RESEARCH ARTICLE



Four new species of Mycena sect. Calodontes (Agaricales, Mycenaceae) from northeast China

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Abstract

Species of *Mycena* sect. *Calodontes* are representative of the *Mycena* genus as a whole and are easily recognised by the pinkish, reddish, purplish to brownish pileus and larger basidiomata. Furthermore, the colour of the pileus in the species of sect. *Calodontes* often has a transition or changes in different stages and the combination of the colour of the pileus with cystidia and basidiospores can be used to recognise taxa within this section. To date, 19 species of *Mycena* sect. *Calodontes* have been reported worldwide. Including our recent description of *M. yuezhuoi*, five species of sect. *Calodontes* have been recorded in China. During examination of specimens collected in coniferous forests or mixed broadleaf-conifer forests in temperate regions of China, additional taxa assigned to sect. *Calodontes* were identified. Four new species are recognised, based mostly on characters of the pileus and cystidia. Phylogenetic analysis of sequence data from multiple DNA regions (ITS + *rpb1* + *tef1*) supported the morphological evidence. Here, we propose *M. polycystidiata*, *M. rufobrunnea*, *M. shengshanensis* and *M. subulata* as new species in *Mycena* sect. *Calodontes*. Morphological descriptions, line drawings, habitat photos and comparisons with closely-related taxa are provided. A key to the 23 known species of sect. *Calodontes* is presented.

Keywords

coniferous forest, new taxa, phylogeny, saprobic, taxonomy

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Introduction

Mycena sect. *Calodontes* (Fr. ex Berk.) Quél. comprises the taxa in *Mycena* (Pers.) Roussel with a pinkish, reddish, purplish to brownish and mostly hygrophanous pileus, interveined lamellae, smooth cheilocystidia and pleurocystidia (if present) and mostly amyloid spores (Fries 1821; Berkeley 1836; Maas Geesteranus 1992a, 1992b; Harder et al. 2010). The group was initially proposed as *Agaricus* trib. *Clitocybe* subtrib. *Calodontes* Fries consisting of six species, then elevated to section rank within *Agaricus* subgen. *Clitocybe* Fr. ex Berk. and finally assigned to *Mycena* in 1872 (Fries 1821; Berkeley 1836; Quélet 1872). To date, 19 species are known, mostly from Europe and North America, but five species have been described from Asia (specifically, China, India and Peninsular Malaysia) (Smith 1947; Maas Geesteranus 1980; Perry 2002; Grgurinovic 2003; Robich 2003; Chew et al. 2014; Aravindakshan and Manimohan 2015; Aronsen and Læssøe 2016; Na 2019; Liu et al. 2021).

The circumscription of subsections within sect. Calodontes is problematic. Initially, Smith (1947) divided sect. Calodontes into two subsections, Granulatae and Ciliatae, according to whether the cheilocystidia were smooth or not, but this system was not widely adopted because most of the species have previously been classified in other sections of Mycena on account of the red, yellow or orange basidiomata, coloured lamellar edge and echinulate or diverticulate cheilocystidia, pleurocystidia, pileipellis or stipitipellis (Maas Geesteranus 1980, 1992a, 1992b; Perry 2002; Grgurinovic 2003; Robich 2003; Harder et al. 2010; Chew et al. 2014). Based on the lamellar edge colour, spore amyloid reaction and cystidial features, a widely accepted subsectional classification of sect. Calodontes was proposed by Maas Geesteranus (1980) and subsequent taxonomists (Grgurinovic 2003; Harder et al. 2010; Chew et al. 2014). The primary criteria for segregation were dominated by microcharacters: subsect. Purae (Konrad & Maubl.) Maas Geest. with amyloid spores and colourless pleuro- and cheilocystidia; subsect. Violacellae Sing. ex Maas Geest. with inamyloid spores and no pleurocystidia; and subsect. Marginatae J.E. Lange with amyloid spores and pleurocystidia and cheilocystidia with purplish-brown contents (Maas Geesteranus 1980; Grgurinovic 2003; Harder et al. 2010; Chew et al. 2014). Although the infrasectional classification of Maas Geesteranus (1980) is generally accepted, phylogenetic analyses have provided only weak support because subsect. Purae and subsect. Violacellae are polyphyletic (Perry 2002; Grgurinovic 2003; Robich 2003; Harder et al. 2010, 2012, 2013; Chew et al 2014; Na 2019). In studying Mycena pura (Pers.) P. Kumm., the type of subsect. Purae, Maas Geesteranus (1992a, 1992b) proposed eight forma based on pileus colour. However, 11 clades have been resolved amongst materials collected from Europe and the Americas, which suggests that there may be additional undescribed taxa in subsect. Purae (Harder et al. 2013).

Including our recent description of *M. yuezhuoi* Z.W. Liu, Y.P. Ge & Q. Na from Kunyushan National Nature Reserve (Yantai, Shandong Province), five species of *Mycena* sect. *Calodontes* have been previously recorded in China (Li et al. 2015; Na 2019; Liu et al. 2021). In this paper, we propose an additional four new species classified in

sect. *Calodontes* from the temperate zone of northeast China. The four new species share a unique set of striking morphological characters and contribute to an improved understanding of the classification of sect. *Calodontes*.

Materials and methods

Morphological observations

Thirteen fungal specimens were examined in this study, which were mainly collected in coniferous forests and some from mixed broadleaf-conifer forests in 2021. Macrocharacters were recorded from fresh specimens. Colour codes in descriptions follow those of Kornerup and Wanscher (1978). Microcharacters were observed from tissues sampled from dried specimens and rehydrated with 5% potassium hydroxide (KOH) and stained with Congo red (1% [w/v] aqueous solution), if necessary, using a Lab A1 microscope (Carl Zeiss AG, Jena, Germany). The amyloid reaction was tested with Melzer's Reagent (Clémençon et al. 2004; Horak 2005). Twenty basidiospores were measured per specimen. For the holotype, 40 basidiospores from different basidiomata were selected for measurement. Basidiospore statistics are expressed as (a/b/c) (d)e-f $g(h) \times (i)i - k - l(m) \mu m [Q = (n)o - p(q), Q = r \pm s]$, where *a*-*c* represent *a* basidiospores of b basidiomata from c specimens measured; d and h are the minimum and maximum length (5% extremum), respectively, e and g indicate the range of values for the remaining 90% of the spores and f is the average length; width (i-m) and Q values (n-q) are expressed in a similar manner; and r and s are the average Q value and its standard deviation, respectively (Ge et al. 2021; Liu et al. 2021; Na et al. 2021; Na et al. 2022). The measurement of basidia, cystidia and other characters were each based on 20 observations. All specimens have been deposited in the Fungarium of the Fujian Academy of Agricultural Sciences (FFAAS).

DNA extraction, PCR, cloning and DNA sequencing

The Plant Genomic DNA Kit (CoWin Biosciences, Beijing, China) was used to isolate total genomic DNA from dried specimens in accordance with the manufacturer's instructions. Three nuclear loci were sequenced, comprising the internal transcribed spacer (ITS), RNA polymerase II largest subunit (*rpb1*) and translation elongation factor-1 alpha (*tef1*). The primer pairs ITS1/ITS4, *rpb1*Mp_f1/*rpb1*Mp_r1 and tEF-Mp_f2/ tEFMp_r2 were selected to amplify ITS, *rpb1* and *tef1*, respectively (White et al. 1990; Harder et al. 2013; Yu et al. 2020). The PCR reactions were performed in a total volume of 25 µl containing 2 µl DNA template, 1 µl for each primer, 8.5 µl nuclease-free H₂O and 12.5 µl 2× Utaq PCR MasterMix (ZomanBio, Beijing, China). The PCR protocol for amplification of the ITS region was as follows: 94 °C for 4 min, then 34 cycles of 94 °C for 45 s, 52 °C for 45 s and 72 °C for 1 min, with a final extension of 72 °C for 10 min (Na et al. 2022). The PCR protocol for amplification of the *rpb1* and *tef1* regions followed that of Harder et al. (2013): 94 °C for 60 s, then 10 cycles of 94 °C for 35 s, 53 °C for 45 s and 72 °C for 45 s; then 25 cycles of 94 °C for 35 s, 56 °C for 45 s, 72 °C for 45 s and final extension of 72 °C for 10 min. The PCR products were purified by gel electrophoresis or filter membrane and subjected to Sanger dideoxy sequencing by the Beijing Genomics Institute (Beijing, China).

Phylogenetic analysis

A combined ITS, rpb1 and tef1 dataset was analysed to infer relationships of the new taxa with other members of sect. Calodontes. We used sequences included in previous studies of sect. *Calodontes* and from members of the most closely-related section deposited in the GenBank database, which were mainly submitted by Harder et al. (2013), Osmundson et al. (2013), Chew et al. (2014) and Liu et al. (2021). For the analysis, representative species of Mycena sect. Supinae Konrad & Maubl., which is closely related to sect. Calodontes, were selected as the outgroup (Osmundson et al. 2013; Na and Bau 2019). Sequences for each DNA region (ITS, rpb1 and tef1) were aligned in MAFFT version 7 online and the aligned matrices were manually checked with BIOEDIT 7.2.5.0 (Hall 1999; Kuraku et al. 2013; Katoh et al. 2019). The best-fit substitution model for each gene partition was determined with MODELTEST 2.3, based on the Akaike Information Criterion (Posada and Crandall 1998). Maximum Likelihood (ML) analysis was conducted by raxmlGUI 2.09 (Edler et al. 2020). The phylogenetic analysis was performed by a single analysis with six partitions (ITS1, 5.8S, ITS2, rpb1 exons, tef1 exons, intron of rpb1 + introns of tef1), using the GTR-GAMMA model and 1,000 rapid bootstrap (BS) replicates. For Bayesian Inference (BI), two runs of six chains were run for 15,000,000 generations and sampled every 10,000 generations by MrBayes 3.2.6. At the end of the run, the average deviation of split frequencies was 0.007821, ESS (effective sample size) was 1300.3 and the average Potential Scale Reduction Factor (PSRF) parameter values (excluding NA and > 10.0) = 1.000 and the "sump" and "sumt" commands were used to summarise sampled parameters with 25% burn-in (Ronquist and Huelsenbeck 2003).

Results

Phylogenetic relationships

The dataset consisted of 192 sequences, comprising 39 newly-generated sequences (13 ITS, 13 *rpb1* and 13 *tef1*) and 153 sequences (61 ITS, 46 *rpb1* and 46 *tef1*) down-loaded from GenBank. In total, 74 accessions of 19 species were included in the dataset. Detailed information for all sequences is presented in Table 1. The aligned dataset contained 1459 nucleotide sites including gaps (229 sites for ITS1, 159 sites for 5.8S, 177 sites for ITS2, 55 sites for *rpb1* exons, 295 sites for *tef1* exons, 544 sites for intron of *rpb1* + introns of *tef1*), of which 1146 were conserved, 257 were parsimony-

No.	Species	Specimen	GenBar	nk accession 1	numbers	Locality	Reference
		voucher	ITS	rpb1	tef1	-	
1.	Mycena aff. pura	TL8052	FN394623	KF723687	KF723641	Ecuador	Harder et al. (2010, 2013)
2.	M. aff. pura	TL9433	FN394622	KF723688	KF723642	Ecuador	Harder et al. (2010, 2013)
3.	M. aff. pura	TL9450	KJ144653	KF723689	KF723643	Ecuador	Harder et al. (2010, 2013)
4.	M. aff. pura	TL9678	FN394621	KF723690	KF723644	Ecuador	Harder et al. (2010, 2013)
5.	M. arcangeliana	252b	JF908401	_	_	Spain	Osmundson et al. (2013)
6.	M. arcangeliana	252f	JF908402	_	_	Spain	Osmundson et al. (2013)
7.	M. cahaya	ACL134	KF537248	-	-	Malaysia	Chew et al. (2014)
8.	<i>M.</i> cf. <i>pura</i> I	CBH039	FN394588	KF723680	KF723634	Denmark	Harder et al. (2010, 2013)
9.	<i>M.</i> cf. <i>pura</i> II	CBH105	FN394581	KF723671	KF723625	Denmark	Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> II	CBH169	FN394579	KF723672	KF723626	Denmark	Harder et al. (2010, 2013)
11.	M. cf. pura II	CBH366	FN394572	KF723673	KF723627	Denmark	Harder et al. (2010, 2013)
	M. cf. pura II	CBH404	FN394566	KF723674	KF723628	Denmark	Harder et al. (2010, 2013)
	M. cf. pura III	CBH019	FN394605	KF723675	KF723629	Denmark	Harder et al. (2010, 2013)
	<i>M</i> . cf. <i>pura</i> III	CBH022	FN394574	KF723676	KF723630	Denmark	Harder et al. (2010, 2013)
	<i>M</i> . cf. <i>pura</i> III	KK	FN394606	KF723677	KF723631	Slovakia	Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> IV	CBH410	FN394595	KF723667	KF723621	Denmark	Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> IV	JV06979	FN394585	KF723668	KF723622		Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> IV	TL1279(FN394583	KF723669	KF723623	Denmark	Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> IV	TL12786 CBH226	FN394591	KF723670 KF723664	KF723624 KF723618	Sweden	Harder et al. (2010, 2013) Harder et al. (2010,
	M. cf. pura V		FN394604				2013)
	<i>M</i> . cf. <i>pura</i> V	TL5614	FN394602				2013)
	<i>M.</i> cf. <i>pura</i> VI	BAP132	FN394561	KF723660	KF723614	USA	Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> VIII	CBH216	FN394598	KF723662	KF723616		Harder et al. (2010, 2013)
	<i>M.</i> cf. <i>pura</i> VIII	CBH402	FN394599	KF723663	KF723617	Denmark	Harder et al. (2010, 2013)
25.	M. cf. pura IX	CBH166	FN394607	KF723701	KF723655	Denmark	Harder et al. (2010, 2013)

 Table 1. Specimens used in phylogenetic analysis and GenBank accession numbers.

No.	Species	Specimen	GenBar	nk accession r	umbers	Locality	Reference
		voucher	ITS	rpb1	tef1		
26.	M. cf. pura IX	CBH358	FN394608	KF723702	KF723656	Denmark	Harder et al. (2010, 2013)
27.	<i>M.</i> cf. <i>pura</i> IX	CBH367	KF913022	KF723703	KF723657	Denmark	Harder et al. (2013)
	M. cf. pura IX	CBH371	KF913023	KF723704	KF723658	Denmark	Harder et al. (2013)
	M. cf. pura X	BAP165A	FN394563	KF723698	KF723652	USA	Harder et al. (2010, 2013)
30.	<i>M.</i> cf. <i>pura</i> XI	CBH187	FN394564	KF723678	KF723632	Sweden	Harder et al. (2010, 2013)
31.	<i>M.</i> cf. <i>pura</i> XI	CBH386	FN394565	KF723679	KF723633	Denmark	Harder et al. (2010, 2013)
32.	M. diosma	CBH400	FN394617	KF723699	KF723653	Denmark	Harder et al. (2010, 2013)
33.	M. diosma	LK1191/2000	FN394619	KF723700	KF723654	Germany	Harder et al. (2010, 2013)
34.	M. dura	10315	FN394560	KF723694	KF723648	Austria	Harder et al. (2010, 2013)
35.	M. lammiensis	TUR165927	FN394552	KF723697	KF723651	Finland	Harder et al. (2010, 2013)
36.	M. meliigena	39	JF908423	-	_	Italy	Osmundson et al. (2013)
37.	M. meliigena	39d	JF908429	-	_	Italy	Osmundson et al. (2013)
38.	M. pearsoniana	CBH068	FN394614	KF723691	KF723645	Germany	Harder et al. (2010, 2013)
39.	M. pearsoniana	JV06890	FN394612	KF723692	KF723646	Denmark	Harder et al. (2010, 2013)
40.	M. pearsoniana	LK880/2002	FN394613	KF723693	KF723647	Germany	Harder et al. (2010, 2013)
41.	M. pelianthina	CBH015	FN394549	KF723695	KF723649	Denmark	Harder et al. (2010, 2013)
42.	M. pelianthina	CBH016	FN394547	KF723696	KF723650	Denmark	Harder et al. (2010, 2013)
43.	M. polycystidiata	FFAAS0417 Holotype	ON427731	ON468456	ON468469	China	This study
44.	M. polycystidiata	FFAAS0418	ON427732	ON468457	ON468470	China	This study
	M. polycystidiata	FFAAS0421	ON427733	ON468458	ON468471	China	This study
46.	M. polycystidiata	FFAAS0422	ON427734	ON468459	ON468472	China	This study
47.	M. pseudocorticola	124a	JF908386	_	_	Italy	Osmundson et al. (2013)
48.	M. pura	IS10/11/2000	FN394611	-	-	USA	Harder et al. (2010)
49.	M. pura f. lutea	DB2005/152	FN394603	-	-	Denmark	Harder et al. (2010)
	M. rosea	UP2	FN394550	-	-	UK	Harder et al. (2010)
51.	M. rosea	CBH097	FN394556	KF723681	KF723635	Denmark	Harder et al. (2010, 2013)
52.	M. rosea	CBH383	FN394553	KF723682	KF723636	Denmark	Harder et al. (2010, 2013)
53.	M. rosea	CBH409	FN394551	KF723683	KF723637	Germany	Harder et al. (2010, 2013)
54.	M. rosea	TL12393	FN394555	KF723684	KF723638	Denmark	Harder et al. (2010, 2013)

No. Species		Specimen	GenBank accession numbers			Locality	Reference
		voucher	ITS	rpb1	tef1		
55.	M. rosea	TL12409	FN394557	KF723685	KF723639	Denmark	Harder et al. (2010, 2013)
56.	M. rufobrunnea	FFAAS0414	ON427728	ON468453	ON468466	China	This study
57.	M. rufobrunnea	FFAAS0415	ON427729	ON468454	ON468467	China	This study
58.	M. rufobrunnea	FFAAS0416	ON427730	ON468455	ON468468	China	This study
		Holotype					
59.	M. seminau	ACL136	KF537250	-	-	Malaysia	Chew et al. (2014)
60.	M. seminau	ACL308	KF537252	-	-	Malaysia	Chew et al. (2014)
61.	M. shengshanensis	FFAAS0424	ON427739	ON468464	ON468477	China	This study
		Holotype					
62.	M. shengshanensis	FFAAS0425	ON427740	ON468465	ON468478	China	This study
63.	M. sinar	ACL092	KF537247	-	-	Malaysia	Chew et al. (2014)
64.	M. sinar	ACL135	KF537249	-	-	Malaysia	Chew et al. (2014)
65.	M. sinar var.	ACL307	KF537251	-	-	Malaysia	Chew et al. (2014)
	tangkaisinar						
66.	M. subulata	FFAAS0419	ON427735	ON468460	ON468473	China	This study
67.	M. subulata	FFAAS0420	ON427736	ON468461	ON468474	China	This study
68.	M. subulata	FFAAS0423	ON427737	ON468462	ON468475	China	This study
		Holotype					
69.	M. subulata	FFAAS0426	ON427738	ON468463	ON468476	China	This study
70.	M. supina	128a	JF908388	-	-	Italy	Osmundson et al.
							(2013)
71.	M. yuezhuoi	FFAAS0344	MW581490	MW868166	MW882249	China	Liu et al. (2021)
72.	M. yuezhuoi	FFAAS0345	MW581491	MW868169	MW882250	China	Liu et al. (2021)
73.	M. yuezhuoi	FFAAS0346	MW581492	MW868168	MW882251	China	Liu et al. (2021)
74.	M. yuezhuoi	FFAAS0347	MW581493	MW868167	MW882252	China	Liu et al. (2021)

informative and 56 were variable, but parsimony-uninformative. For Bayesian Inference (BI), the selected models for each DNA region of the concatenated dataset were as follows: HKY+G for ITS1 and intron of *rpb1* + introns of *tef1*, JC for 5.8S and *rpb1* exons, HKY+I+G for ITS2 and SYM+I+G for *tef1* exons. The BI and ML analyses resulted in almost identical topologies; thus, the BI topology is presented as a master tree (Fig. 1).

The phylogenetic analysis revealed that sect. *Calodontes* was strong support (BS/ Bayesian posterior probability [BPP] = 100/1.00) (Fig. 1). Fifteen species and eleven *M. pura* complex clades within sect. *Calodontes* were retrieved. Four new species were resolved as monophyletic, each with strong support: *M. polycystidiata* (BS/ BPP = 100/1.00), *M. rufobrunnea* (BS/BPP = 100/1.00), *M. shengshanensis* (BS/ BPP = 90/1.00) and *M. subulata* (BS/BPP = 100/1.00). A sister relationship between *M. shengshanensis* and *M. pearsoniana* Dennis ex Singer was well supported. *Mycena subulata* was resolved as sister, but genetically distant from *M. pearsoniana* and *M. shengshanensis* clade. In addition, the sister relationship of *Mycena polycystidiata* and *M. rufobrunnea* were unresolved.

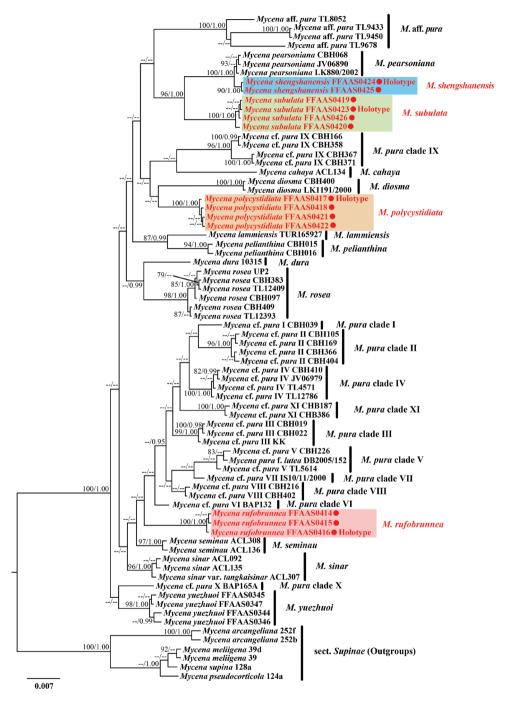


Figure 1. Bayesian Inference analysis of *Mycena* sect. *Calodontes* with ITS, *rpb1* and *tef1* sequence data. Species in *Mycena* sect. *Supinae* served as outgroup. Bootstrap values (BS) from Maximum Likelihood \geq 75 and Bayesian posterior probabilities (BPP) \geq 0.95 are shown on each branch (BS/BPP). The new species are marked in red.

Taxonomy

Mycena polycystidiata Z.W. Liu, Y.P. Ge, L. Zou & Q. Na, sp. nov.

MycoBank No: 843977 Figs 2–5

Diagnosis. Pileus greyish-rose, umbo brownish-orange, hygrophanous. Stipe pubescent. Pleurocystidia polymorphic in shape. Stipitipellis a cutis, with numerous projecting hyphae.

Holotype. CHINA. Heilongjiang Province: Liangshui National Nature Reserve, Yichun City, 47°12'74"N, 128°52'86"E, 20 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0417* (collection number MY0633).

Etymology. Refers to the variable shape of pleurocystidia.

Description. *Pileus* 14–31 mm in diam., campanulate to hemispherical when young, plano-convex with age, with obtuse umbo at centre, margin slightly revolute, at times cracked at mature; umbo brownish-orange (7C3–7C5), disc purplish-grey (13C2, 14B2, 14C2), reddish-grey (12B2, 12C2) to greyish-rose (12B3), near margin reddish-grey (12D2), greyish-ruby (12D3) or purplish-grey (13D3), margin whitish; striate none or indistinct, greyish-ruby (12E3–12E4), towards the centre up to 1/3 diam.; surface dry and rugose, hygrophanous, generally tomentose. *Context* white, 2 mm thick, fragile. *Lamellae* emarginate, slightly decurrent when old, 20–28 reaching the stipe, 1–2 tiers of lamellulae, white, irregularly intervenose, edge concolorous, wavy. *Stipe* 34–73 × 3–7 mm, central, cylindrical, base occasionally compressed with age; apex violet brown (11E3–11E4, 11F4), greyish-ruby (12E4), lower part brownish-grey (11D2, 11E2) to greyish-brown (11D3, 11E3) or purplish-grey (13C2), fragile, hollow; apex to middle densely pubescent, sparser towards base; whitely villose at base. *Odour* strongly raphanoid, *taste* indistinct.

Basidiospores (130/5/4) (6.4)6.7–7.4–8.3(8.8) × (3.2)3.5–3.9–4.3(4.6) µm $[Q = (1.62)1.72 - 2.05(2.18), Q = 1.90 \pm 0.11]$ [holotype (70/2/1) (6.7)6.9-7.6-8.5(8.7) \times (3.4)3.6–4.0–4.4(4.6) µm, Q = (1.71)1.76–2.05(2.13), Q = 1.90 ± 0.09], elongated ellipsoid to cylindrical, colourless, smooth, thin-walled, amyloid. Basidia 21-31 × 6-8 µm, 4-spored, clavate, hyaline, sterigmata approximately 4 µm in length. Cheilocys*tidia* thin-walled, hyaline, differs in two shapes, mainly utriform, $50-65 \times 20-31 \mu m$, some subclavate, 54-78 × 14-19 µm. *Pleurocystidia* abundant, thin-walled, hyaline, multi-shaped: lanceolate and mostly round to blunt apices, $37-81 \times 12-20 \mu m$, lanceolate and acute apices, $51-87 \times 14-22 \mu m$, elliptical, $30-86 \times 12-31 \mu m$, ovate and acute apices, $49-71 \times 15-24 \mu m$, ovate and mostly round to blunt apices, $49-73 \times 16-22 \mu m$. *Pileipellis* a cutis composed of four to five layers cylindrical cells, $51-81 \times 4-5 \mu m$, smooth and thin-walled; terminal cells cylindrical or fusiform, $50-69 \times 3-22 \mu m$, thinwalled, hyaline. Hypodermium formed by fusiform to subglobose hyphae, 32-69 × 18–54 μm, thin-walled, hyaline. *Lamellar trama* subregular, dextrinoid. *Stipitipellis* a cutis composed of cylindrical hyphae $5-8 \mu m$ in diam., smooth, thin-walled, with numbers of projecting hyphae 2–6 μ m in diam.; *caulocystidia* 29–74 × 6–19 μ m, clavate or fusiform, thin-walled, smooth, hyaline. *Clamps* present in all tissues.



Figure 2. Basidiomata of *Mycena polycystidiata* Z.W. Liu, Y.P. Ge, L. Zou & Q. Na **a, b** *FFAAS0422* **c, d** *FFAAS0417*, holotype **e-g** *FFAAS0421* **h** *FFAAS0418* Scale bars: 10 mm (**a-h**). Photographs **a-e, h** by Qin Na **f, g** by Yupeng Ge.

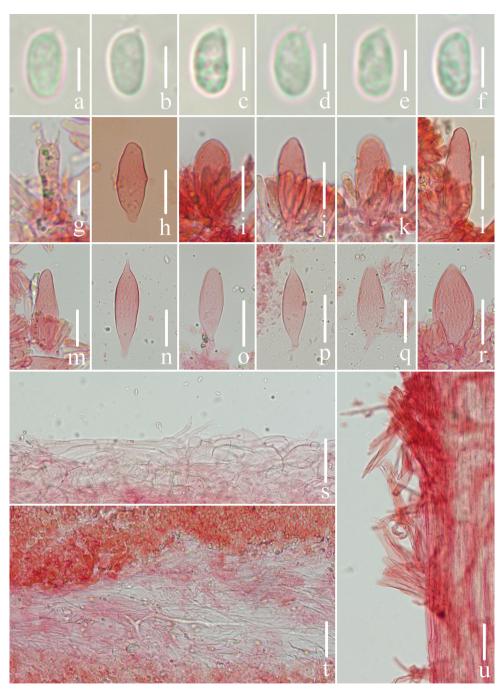


Figure 3. Microscopic features of *Mycena polycystidiata* (*FFAAS0417*, holotype) **a–f** basidiospores **g** basidia **h–l** cheilocystidia **m–r** pleurocystidia **s** pileipellis and hypodermium **t** lamellar trama **u** stipitipellis and caulocystidia. Scale bars: 5 μm (**a–f**); 10 μm (**g**); 30 μm (**h–r**); 40 μm (**s–u**).

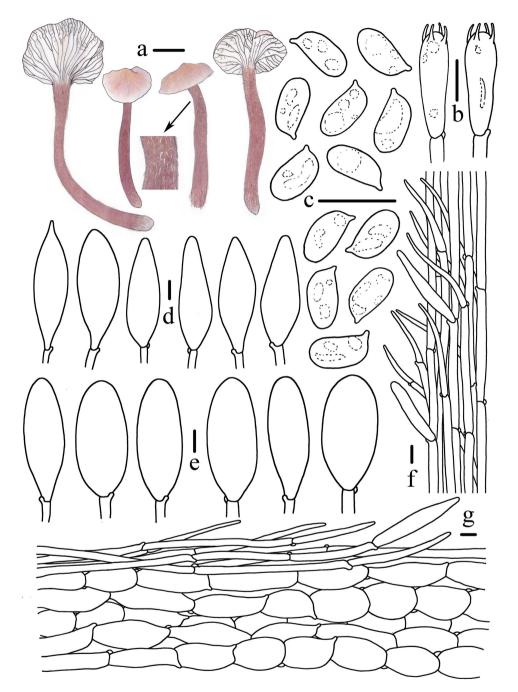


Figure 4. Morphological features of *Mycena polycystidiata* (*FFAAS0417*, holotype) **a** basidiomata **b** basidia **c** basidiospores **d** pleurocystidia **e** cheilocystidia **f** stipitipellis and caulocystidia **g** pileipellis and hypodermium. Scale bars: 10 mm (**a**); 10 μm (**b–g**). Drawings by Zewei Liu.

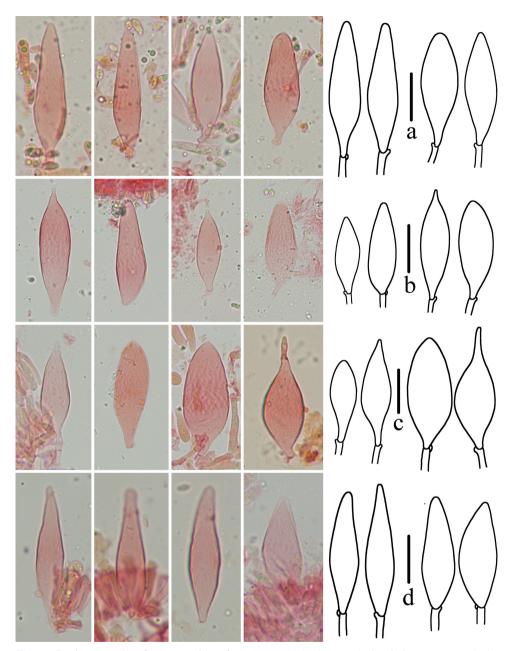


Figure 5. Pleurocystidia of *Mycena polycystidiata* **a** *FFAAS0422* **b** *FFAAS0417*, holotype **c** *FFAAS0418* **d** *FFAAS0421*. Scale bars: 25 μm (**a–d**).

Habitat. Scattered on the litter layers in *Pinus koraiensis* and *Larix gmelinii* mixed forests.

Known distribution. Heilongjiang Province, China.

Additional material examined. CHINA. Heilongjiang Province: Liangshui National Nature Reserve, Yichun City, 47°12'82"N, 128°52'94"E, 20 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0418* (collection number MY0634); same location, 21 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0421* (collection number MY0659); same location, 21 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0422* (collection number MY0661).

Notes. Macroscopically, *Mycena luteovariegata* Harder & Læssøe and *M. pura* resemble *M. polycystidiata* in pileus colour, but the latter possesses more typically utriform cheilocystidia and uncontracted pleuro- and cheilocystidia (Perry 2002; Robich 2003; Harder et al. 2013; Aronsen and Læssøe 2016; Na 2019). *Mycena pearsoniana* also has a rose to violaceous pileus, but differs from *M. polycystidiata* in having inamyloid spores and lacking pleurocystidia (Aronsen and Læssøe 2016; Na 2019). Compared with *M. polycystidiata*, *M. sirayuktha* Aravind. & Manim. has similar cheilocystidia, but has an obviously greyish-brown striate pileus, inamyloid spores and slightly glutinous pileipellis with finger-like excrescences (Aravindakshan and Manimohan 2015).

The pleurocystidia of *M. polycystidiata* varied in shape amongst specimens (Fig. 5). In all four specimens, most pleurocystidia were lanceolate and with round to blunt apices, but pleurocystidia with lanceolate and acute apices, elliptical and ovate and acute apices were also observed in *FFAAS0417* (Holotype) and *FFAAS0418*, while elongated lageniform-lanceolate or round apices ovate were detected in *FFAAS0421* and *FFAAS0422*. The multi-shaped pleurocystidia may show a morphological continuum that changes between developmental stages. Nevertheless, the multi-shaped pleurocystidia are unquestionably diagnostic for identification of this species.

Mycena rufobrunnea Z.W. Liu, Y.P. Ge & Q. Na, sp. nov.

MycoBank No: 843978 Figs 6–8

Diagnosis. Pileus dark brown at centre, disc gradually turning paler to reddish-brown to greyish-brown, edge white. Lamellae obviously intervenose. Stipe apex to middle greyish-magenta to dull violet, lower part darker to dark purple or dark magenta. Cheilocystidia utriform, sometimes clavate. Pleurocystidia absent. Caulocystidia clavate or fusiform. Pileipellis with fusiform terminal cells.

Holotype. CHINA. Jilin Province: Dayangcha, Erdaobaihe Town, Antu County, Yanbian Korean Autonomous Prefecture, 42°20'73"N, 127°56'06"E, 16 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0416* (collection number MY0581).

Etymology. Refers to reddish-brown pileus.

Description. *Pileus* 12–34 mm in diam., hemispherical to convex when young, then plano-convex, sometimes an unclear umbo at centre, margin slightly revolute, acute to subacute, at times cracked at mature; dark brown (8F6–8F8) at centre, disc gradually turning paler to reddish-brown (8D4–8D5, 8E6–8E8) to greyish-brown



Figure 6. Basidiomata of *Mycena rufobrunnea* Z.W. Liu, Y.P. Ge & Q. Na **a,b** *FFAAS0414* **c,d** *FFAAS0415* **e-h** *FFAAS0416*, holotype. Scale bars: 10 mm (**a-h**). Photographs **a-h** by Qin Na.

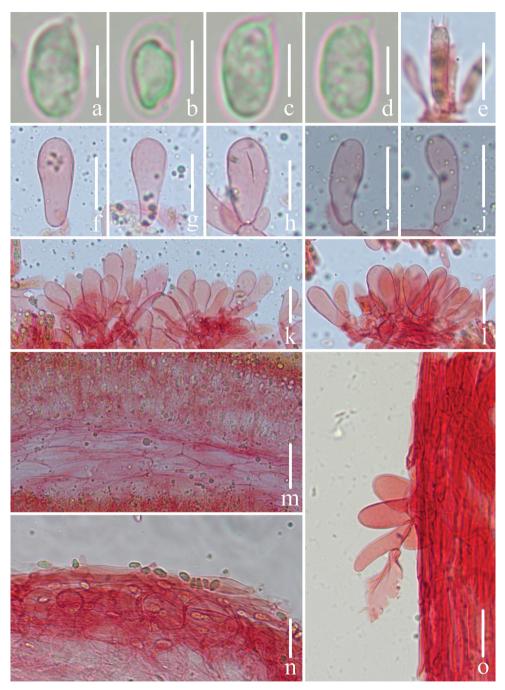


Figure 7. Microscopic features of *Mycena rufobrunnea* (*FFAAS0416*, holotype) **a–d** basidiospores **e** basidia **f–l** cheilocystidia **m** lamellar trama **n** pileipellis and *hypodermium* **o** stipitipellis and caulocystidia. Scale bars: 5 μm (**a–d**); 20 μm (**e–o**).

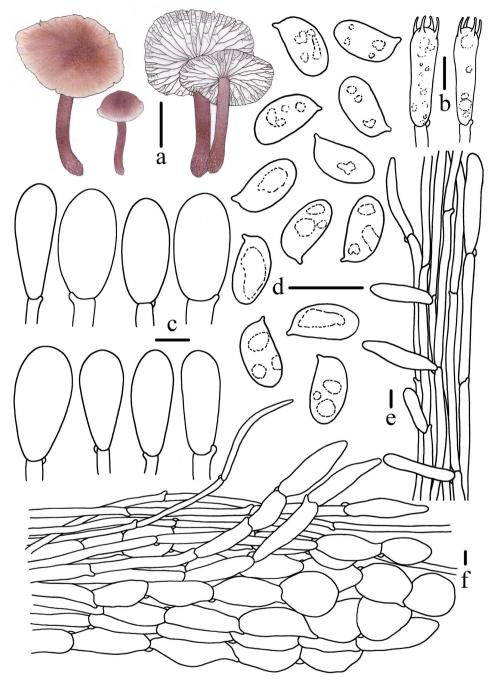


Figure 8. Morphological features of *Mycena rufobrunnea (FFAAS0416*, holotype) **a** basidiomata **b** basidia **c** cheilocystidia **d** basidiospores **e** stipitipellis and caulocystidia **f** pileipellis and hypodermium. Scale bars: 10 mm (**a**); 10 μm (**b–f**). Drawings by Zewei Liu.

(8D3) and turning to whitish at margin; striate reddish-brown (8D4–8D5, 8E6–8E8), towards the centre up to 1/3-1/2 diam.; surface humidus when wet. **Context** white, 1.5 mm thick, fragile. **Lamellae** adnexed to emarginate, 20–23 reaching the stipe, 1–3 tiers of lamellulae, white, irregularly intervenose, edge concolorous, slightly serrulate. **Stipe** 19–62 × 2–6 mm, central, cylindrical, apex to middle greyish-magenta (14E4–14E5) to dull violet (16E3–16E4), lower part darker to dark purple (14F4–14F5) or dark magenta (13F3), fragile, hollow, base slightly swollen with whitish villose. **Odour** raphanoid, **taste** indistinct.

Basidiospores (80/4/3)(7.1)7.6 - 8.4 - 9.2(9.6)× (3.8)4.0-4.5-5.0 μm $[Q = (1.73)1.77 - 1.98(2.05), Q = 1.88 \pm 0.07]$ [holotype (40/2/1) (7.9)8.1-8.6- $9.2(9.4) \times 4.2-4.6-5.0 \ \mu m, \ Q = (1.73)1.77-1.96(1.98), \ Q = 1.87 \pm 0.06], \ elon$ gated ellipsoid to cylindrical, colourless, smooth, thin-walled, amyloid. Basidia 24-34 × 7-10 µm, 4-spored, clavate, hyaline, sterigmata 2-3 µm in length. Cheilocyst*idia* thin-walled, hyaline, utriform, sometimes clavate, $23-44 \times 7-17 \mu m$, abundant. Pleurocystidia absent. Pileipellis a cutis composed of four to five slightly interwoven layers of cylindrical cells, $44-70 \times 4-7 \mu m$, smooth, thin-walled; terminal cells cylindrical or fusiform, $34-65 \times 4-17 \mu m$, thin-walled, hyaline. *Hypodermium* formed by fusiform, subcylindrical to subglobose hyphae, $15-50 \times 12-37 \mu m$, thin-walled, hyaline. Lamellar trama subregular, dextrinoid. Stipitipellis a cutis composed of hyphae 3-9 µm in diam., smooth, thin-walled; *caulocystidia* common in the apex, sparse in the middle and base, $23-76 \times 6-14 \mu m$, clavate and fusiform, thin-walled, hyaline, smooth. *Clamps* present in all tissues.

Habitat. Scattered on the decayed logs of *Acer, Larix, Pinus, Populus, Quercus* and *Ulmus* mixed forests.

Known distribution. Jilin Province, China.

Additional material examined. CHINA. Jilin Province: Dayangcha, Erdaobaihe Town, Antu County, Yanbian Korean Autonomous Prefecture, 42°20'72"N, 127°56'08"E, 16 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0414* (collection number MY0579); same location, 16 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0415* (collection number MY0580).

Notes. Species of sect. *Calodontes* that are macroscopically similar to *Mycena rufobrunnea* have been recorded in many regions of the world. Most taxa resemble *M. rufobrunnea* in pileus colour (Smith 1947; Maas Geesteranus 1992a, 1992b; Gr-gurinovic 2003; Robich 2003; Chew et al. 2014; Aronsen and Læssøe 2016). *Mycena dura* Maas Geest. & Hauskn., recorded in Europe, also has a dark brown to greyishbrown pileus, but can be distinguished from *M. rufobrunnea* in having a white stipe and having pleurocystidia (Robich 2003; Aronsen and Læssøe 2016). *Mycena kuehne-riana* A.H. Sm., which is recorded from the United States and Canada, is distinguished from *M. rufobrunnea* in that its pileus is pale avellaneous with rose and lilac, almost white when faded and the spores are obviously smaller ($5-6 \times 2-3 \mu m$) (Smith 1947; Maas Geesteranus 1992a, 1992b). *Mycena clarkeana* Grgur. and *M. nullawarrensis* Gr-gur., described from Australia, are similar to *M. rufobrunnea* in having a reddish-brown pileus, but both species have broader spores and possess pleurocystidia (Grgurinovic 2003). Mycena cahaya A.L.C. Chew & Desjardin, M. seminau A.L.C. Chew & Desjardin and M. sinar A.L.C. Chew & Desjardin, known from Malaysia, resemble M. rufobrunnea owing to the brown pileus, but differ in having adnate to subdecurrent lamellae, a yellowish-grey or brownish-orange stem, mucronate cheilocystidia and lack caulocystidia (Chew et al. 2014). Microscopically, utriform or clavate cheilocystidia and absence of pleurocystidia are key characteristics of M. rufobrunnea. Mycena diosma Krieglst. & Schwöbel has similar cheilocystidia and pleurocystidia are absent or rare, but it has a strongly hygrophanous pileus and a remarkable change in colour (Robich 2003; Aronsen and Læssøe 2016). Mycena pura, M. sirayuktha and M. vinacea Cleland have similar cheilocystidia, but are easily distinguished from M. rufobrunnea by the presence of pleurocystidia (Perry 2002; Grgurinovic 2003; Robich 2003; Aravindak-

Mycena shengshanensis Z.W. Liu, Y.P. Ge & Q. Na, sp. nov.

shan and Manimohan 2015; Aronsen and Læssøe 2016; Na 2019).

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Diagnosis. Pileus brown to violet-brown at centre, disc light brown to dull red. Cheilocystidia clavate with slightly inflated apex, thick-walled. Pleurocystidia absent. Caulocystidia clavate with tapered apices, apex to middle thick-walled. Scattered to gregarious under *Larix gmelinii*.

Holotype. CHINA. Heilongjiang Province: Shengshan National Nature Reserve, Heihe City, 49°37'45"N, 126°47'39"E, 23 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0424* (collection number MY0686).

Etymology. Refers to the type locality.

Description. Pileus 13-26 mm in diam., when young parabolic to convex, with obtuse umbo at centre, then plano-convex, margin wavy and revolute, at times cracked at mature; centre light brown (7D5–7D6), brown (7E4–7E8), dark brown (8F5–8F6), violet brown (11F4-11F6), disc paler to light brown (7D4-7D5), brown (7E5-7E6), greyish-brown (8D3), reddish-brown (8D4, 8E4), brownish-grey (11C2), dull red (11C3), margin whitish; striate indistinct, brownish-orange (7C3), greyish-brown (7D3), reddish-grey (12D2), greyish-ruby (12E3-12E5), towards the centre up to 1/3–1/2 diam.; surface slightly moist, smooth. *Context* white, 1–2 mm thick, fragile. *Lamellae* sinuate to subdecurrent, 19–25 reaching the stipe, 1–3 tiers of lamellulae, white, irregularly intervenose, edge concolorous, wavy and slightly serrulate. Stipe 26-42 × 2-4 mm, central, cylindrical, apex reddish-brown (8E4-8E5), greyish-ruby (12E4–12E5), grevish-brown (11F3), violet brown (11F4–11F5), dark ruby (12F4– 12F5), lower part paler to brownish-grey (7C2), brownish-orange (7C3), greyishbrown (11E3), greyish-ruby (12E3), base darker to brown (7E5), reddish-brown (8E4–8E5), grevish-brown (11F3), violet brown (11F4–11F5), grevish-ruby (12E3), dark ruby (12F6–12F8), fragile, hollow, base swollen with white fibrils. **Odour** raphanoid, taste indistinct.



Figure 9. Basidiomata of *Mycena shengshanensis* Z.W. Liu, Y.P. Ge & Q. Na **a–d** *FFAAS0424*, holotype **e–h** *FFAAS0425* Scale bars: 10 mm (**a–h**). Photographs **a–c, e–g** by Yupeng Ge **d, h** by Zewei Liu.

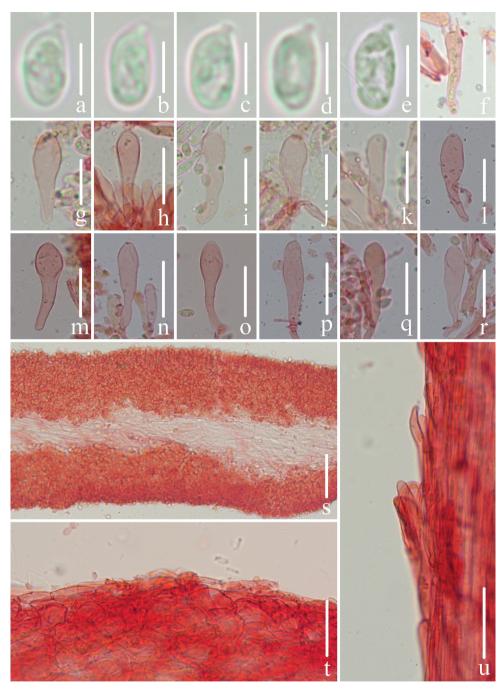


Figure 10. Microscopic features of *Mycena shengshanensis* (*FFAAS0424*, holotype) **a–e** basidiospores **f** basidia **g–r** cheilocystidia **s** lamellar trama **t** pileipellis and hypodermium **u** stipitipellis and caulocystidia. Scale bars: 5 μm (**a–e**); 20 μm (**g–r**); 40 μm (**s–u**).

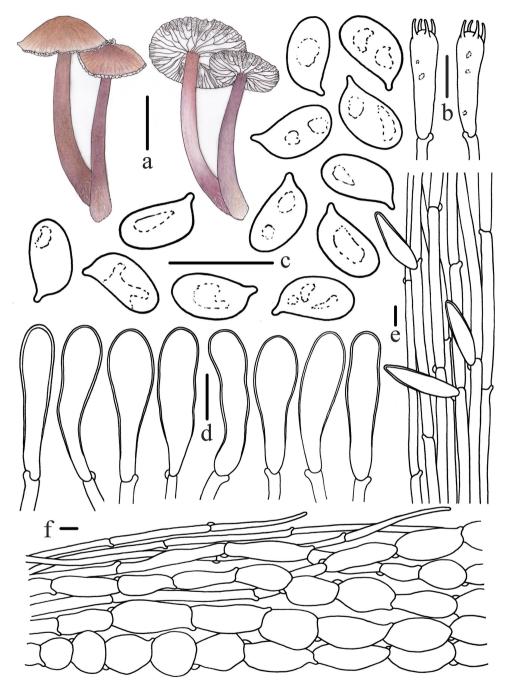


Figure 11. Morphological features of *Mycena shengshanensis* (*FFAAS0424*, holotype) **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** stipitipellis and caulocystidia **f** pileipellis and hypodermium. Scale bars: 10 mm (**a**); 10 μm (**b–f**). Drawings by Zewei Liu.

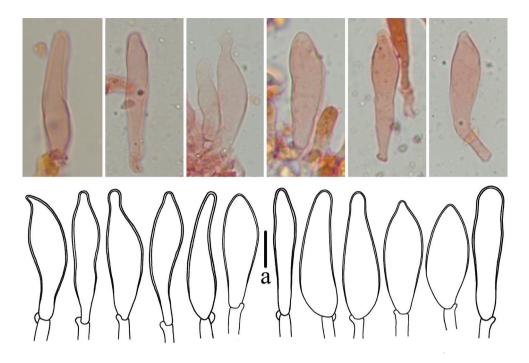


Figure 12. Cheilocystidia of Mycena shengshanensis FFAAS0424, holotype. Scale bars: 10 µm (a).

Basidiospores (60/3/2) (5.9)6.1–6.9–8.1(8.7) × 3.4–4.0–4.4(4.7) µm [Q = 1.62– 1.93 (1.97), Q = 1.75 \pm 0.09] [holotype (40/2/1) 6.1–7.0–8.2(8.7) × 3.4–4.0– 4.4(4.7) µm, Q = (1.62)1.65–1.93(1.97), Q = 1.77 \pm 0.08], elongated ellipsoid, colourless, smooth, thin-walled, amyloid. **Basidia** 22–32 × 6–8 µm, 4-spored, clavate, hyaline, sterigmata 3–4 µm in length. **Cheilocystidia** moderately thickwalled (0.5–0.6 µm), clavate with slightly inflated apex, 25–63 × 6–12 µm, hyaline. **Pleurocystidia** absent. **Pileipellis** a cutis composed of three to four layers of cylindrical cells, 24–57 × 3–5 µm, smooth and thin-walled; terminal cells cylindrical, apically narrow, 28–49 µm in length, apex 1–3 µm and base 2–5 µm in diam., thin-walled, hyaline. **Hypodermium** formed by fusiform to subglobose hyphae, 19–53 × 13–30 µm, thin-walled, hyaline. **Lamellar trama** subregular, weakly dextrinoid to dextrinoid. **Stipitipellis** a cutis composed of hyphae 4–9 µm in diam., smooth, thin-walled; **caulocystidia** 22–61 × 5–20 µm, clavate with tapered apices, apex to middle thickwalled, smooth, hyaline. **Clamps** present in all tissues.

Habitat. Scattered to gregarious on the litter layer in Larix gmelinii.

Known distribution. Heilongjiang Province, China.

Additional material examined. CHINA. Heilongjiang Province: Shengshan National Nature Reserve, Heihe City, 23 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0425* (collection number MY0687).

Notes. In sect. Calodontes, M. diosma, M. pearsoniana and M. yuezhuoi also have clavate cheilocystidia with a slightly inflated apex and lack pleurocystidia, similar to M. shengshanensis, but M. diosma differs in pileus characters, M. pearsoniana has inamyloid spores and *M. yuezhuoi* has a more purple pileus and subcellular lamellar trama (Robich 2003; Aronsen and Læssøe 2016; Na 2019; Liu et al. 2021). Clavate cheilocystidia are also present in *M. luteovariegata* and *M. pura*, but these species differ in having pleurocystidia (Perry 2002; Robich 2003; Harder et al. 2013; Aronsen and Læssøe 2016; Na 2019). Macroscopically, several species in sect. *Calodontes* also have a brown with reddish or violet pileus or stipe (Smith 1947; Maas Geesteranus 1992a, 1992b; Grgurinovic 2003; Robich 2003; Aronsen and Læssøe 2016). Mycena dura, M. kuehneriana and M. nullawarrensis are distinguished by basidiospore size (Smith 1947; Maas Geesteranus 1992a, 1992b; Grgurinovic 2003; Robich 2003; Aronsen and Læssøe 2016). Two species, M. seminau and M. sirayuktha, reported from Southeast Asia, are similar to *M. shengshanensis* owing to the brown pileus, but they differ in having gelatinised or sometimes mucronate cheilocystidia and caulocystidia have not been observed (Chew et al. 2014; Aravindakshan and Manimohan 2015). Fusiform, obclavate, ovate and clavate cheilocystidia with a subcapitate protuberance were observed occasionally, but clavate cheilocystidia with a slightly inflated apex represented the predominant morphological type in *M. shengshanensis* (Fig. 12).

Mycena subulata Z.W. Liu, Y.P. Ge & Q. Na, sp. nov.

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Diagnosis. Pileus reddish-grey to dull red, slightly hygrophanous. Cheilocystidia thick-walled, slenderly fusiform with distinctly long and narrow protuberance. Stipitipellis a cutis, with projecting hyphae, caulocystidia thick-walled.

Holotype. CHINA. Heilongjiang Province: Liangshui National Nature Reserve, Yichun City, 47°13'13"N, 128°53'21"E, 21 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0423* (collection number MY0671).

Etymology. Refers to cheilocystidia with distinctly long and narrow protuberance.

Description. *Pileus* 9–32 mm in diam., convex to campanulate when young, hemispherical to applanate with age, margin sometimes wavy, slightly deflexed; at centre dull red (8C3), brownish-grey (8D2), greyish-brown (8D3), reddish-brown (8D4, 8E4–8E5) and dark brown (8F5), disc paler to reddish-grey (8B2, 9B2), brownish-grey (9C2), dull red (9B3, 9C3), greyish-magenta (13D3), margin light brown (7D4), brown (7E4) or dull red (9C3); striate none or indistinct, reddish-brown (8E4–8E5), towards the centre up to 1/5 diam.; surface dry, unclearly rugose or none, margin slightly hygrophanous. *Context* white, 1 mm thick, fragile. *Lamellae* sinuate to subdecurrent, 31–33 reaching the stipe, 1–3 tiers of lamellulae, white, irregularly intervenose, edge concolorous, wavy and slightly serrulate. *Stipe* 27–75 × 2–5 mm, central, cylindrical, apex to middle brownish-orange (7C3), dull red (8C3), brownish-grey (7D2), greyish-magenta (14D3), lower part brownish-grey



Figure 13. Basidiomata of *Mycena subulata* Z.W. Liu, Y.P. Ge & Q. Na **a**, **b** *FFAAS0419* **c**, **d** *FFAAS0420* **e–g** *FFAAS0423*, holotype **h** *FFAAS0426* Scale bars: 10 mm (**a–h**). Photographs **a**, **b**, **e**, **f** by Yupeng Ge **c**, **d** by Qin Na **g** by Zewei Liu **h** by Shixin Wang.

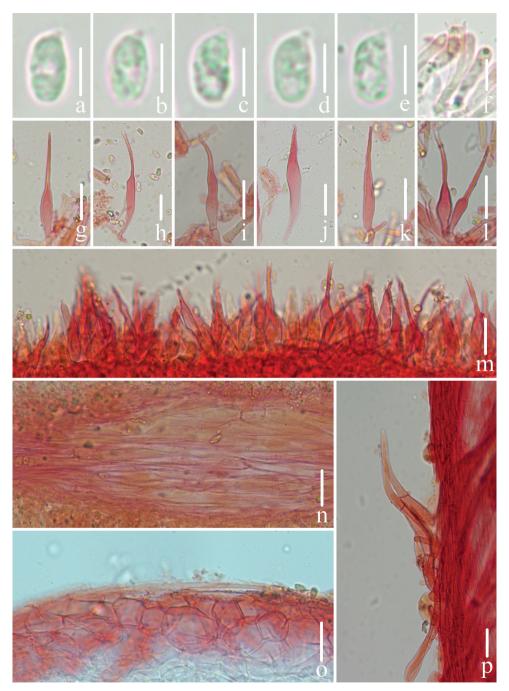


Figure 14. Microscopic features of *Mycena subulata* (*FFAAS0423*, holotype) **a–e** basidiospores **f** basidia **g–m** cheilocystidia **n** lamellar trama **o** pileipellis and hypodermium **p** stipitipellis and caulocystidia. Scale bars: 5 μm (**a–e**); 10 μm (**f**); 20 μm (**g–p**).

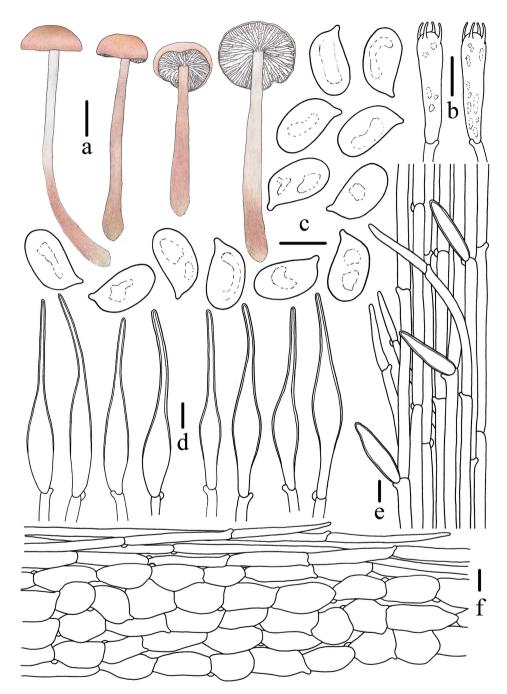


Figure 15. Morphological features of *Mycena subulata* (*FFAAS0423*, holotype) **a** basidiomata **b** basidia **c** basidiospores **d** cheilocystidia **e** stipitipellis and caulocystidia **f** pileipellis and hypodermium. Scale bars: 10 mm (**a**); 5 μm (**c**); 10 μm (**b**, **d–f**). Drawings by Zewei Liu.

(8C2), greyish-brown (8D3), reddish-brown (8D4), fragile, hollow, white granular near apex, base slightly swollen with white fibrils. *Odour* raphanoid, *taste* indistinct.

Basidiospores (100/5/4) 6.0–6.7–7.3(7.9) × 3.3–3.8–4.3(4.6) µm [Q = (1.61)1.65–1.87(1.90), Q = 1.76 ± 0.07] [holotype (40/2/1) (6.0)6.2–6.6–7.1 × 3.4–3.7–4.0(4.2) µm, Q = 1.65–1.87(1.90), Q = 1.78 ± 0.07], elongated ellipsoid, colourless, smooth, thin-walled, amyloid. **Basidia** 23–34 × 5–6 µm, 4-spored, clavate, hyaline, sterigmata 2–3 µm in length. **Cheilocystidia** moderately thick-walled (0.5–0.6 µm), hyaline, narrowly fusiform with long and narrow protuberance, 43–82 × 4–11 µm, protuberance 14–36 × 1–2 µm. **Pleurocystidia** absent. **Pileipellis** a cutis composed of three to four layers of cylindrical cells, 20–89 × 3–7 µm, smooth and thin-walled; terminal cells cylindrical, apically narrow, 24–61 µm in length, the apex 3–4 µm and base 4–7 µm in diam., thin-walled, hyaline. **Hypodermium** formed by fusiform to subglobose hyphae, 21–41 × 17–25 µm, thin-walled, hyaline. **Lamellar trama** subregular, dextrinoid. **Stipitipellis** a cutis composed of hyphae 3–8 µm in diam., smooth, thin-walled, with projecting hyphae 3–8 µm in diam.; **caulocystidia** 29–47 × 6–12 µm, clavate and apices tapered, thick-walled (0.5–0.6 µm), smooth. **Clamps** present in all tissues.

Habitat. Scattered on the litter layer in *Pinus koraiensis*, *Larix gmelinii* and *Tilia* sp. mixed forests.

Known distribution. Heilongjiang Province, China.

Additional material examined. CHINA. Heilongjiang Province: Liangshui National Nature Reserve, Yichun City, 21 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0419* (collection number MY0654); same location, 21 August 2021, Zewei Liu, Yupeng Ge, Qin Na and Shixin Wang, *FFAAS0420* (collection number MY0657); Heilongjiang Province: Taipinggou National Nature Reserve, Hegang City, 3 September 2021, Shixin Wang, *FFAAS0426* (collection number MY0795).

Notes. Cheilocystidia with a long and narrow protuberance is the key microscopic character that distinguishes *M. subulata* and is uncommon in sect. *Calodontes* (Smith 1947; Maas Geesteranus 1992a, 1992b; Grgurinovic 2003; Robich 2003; Chew et al. 2014; Aravindakshan and Manimohan 2015; Aronsen and Læssøe 2016; Na 2019; Liu et al. 2021). Mycena lammiensis Harmaja and M. pelianthina (Fr.) Quél. have similar cheilocystidia, but differ from *M. subulata* by their broader cheilocystidia with purplish-brown contents and having pleurocystidia (Smith 1947; Robich 2003; Aronsen and Læssøe 2016). The cheilocystidia of M. subcorticalis (Cooke & Massee) Sacc. with a protuberance are similar to those of *M. subulata*. However, *M. subcorticalis* has larger and inamyloid spores, a gelatinised pileipellis and a stipitipellis with sparse excrescences (Grgurinovic 2003). More rarely, mucronate cheilocystidia and absence of pleurocystidia have been described for *M. pearsoniana* and its clay pink pileus is similar to that of *M. subulata*, but *M. pearsoniana* differs in having a slightly glutinous pileus when wet and inamyloid spores (Aronsen and Læssøe 2016; Na 2019). Other species that are macroscopically similar to M. subulata, namely M. luteovariegata, M. nullawarrensis and M. pura, can be distinguished by cheilocystidia shape and presence of pleurocystidia (Perry 2002; Robich 2003; Grgurinovic 2003; Harder et al. 2013; Aronsen and Læssøe 2016; Na 2019).

Key to species of sect. Calodontes known worldwide

1	Stipe white2
_	Stipe coloured
2	Pileus whiteMycena subaquosa
_	Pileus coloure
3	Pileus pink and lamellae emarginate, pileipellis without inflated terminal cells
_	Pileus brown and lamellae adnate, pileipellis with fusiform, subcylindrical to lageniform terminal cells
4	Lamellae edge with coloured dots
т	Lamellae edge white or without dots
5	Caulocystidia present, spores almost broader than 4 µm
)	Mycena lammiensis
	Caulocystidia absent, spores almost narrower than 4 µm
_	Caulocysticia absent, spores almost narrower than 4 μm
6	Basidiospores inamyloid
6	
- 7	Basidiospores amyloid
7	Pleurocystidia absent
_	9 Pleurocystidia present
8	Stipitipellis and caulocystidia smooth
_	Stipitipellis and caulocystidia with nodulose excrescences
	Mycena subcorticalis
9	Pileipellis gelatinised, caulocystidia absent, cheilo- and pleurocytsidia base
	uncontracted, disc greyish-red or orange white in pileus
-	Pileipellis not gelatinised, caulocystidia present, cheilo- and pleurocytsidia base contracted, disc wood brown or reddish-brown in pileus
10	Pleurocystidia present11
_	Pleurocystidia absent16
11	Caulocystidia absent, almost cheilocystidia apically mucronate or subcapi-
	tate
_	Caulocystidia present, almost cheilocystidia apically broadly rounded12
12	Caulocystidia with apical excrescences, spores more than 5.6 µm width
_	Caulocystidia without apical excrescences, spores less than 5.6 µm width
	13
13	Cheilocystidia base uncontracted Mycena polycystidiata
_	Cheilocystidia base contracted
14	Stipe brown to dark brown, $Q_{av} = 1.5$
_	Stipe not brown to dark brown, $Q_{av} = 1.5$
-	Superior brown to tark brown, $\zeta_{av} > 1.5$

15	Pileus sulphur yellow to reddish-grey, stipe reddish-grey
	Mycena luteovariegata
_	Pileus generally pinkish or purplish, stipe whitish to pinkish-purple
16	Caulocystidia absent
_	Caulocystidia present
17	Pileus brown to dark brown, spores weakly amyloidMycena seminau
_	Pileus brownish-orange to greyish-yellow, spores amyloid Mycena sinar
18	Cheilocystidia slender fusiform, with distinctly long and narrow protuber-
	ance Mycena subulata
_	Cheilocystidia clavate, utriform, subfusiform, or subcylindrical, with short
	mucronate or none
19	Spores less than 6 µm length
-	Spores more than 6 µm length20
20	Pileus more than 35 mm in diam., lamellae dark brownish-violet to reddish-
	violetMycena diosma
-	Pileus less than 35 mm in diam., lamellae white21
21	Lamellar trama subcellular, pileus lilac to purple Mycena yuezhuoi
-	Lamellar trama subregular, pileus brownish22
22	Cheilocystidia utriform, sometimes clavate, thin-walled, lamellae adnexed to
	emarginateMycena rufobrunnea
_	Cheilocystidia clavate with slightly inflated apex, thick-walled, lamellae sinu-
	ate to subdecurrent

Discussion

Maas Geesteranus (1980) proposed that *Mycena* sect. *Calodontes* could be divided into three subsections based on the colour of the lamellar edge and the amyloid reaction of the basidiospores. Subsequently, taxonomists have followed this division, but opinions have differed on the diagnostic characters that support this classification (Grgurinovic 2003; Robich 2003; Harder et al. 2010; Chew et al. 2014). Some taxonomists classified the subsections according to the amyloid reaction of basidiospores, cheilocystidia and pleuro-cystidia contents and presence or absence of pleurocystidia, but the shapes of cheilocystidia and pleuro-cystidia were not considered (Grgurinovic 2003; Harder et al. 2010; Chew et al. 2014). Robich (2003) also did not consider the shapes of cheilocystidia and pleuro-cystidia, but the colour of the lamellar edge and cheilocystidia contents were emphasised to distinguish subsections. According to the historical infrasectional classification of sect. *Calodontes, M. polycystidiata* could be classified in subsect. *Purae*, whereas *M. rufobrunnea, M. shengshanensis* and *M. subulata* cannot be assigned to a subsection owing to their having amyloid spores and lacking pleurocystidia (Maas Geesteranus 1980; Harder et al. 2010).

Phylogenetic reconstructions do not fully support recognition of three subsections defined by morphological characters; notably, subsect. *Violacellae* and subsect. *Purae*

are polyphyletic in the phylogenies (Harder et al. 2010, 2012, 2013). Chew et al. (2014) supported the views of Harder et al. (2010, 2012) and the new taxa proposed by the former authors were not assigned to a subsection. Additionally, subsect. *Purae* was proved to be polyphyletic in our combined analysis of ITS, *rpb1* and *tef1* dataset, which also supported analysis, based on single gene region (Harder et al. 2013).

The five taxa of *Mycena* sect. *Calodontes* recorded from China show obvious differences in pileus colour and in the shapes of cheilocystidia and pleurocystidia (if present) (Liu et al. 2021). The colour of the pileus includes greyish-rose, reddish-grey, purple, reddish-brown and violet-brown and most show a gradual transition with age. Clavate, obclavate, utriform and fusiform cheilocystidia with a long, narrow protuberance are observed, but pleurocystidia are present only in *M. polycystidiata*. Forms and variations within *M. pura* complex had a wide range of pileus colour, but the shape of cheilocystidia was highly similar and could be clearly distinguished from the four new taxa (Robich 2003).

In our phylogenetic analysis, four new species all formed separate clades with high support and had obvious genetic distance from other species in sect. *Calodontes. Mycena rufobrunnea* is more closely related to the phylogenetic species within *M. pura* complex by Harder et al. (2013). While the other three new species are significantly more distant from *M. pura* complex genetically, *M. shengshanensis* and *M. subulata*, formed a sister relationship with high support from *M. pearsoniana*; *M. polycystidiata* clustered with *M. diosma*, but is poorly supported.

Based on extensive field work in China, most specimens of sect. *Calodontes* have been observed in coniferous forests or mixed coniferous-broadleaved forests in early autumn (Na 2019; Liu et al. 2021). Specimens of the four new taxa described in the present study were collected from Changbai Mountain and the Lesser Khinggan Mountains in northeast China from mixed broadleaf-Korean pine (*Pinus koraiensis*) forests (Zhao et al. 2004; Wang and Guo 2016). In particular, *M. polycystidiata* and *M. subulata* were both distributed in the Liangshui National Nature Reserve on the Lesser Khinggan Mountains, where the dominant forest species is *P. koraiensis*, mixed with fewer *Betula*, *Tilia*, *Quercus* and *Picea* individuals (She et al. 2022). Moreover, more specimens were located in the northern region of China with an average temperature not more than 20 °C in August. For example, the average temperature is 16.4 °C in Liangshui National Nature Reserve and 16.3 °C in Shengshan National Nature Reserve (Liu 2017). Therefore, we speculate that members of this section in China prefer the climate types Dwa, Dwb and Dwc according to the Köppen climate classification (Kottek et al. 2006; Wang et al. 2020).

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References

- Aravindakshan DM, Manimohan P (2015) Mycenas of Kerala. PhD Thesis, University of Calicut, Kerala, India. http://dx.doi.org/10.13140/RG.2.1.2116.4003
- Aronsen A, Læssøe T (2016) The Genus Mycena s.l. Fungi of Northern Europe Vol. 5. Narayana Press, Gylling, Denmark, 373 pp.
- Berkeley MJ (1836) Fungi. In: Smith JE (Eds) The English flora. Longman, Hurst, Rees, Orme, Brown, and Green, London, 5(2): 43. https://doi.org/10.5962/bhl.title.6340
- Chew AL, Tan YS, Desjardin DE, Musa MY, Sabaratnam V (2014) Four new bioluminescent taxa of *Mycena* sect. *Calodontes* from Peninsular Malaysia. Mycologia 106(5): 976–988. https://doi.org/10.3852/13-274
- Clémençon H, Emmett V, Emmett EE (2004) Cytology and Plectology of the Hymenomycetes. Bibliotheca Mycologica, Vol. 199. J. Cramer, Berlin, Stuttgart, 488 pp.
- Edler D, Klein J, Antonelli A, Silvestro D (2020) RaxmlGUI 2.0: A graphical interface and toolkit for phylogenetic analyses using RAxML. Methods in Ecology and Evolution 12(2): 1–5. https://doi.org/10.1111/2041-210X.13512

Fries EM (1821) Systema Mycologicum Vol. l. Lundæ, ex officina Berlingiana, Sweden, 520 pp.

- Ge YP, Liu ZW, Zeng H, Cheng XH, Na Q (2021) Updated description of *Atheniella* (Mycenaceae, Agaricales), including three new species with brightly coloured pilei from Yunnan Province, southwest China. MycoKeys 81: 139–164. https://doi.org/10.3897/mycokeys.81.67773
- Grgurinovic CA (2003) The Genus *Mycena* in south-eastern Australia. Fungal Diversity Press, 329 pp.
- Hall TA (1999) BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98.
- Harder CB, Læssøe T, Kjøller R, Frøslev TG (2010) A comparison between ITS phylogenetic relationships and morphological species recognition within *Mycena* sect. *Calodontes* in Northern Europe. Mycological Progress 9(3): 395–405. https://doi.org/10.1007/s11557-009-0648-7
- Harder CB, Lodge DJ, Petersen RH, Hughes KW, Blanco JC, Frøslev TG, Læssøe T (2012) Amyloidity is not diagnostic for species in the *Mycena pearsoniana* complex (*Mycena* section *Calodontes*). Mycological Progress 11(3): 725–732. https://doi.org/10.1007/s11557-011-0782-x
- Harder CB, Læssøe T, Frøslev TG, Ekelund F, Rosendahl S, Kjøller R (2013) A three-gene phylogeny of the *Mycena pura* complex reveals 11 phylogenetic species and shows ITS to

be unreliable for species identification. Fungal Biology 117(11–12): 764–775. https://doi.org/10.1016/j.funbio.2013.09.004

- Horak E (2005) Röhrlinge und Blätterpilze in Europa: Bestimmungsschlüssel für Polyporales, Boletales, Agaricales, Russulales. Elsevier, Spektrum Akad. Verlag, 555 pp.
- Katoh K, Rozewicki J, Yamada KD (2019) MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics 20(4): 1160–1166. https://doi.org/10.1093/bib/bbx108
- Kornerup A, Wanscher JH (1978) Methuen handbook of colour. Eyre Methuen, London, 252 pp.
- Kottek M, Grieser J, Beck C, Rudolf B, Rubel F (2006) World Map of the Koppen-Geiger Climate Classification Updated. Meteorologische Zeitschrift (Berlin) 15(3): 259–263. https://doi.org/10.1127/0941-2948/2006/0130
- Kuraku S, Zmasek CM, Nishimura O, Katoh K (2013) aLeaves facilitates on-demand exploration of metazoan gene family trees on MAFFT sequence alignment server with enhanced interactivity. Nucleic Acids Research 41(W1): W22–W28. https://doi.org/10.1093/nar/gkt389
- Li Y, Li TH, Yang ZL, Bau T, Dai YC (2015) Atlas of Chinese Macrofungal Resources. Central Chinese Farmer Press, Zhengzhou, China, 1351 pp.
- Liu M (2017) Research of dynamic changes and responses of Korean pine growth under climate change in northeastern China. PhD Thesis, Northeast Forestry University, Heilongjiang, China.
- Liu ZW, Na Q, Cheng XH, Wu XM, Ge YP (2021) Mycena yuezhuoi sp. nov. (Mycenaceae, Agaricales), a purple species from the peninsula areas of China. Phytotaxa 511(2): 148–162. https://doi.org/10.11646/phytotaxa.511.2.3
- Maas Geesteranus RA (1980) Studies in Mycenas-15, A tentative subdivision of the genus *Mycena* in the Northern Hemisphere. Persoonia 11: 93–120.
- Maas Geesteranus RA (1992a) Mycenas of the Northern Hemisphere I. Studies in Mycenas and other papers. Proceedings van de Koninklijke Nederlandse Akademie van Wetenschappen, Amsterdam, North-Holland.
- Maas Geesteranus RA (1992b) Mycenas of the Northern Hemisphere II. Conspectus of the Mycenas of the Northern Hemisphere. Proceedings van de Koninklijke Nederlandse Akademie van Wetenschappen, Amsterdam, North-Holland.
- Na Q (2019) Taxonomy and Molecular Phylogeny of *Mycena* in China. PhD Thesis, Jilin Agriculture University, Jilin, China. https://doi.org/10.27163/d.cnki.gjlnu.2019.000016
- Na Q, Bau T (2019) Recognition of *Mycena* sect. Amparoina sect. nov. (Mycenaceae, Agaricales) including four new species and revision of the limits of sect. Sacchariferae. MycoKeys 52: 103–124. https://doi.org/10.3897/mycokeys.52.34647
- Na Q, Hu YP, Liu ZW, Zeng H, Qi LL, Ding H, Cheng XH, Ge YP (2021) The first reported occurrence of *Leucoinocybe* (Porotheleaceae, Agaricales) in China: *Leucoinocybe lishuiensis* sp. nov. from Zhejiang Province. Nova Hedwigia 113(3–4): 453–469. https://doi. org/10.1127/nova_hedwigia/2021/0661
- Na Q, Hu YP, Zeng H, Song ZZ, Ding H, Cheng XH, Ge YP (2022) Updated taxonomy on *Gerronema* (Porotheleaceae, Agaricales) with three new taxa and one new record from China. MycoKeys 89: 87–120. https://doi.org/10.3897/mycokeys.89.79864

- Osmundson TW, Robert VA, Schoch CL, Baker LJ, Smith A, Robich G, Mizzan L, Garbelotto MM (2013) Filling Gaps in Biodiversity Knowledge for Macrofungi: Contributions and Assessment of an Herbarium Collection DNA Barcode Sequencing Project. PLoS ONE 8(4): e62419. https://doi.org/10.1371/journal.pone.0062419
- Perry BA (2002) A taxonomic investigation of *Mycena* in California. PhD Thesis, San Francisco State University, San Francisco.
- Posada D, Crandall KA (1998) Modeltest: Testing the model of DNA substitution. Bioinformatics 14(9): 817–818. https://doi.org/10.1093/bioinformatics/14.9.817
- Quélet L (1872) Les champignons de Jura et des Vosges (Vol. 1). Imprimerie et Lithographie de Henri Barbier, 332 pp.
- Robich G (2003) Mycena D'Europa. Associazione Micologica Bresadola, Fondazione Centro Studi Micologici, Trento, Vicenza, 728 pp.
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics (Oxford, England) 19(12): 1572–1574. https://doi.org/10.1093/ bioinformatics/btg180
- She DQ, Zhang XT, Xiao L, Zhong ZL, Wang HM, Wang WJ (2022) Plant beta diversity and its influence factors in the Liangshui National Nature Reserve in the central region of the Xiaoxing'an Mountains. Shengwu Duoyangxing 30(03): 15–26. https://doi. org/10.17520/biods.2021274
- Smith AH (1947) North American species of *Mycena*. Unversity of Michigan Press, Ann Arbor, Michigan, 687 pp.
- Wang RZ, Guo QX (2016) Woody plants species-area relationships in a broad-leaved Korean pine forest in the Xiaoxing'an Mountains. Acta Ecologica Sinica 36(13): 4091–4098. https://doi.org/10.5846 /stxb201410282107
- Wang T, Zhou DW, Shen XJ, Fan GH, Zhang H (2020) Köppen's climate classification map for China. Journal of the Meteorological Sciences 40(6): 752–760. https://doi. org/10.3969/2019jms.0040
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR Protocols: a Guide to Methods and Applications. Academic Press, New York, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1
- Yu WJ, Chang C, Qin LW, Zeng NK, Wang SX, Fan YG (2020) Pseudosperma citrinostipes (Inocybaceae), a new species associated with Keteleeria from southwestern China. Phytotaxa 450(1): 8–16. https://doi.org/10.11646/phytotaxa.450.1.2
- Zhao SQ, Fang JY, Zong ZJ, Zhu B, Shen MH (2004) Composition, structure and species diversity of plant communities along an altitudinal gradient on the northern slope of Mt. Changbai, Northeast China. Shengwu Duoyangxing 12(1): 164–173. https://doi. org/10.17520/biods.2004020