name suggests forms dense populations of small imbricate fruitbodies. It is most commonly associated with tea-tree (*Kunzea ericoides* and *Leptospermum scoparium*) and often found on the bark at the base of living trees. *Gymnopus hakaroa* is larger, with a dark minutely pruinose cap and again forms dense populations

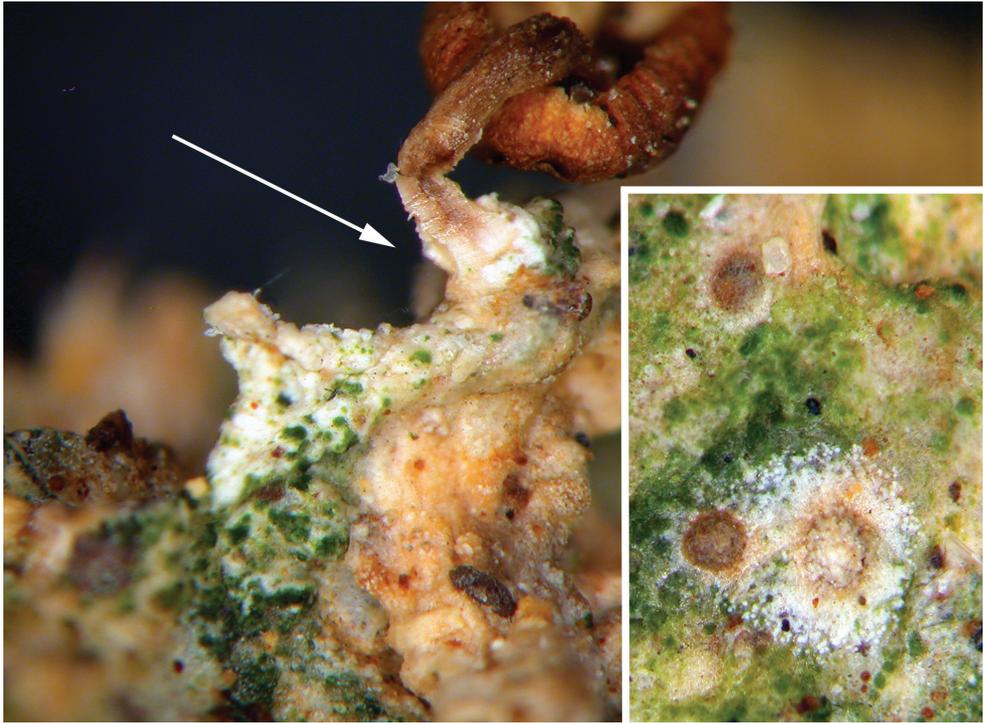


Figure 9. *Gymnopus hakaroa* PDD 96390. Stipe base (arrow) with surrounding algal mat. Inset, primordial arising from algal mat.

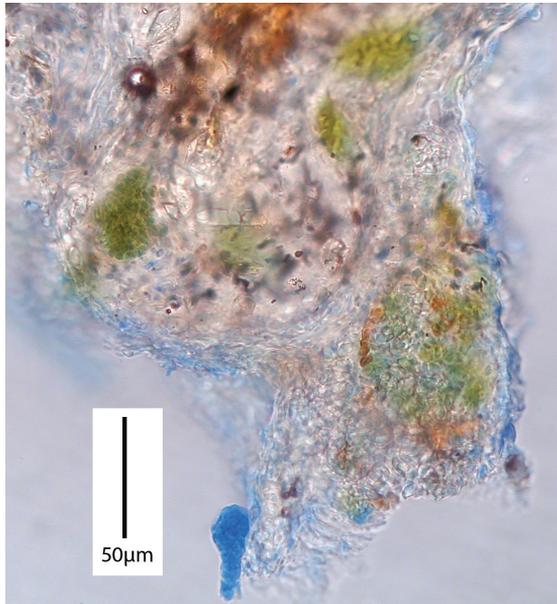


Figure 10. *Gymnopus hakaroa* PDD 96390. Pockets of algal cells embedded in hyphal tissue of stem base (cotton blue stain).

on the bark of dead logs. These two species have smooth stems. *Gymnopus ceraceicola* is distinguished by larger fruitbodies, a pruinose stipe, and is more commonly associated with southern-beech forests on dead fallen logs. The species of *Gymnopus* described here belong in the /micromphale clade of Moncalvo et al. (2002) and share the diagnostic feature of this clade of a foetid odour likely due to the presence of mercaptan-like compounds. In New Zealand this feature is shared with *Mycetinis curranae* (G. Stev.) J.A. Cooper & P. Leonard, a marasmioid fungus distinguished by its ornamented hymeniderm pileipellis. Another very distinctive character common to all three *Gymnopus* species, and visible in the accompanying photographs (Figs 2 and 6), is the presence of a waxy layer of partially gelatinised hyphae on the substrate from which the fruitbodies emerge. This layer is usually green from the presence of embedded algal cells. Interestingly, some published images of *G. foetidus* in the northern hemisphere also show a similar layer, e.g. Antonín and Noordeloos (2010). Detailed examination of our material does show algal cells deeply embedded within the context of the waxy layer and the basal portion of the stipe (Figs 9 and 10), but it would seem unlikely that algal cells are present in sufficient numbers to confer any significant nutritional benefit to the fungus. The morphologically similar *Marasmiellus affixus* (Berk.) Singer, described from Australia and commonly known as the 'little stinker', is also associated with a waxy algae-infected layer. The association of *M. affixus* with alga was noted by Singer (1973) and has been speculated to be a basidio-lichen, although this has not proven (Lepp 2011). A partial, poor quality ITS1 sequence for *M. affixus* obtained during this work (not deposited) suggests it has affinity with *Marasmiellus ramealis* (Bull.) Singer rather than the taxa treated here.

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