

Research Article

New species of *Tropicoporus* (Basidiomycota, Hymenochaetales, Hymenochaetaceae) from India, with a key to Afro-Asian *Tropicoporus* species

Sugantha Gunaseelan¹⁰, Kezhocuyi Kezo¹⁰, Samantha C. Karunarathna^{2,30}, Erfu Yang^{2,40}, Changlin Zhao⁵⁰, Abdallah M. Elgorban⁶⁰, Saowaluck Tibpromma²⁰, Malarvizhi Kaliyaperumal¹⁰

- 1 Centre for Advanced Studies in Botany, University of Madras, Guindy Campus, Chennai 600025, Tamil Nadu, India
- 2 Center for Yunnan Plateau Biological Resources Protection and Utilization, College of Biological Resource and Food Engineering, Qujing Normal University, Qujing, Yunnan 655011, China
- 3 National Institute of Fundamental Studies (NIFS), Kandy, Sri Lanka
- 4 Department of Biology, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand
- 5 College of Biodiversity Conservation, Southwest Forestry University, Kunming 650224, China
- 6 Department of Botany and Microbiology, College of Science, King Saud University, Riyadh 11451, Saudi Arabia

Corresponding authors: Sugantha Gunaseelan (suganthagunaseelan@gmail.com); Malarvizhi Kaliyaperumal (malar.kaliyaperumal@gmail.com)

Abstract

The *Inonotus linteus* complex, predominantly reported from East Asia, Mesoamerica and Caribbean countries, was circumscribed into *Tropicoporus* as one of the new genera, based on morphological and phylogenetic data. The present paper describes four new species of *Tropicoporus* from India. Morphological characteristics and phylogenetic analyses, based on ITS and nLSU data, delimited the new species, which are named *T. cleistanthicola*, *T. indicus*, *T. pseudoindicus* and *T. tamilnaduensis*. The pairwise homoplasy index (PHI) test was done to confirm the distinctive nature of the new species. The traits of Indian species remain distinct from one another, except for the pileate basidiome with the mono-dimitic hyphal system, cystidioles and broadly ellipsoid basidiospores. Descriptions, illustrations, PHI test results and a phylogenetic tree to show the position of the new species are provided. In addition, an identification key to *Tropicoporus* rus in Asia and an African species is given.

Key words: DNA, *Inonotus linteus* complex, mushroom, new species, taxonomy, wood decaying fungi

Introduction

The morpho-taxonomy and phylogenetic analyses, based on the nLSU and ITS genetic markers, revealed that the *Inonotus linteus* complex comprises two clades and are respectively treated as two new genera, *Sanghuangporus* and *Tropicoporus* (Zhou et al. 2015). *Tropicoporus* is characterised by their annual to perennial, resupinate, effused-reflexed to pileate basidiome with glabrous, uncracked to radially cracked pilear surface, homogeneous to duplex context, with or without a black line. A mono-dimitic or dimitic hyphal system with simple septate generative hyphae, presence or absence of cystidioles, presence



Academic editor: Ajay Kumar Gautam Received: 8 December 2023 Accepted: 21 January 2024 Published: 5 February 2024

Citation: Gunaseelan S, Kezo K, Karunarathna SC, Yang E, Zhao C, Elgorban AM, Tibpromma S, Kaliyaperumal M (2024) New species of *Tropicoporus* (Basidiomycota, Hymenochaetales, Hymenochaetaceae) from India, with a key to Afro-Asian *Tropicoporus* species. MycoKeys 102: 29–54. https://doi.org/10.3897/ mycokeys.102.117067

Copyright: © Sugantha Gunaseelan et al. This is an open access article distributed under terms of the Creative Commons Attribution License (Attribution 4.0 International – CC BY 4.0). of hymenial setae with smooth, fairly thick-walled to thick-walled, yellowish, subglobose to ellipsoid basidiospores are microscopic characteristic features of *Tropicoporus* (Zhou et al. 2015; Wu et al. 2022).

A total of forty-eight Tropicoporus species have been recorded in MycoBank with fifteen new species and thirty-three new combinations (as of 12 January 2024). Two new species, namely Tropicoporus excentrodendri L.W. Zhou & Y.C. Dai and T. guanacastensis L.W. Zhou, Y.C. Dai & Vlasák have been delimited, based on nLSU and ITS datasets (Zhou et al. 2015). In addition, Tropicoporus boehmeriae (L.W. Zhou & F. Wu) Y. C. Dai & F. Wu, T. drechsleri Salvador-Montoya & Popoff, T. flabellatus V.R.T. Oliveira, J.R.C. Oliveira-Filho, Xavier de Lima & Gibertoni, T. nullisetus Xavier de Lima, V.R.T. Oliveira & Gibertoni, T. stratificans Y.C. Dai & F. Wu. and T. texanus A.A. Brown, D.P. Lawr. & K. Baumgartner were reported across the world, based on the morphological and molecular data (Wu et al. 2015; Coelho et al. 2016; Salvador-Montoya et al. 2018; Brown et al. 2019; Lima et al. 2022). Recently, seven new species viz., T. angustisulcatus Y.C. Dai & F. Wu, T. hainanicus Y.C. Dai & F. Wu, T. lineatus Y.C. Dai & F. Wu, T. minus Y.C. Dai & F. Wu, T. ravidus Y.C. Dai & F. Wu, T. substratificans Y.C. Dai & F. Wu and T. tenuis Y.C. Dai & F. Wu with twenty-four new combinations were reported, based on the combined dataset of ITS and nLSU sequences (Wu et al. 2022). Of the forty-eight legitimate Tropicoporus species, only T. nullisetus was reported without setae (Lima et al. 2022).

Tropicoporus linteus (also known as *Phellinus linteus*) is used as a renowned Chinese medicine. Due to the presence of *P. linteus* polysaccharides (PLPs), it may play a vital role in anti-aging, anti-bacterial, anti-inflammation, anti-tu-mour, anti-oxidant, hepatoprotective and hypoglycaemic processes (Chen et al. 2019). On the other hand, *Tropicoporus tropicalis* has been reported to cause diseases in humans (Sutton et al. 2005; Haidar et al. 2017; Gupta et al. 2022).

In India, hymenochaetoid fungi from Himachal Pradesh were studied (Kaur et al. 2022). Fourteen hymenochaetoid members were documented from Tamil Nadu (Natarajan and Kolandavelu 1998). Nevertheless, studies on Indian hymenochaetoid fungi, based on molecular data have not been attempted, which makes it difficult to understand their evolutionary history, phylogenetic relationships and accuracy of species delimitation. This study is the first attempt to describe new *Tropicoporus* species from India, based on morphology and molecular evidence. In addition, an identification key to Afro-Asian *Tropicoporus* is given.

Materials and methods

Morphological analyses

Eight specimens were collected from parts of Eastern Ghats and the plain region of Tamil Nadu, southern India. Macro-morphological characteristics such as shape, size of basidiome, perennial or annual, colour, texture, margin (acute or obtuse), context (homogenous, duplex with or without black line), tube layer (colour, length, stratification) and pores (size and shape) were examined in the fresh sample and recorded. Colour descriptions were based on the Methuen Handbook (Kornerup and Wanscher 1978). To analyse the micro-morphological characteristics, free-hand sections of dry specimens were mounted in water, 5% potassium hydroxide (KOH) (v/w), cotton blue (CB) and Melzer's reagent (IK). Sections were studied and photos were taken at magnification up to 1000× using a LABOMED OPTIC-CX BINO LED microscope. The drawings were made using LABOMED CxL2 compound microscope. Microscopic measurements and illustrations were made in 5% KOH solution. Basidiospores measurements (as minimum-mean-maximum) and Q values (length/width ratios) were recorded. The following abbreviations are used: IKI⁻ (inamyloid), IKI⁺ (amyloid), CB⁻ (acyanophilous), CB⁺ (cyanophilous), L = mean spore length (arithmetic average of all spores), Q = (variation in the L/W ratios; and basidium length excludes the lengths of the sterigmata) and n = number of spores measured. For measuring the spores, an average of 50 spores were considered. Specimens in this study were deposited in the Madras University Botany Laboratory (**MUBL**), Centre for Advanced Studies in Botany, University of Madras, India.

Genomic DNA extraction, PCR amplification and sequencing

Extraction of total genomic DNA from mycelium and dried basidiome followed the protocol of Doyle and Doyle (1987), modified by Góes-Neto et al. (2005). The ITS and nLSU regions were amplified and sequenced with the primers ITS1/ITS4 and LR0R/LR7, respectively (Vilgalys and Hester 1990; White et al. 1990). The polymerase chain reaction (PCR) procedure for ITS was as follows: initial denaturation at 95 °C for 3 min, followed by 32 cycles at 95 °C for 30 s, 52 °C for 30 s and 72 °C for 1 min and a final extension of 72 °C for 3 min. The PCR procedure followed for nLSU was as follows: initial denaturation at 94 °C for 1 min, followed by 34 cycles at 94 °C for 30 s, 45 °C for 30 s and 72 °C for 1.5 min and final extension at 72 °C for 10 min. The PCR products were sequenced at Eurofins Genomics India Pvt. Ltd., Karnataka, India.

Phylogenetic analyses

The dataset comprised ITS and nLSU sequences of Fulvifomes, Inonotus, Phellinus, Phylloporia, Sanghuangporus and Tropicoporus retrieved from GenBank (NCBI), along with the outgroup (Fomitiporella caryophylli, CBS 448.76) and the newly-generated sequences (deposited at GenBank (Sayers et al. 2023); for accession numbers, see Table 1). The dataset was aligned using MEGA X v.10.0.2 configured for Windows and edited manually to increase the alignment similarity (Kumar et al. 2018). The Maximum Likelihood (ML) tree was constructed using raxmlGUI 2.0 (Edler et al. 2020) with the best-fit evolutionary model estimated by jModelTest 2.1.10 with 1000 rapid bootstrap inferences (BS) (Guindon and Gascuel 2003; Darriba et al. 2012). Bayesian Inference (BI) was performed using MrBayes 3.2.7a with two independent runs and six chains of Metropolis-coupled Markov Chain Monte Carlo iterations for 2,000,000 generations and trees were sampled every 100 generations (Ronguist et al. 2012). A proportion of 0. 25% of all trees (nLSU+ITS, ITS, nLSU) were discarded as burn-in. The final alignments and the retrieved topologies were deposited in TreeBASE (http:// purl.org/phylo/treebase/phylows/study/TB2:S31000).

Table 1. Names, strain numbe	ers, countries of co	ollection and the c	orresponding GenBank
accession numbers of the se	quences used in tl	his study.	

0		0	Accession numbers			
Species	Strain numbers	Country	ITS	nLSU		
Fomitiporella caryophylli	CBS 448.76	_	AY558611	AY558611		
Fulvifomes centroamericanus [™]	JV0611_III	Guatemala	KX960763	KX960764		
F. elaeodendri	CMW47825	South Africa	MH599094	MH599134		
F. nilgheriensis	CBS 209.36	USA	AY558633	AY059023		
F. thailandicus [⊤]	LWZ 2014073-11	Thailand	KR905672	KR905665		
Inonotus pachyphloeus	Wu 0407.6	Taiwan	KP030785	KP030770		
Phellinus laevigatus	CBS 122.40	USA	MH856059	MH867554		
P. populicola [⊤]	CBS 638.75	Finland	MH860960	MH872729		
Phylloporia nodostipitata	FLOR:51153	Brazil	KJ639057	KJ631414		
Sanghuangporus alpinus	Cui12485	China	MF772781	MF772799		
S. baumii	Cui 11769	China	MF772784	MF772803		
S. lonicericola	Dai 8376	China	JQ860308	MF772805		
S. lonicerinus	Dai 17093	China	MF772788	MF772807		
S. quercicola	Dai 13947	China	KY328309	MF772809		
S. sanghuang	Cui 14419	China	MF772789	MF772810		
S. vaninii	DMR 95-1-T	North America	KU139198	KU139258		
S. vitexicola	Wu 2006-21	_	MT906620	MZ437416		
S. weigelae	Dai 16077	China	MF772794	MF772815		
S. zonatus	Dai 10841	China	JQ860306	KP030775		
Tropicoporus angustisulcatus	Dai 17409	Brazil	MZ484584	MZ437417		
T. angustisulcatus [™]	JV 1808/83	French Guiana	MZ484585	MZ437418		
T. boehmeriae [⊤]	LWZ 20140729-10	Thailand	KT223640	MT319393		
T. boehmeriae	LWZ 20140729-13	Thailand	KT223641	MT319394		
	Dai 20522	China	MZ484586	MZ437419		
	Dai 20617	China	MZ484587	MZ437420		
T. cleistanthicola [⊤]	MUBL1089	India	OR272292	OR272337		
T. cleistanthicola	MUBL1090	India	OR272291	OR272336		
T. cubensis	MUCL 47113	Cuba	JQ860324	KP030777		
	MUCL 47079	Cuba	JQ860325	KP030776		
T. dependens	JV 0409/12-J	USA	KC778777	MF772818		
T. detonsus	CBS 617.89	-	AF534077	AY059037		
	IDR 1300012986	USA	KF695121	KF695122		
T. drechsleri [™]	CTES:570140	Argentina	MG242439	MG242444		
T. drechsleri	CTES:570144	Argentina	MG242437	MG242442		
T. excentrodendri	Yuan 6234	China	KP030791	_		
	Yuan 6229	China	KP030789	-		
T. flabellatus [⊤]	VRT0873	Brazil	MT908376	MT906643		
T. guanacastensis	0 19228	Costa Rica	KP030794	_		
T. guanacastensis [⊤]	JV 1408_25	Costa Rica	KP030793	KP030778		
T. hainanicus [™]	Dai 17705	China MZ484588		MZ437421		
T. indicus [⊤]	MUBL1083	India	OR272293	OR272338		
T. indicus	MUBL1084	India	OR272294	OR272339		
T. lineatus [⊤]	Dai 21196	Malaysia	MZ484594	MZ437426		
T. linteus	JV 0904/64	USA	JQ860322	JX467701		
	-		1			

Species	Stroin numbers	Country	Accession numbers		
Species	Strain numbers	Country	ITS	nLSU	
T. linteus	JV 0904/140	USA	JQ860323	KP030780	
T. minor [™]	Dai 21139	China	MZ484592	MZ437424	
T. minus	Dai 18487A	China	MZ484590	MZ437422	
	Dai 21183	China	MZ484593	MZ437425	
T. nullisetus [⊤]	VXLF616	Brazil	MN795129	MN812261	
T. nullisetus	VRTO195	Brazil	MN795118	MN812254	
T. pseudoindicus [⊤]	MUBL1087	India	OR272295	OR272340	
T. pseudoindicus	MUBL1088	India	OR272296	OR272341	
T. pseudolinteus	JV0402/35-K	Venezuela	KC778781	MF772820	
	JV 0312/22.10-J	Venezuela	KC778780	-	
T. ravidus [⊤]	Dai 18165	China	MZ484595	MZ437427	
T. rudis	0 915614	Rwanda	KP030796	-	
	0 915617	Tanzania	KP030797	MH101016	
T. sideroxylicola	JV 1207/4.3-J	USA	KC778783	-	
T. sideroxylicola [⊤]	JV 0409/30-J	USA	KC778782	-	
T. stratificans [⊤]	SMDB 14732	Brazil	KM199689	-	
T. stratificans	VRT0884	Brazil	MN795124	MN812266	
T. substratificans [⊤]	JV 1908/80	French Guiana	MZ484597	MZ437429	
T. tamilnaduensis [⊤]	MUBL1085	India	OR272297	OR272343	
T. tamilnaduensis	MUBL1086	India	_	OR272344	
T. tenuis [⊤]	Dai 19699	China	MZ484598	MZ437430	
T. tenuis	Dai 19724	China	MZ484599	MZ437431	
T. texanus [⊤]	CBS 145357	USA	NR_168219	NG_068906	
T. texanus	TX8	USA	MN108123	MN113949	
T. tropicalis	UTHSC 02-617	USA	AY641432	-	
	UAMH 10376	USA	AY599487	-	

 $^{\intercal}$ = Type material and "-" refers to the data unavailability. Sequences generated from the present study are indicated in bold.

Genealogical concordance phylogenetic species recognition analysis

Genealogical concordance phylogenetic species recognition analysis (GCPSR) by the pairwise homoplasy index (PHI) test was used to determine the recombination level within closely-related species (Bruen et al. 2006). The data were analysed by the software SplitsTree 4 (Bruen et al. 2006; Huson and Bryant 2006). The relationships between closely related taxa were visualised by constructing split graphs from concatenated datasets, using the LogDet transformation and splits decomposition options. If the PHI test value is (Φ w) \leq 0.05, it indicates significant recombination within the dataset. This is an important method to provide further evidence to justify a species. All results are shown in Fig. 1.

Results

Phylogenetic analyses

In total, eight new sequences of the ITS and seven new sequences of the nLSU regions were generated and submitted to GenBank (Table 1). Additionally, 62 taxa (52 nLSU and 62 ITS sequences) were retrieved from GenBank

Sugantha Gunaseelan et al.: New species of Tropicoporus from India



Figure 1. Split graphs show the results of the PHI test of the new species, *Tropicoporus indicus*, *T. tamilnaduensis*, *T. pseudoindicus*, *T. cleistanthicola* and their most closely-related species *T. rudis*, using LogDet transformation and split decomposition options. The PHI test result $\Phi w \le 0.05$ indicates that there is a significant recombination within the dataset.

(Table 1). The combined nLSU and ITS dataset were aligned and the multiple sequence alignment consists of 1,820 characters (914 for nLSU and 902 for ITS) of which 1,017 were constant, 962 were variable and 570 (31%) were parsimony informative. The best-fit evolutionary model (GAMMA+P-Invar Model) was estimated by jModelTest 2.1.10 for the combined datasets. The Maximum Likelihood (ML) trees were constructed using raxmlGUI 2.0 with 1,000 rapid bootstrap inferences (BS). The Bayesian analysis was run for 2,000,000 generations and the average standard deviation reached 0.010166. The phylogenetic topology was selected from Bayesian analysis. The Maximum Likelihood bootstrap values \ge 60% and the Bayesian posterior probabilities \ge 0.90 are summarised in Fig. 2.

Taxonomical descriptions of the four novel species of Tropicoporus

Tropicoporus cleistanthicola **S. Gunaseelan & M. Kaliyaperumal, sp. nov.** MycoBank No: 849484 Figs 3, 4

Etymology. The specific epithet *cleistanthicola* (Lat.) refers to the host *Cleistanthus collinus*.

Diagnosis. Tropicoporus cleistanthicola is characterised by perennial, effused-reflexed to pileate, applanate to triquetrous basidiome with narrowly zonate, glabrous, meagrely warted pilear surface, acute margin, homogenous context, mono-dimitic hyphal system, presence of cystidioles and subglobose to broadly ellipsoid basidiospores measuring $4.7-5.4 \times 4.2-4.9 \mu m$.

Type. INDIA, Tamil Nadu, Thiruvannamalai District, Jawadhu Hills, 12°54'24"N, 78°87'75"E; 15 Nov 2019; Sugantha Gunaseelan; on a living angiosperm tree (*Cleistanthus collinus*); SMK-PMP11 (MUBL1089; Holotype); GenBank: OR272292 (ITS); OR272337 (nLSU).

Description. *Basidiome* perennial, pileate, solitary, hard corky and without distinctive odour or taste when fresh, woody hard and light in weight when dry. *Pilei* effused-reflexed to pileate, dimidiate, triquetrous in section, projecting up to 4 cm, 6.5 cm wide and 3 cm thick at the base; Pileal surface narrowly zonate, glabrous, meagrely warted near attachment, yellowish-brown (5E6; 5E8) to dark



Figure 2. Molecular phylogeny of four new Indian *Tropicoporus* species and other hymenochaetoid species inferred from combined ITS and nLSU sequences. The topology is from the Bayesian analysis. Maximum Likelihood bootstrap values and Bayesian posterior probabilities, above 60% and 0.9, respectively, are labelled at the nodes. The newly-generated sequences are coloured and bold; the type specimens are in bold.

brown (6F5), turning dark brown (7F4) to greyish-brown (6F3). *Margin* acute, 1 mm thick, light brown (6D5). *Pore surface* brown (6E6) to dark brown (6F7); sterile margin up to 2 mm wide, light brown (6D5); pores circular, 5–7 per mm.



Figure 3. Tropicoporus cleistanthicola (MUBL1089 holotype) A basidiomata (Holotype) B pore surface C cross-section of basidiome (arrows indicating stratified tube layers) D hymenial setae E-H basidiospores: E basidiospores in water F basidiospores in KOH G basidiospore in cotton blue H basidiospore in Melzer's reagent. Scale bars: 1 cm (A-C); 5 μ m (D-H).

Context homogenous, up to 1.5 cm thick, brown (6E8). *Tubes* up to 0.5 cm long, tube layers distinctly stratified, each stratum up to 2 mm, brown (6E7).

Hyphal structures. Hyphal system monomitic in the context and dimitic in the trama, tissue darkening with KOH without hyphal swelling.

Context. Generative hyphae, thin to thick-walled, hyaline to golden yellow, simple septate, rarely branched, $2-5 \,\mu\text{m}$ diam.

Trama. Generative hyphae, dominant, thin to thick-walled, hyaline to pale yellow, septate, occasionally branched, $2-4 \mu m$ diam. Skeletal hyphae thick-walled with narrow to wide lumen, yellowish-brown, aseptate, unbranched, $2-3.5 \mu m$ diam.

Hymenium. Hymenial setae dark brown, thick-walled, ventricose to subulate with sharp to blunt tips, $5-32 \times 4-5.5 \mu m$. Cystidia absent. Cystidioles hyaline, thin-walled, ventricose to fusoid with elongated tapering apical portion, $7-45 \times 2-5 \mu m$. Basidia clavate to broadly clavate, $7-15 \times 2.7-6.2 \mu m$, with four sterigmata and a simple septum at the base. Basidioles clavate, $5-13 \times 3.5-6 \mu m$. Basidiospores broadly ellipsoid to subglobose, pale yellow in water, turning golden yellow to brown in KOH, thick-walled, smooth, CB⁻, IKI⁻, (4.7-) 4.9-5.2 (-5.4) × (4.2-) 4.5-4.7 (-4.9) μm (n = 50/2), Q = 1.1 (Q range 1.05-1.2).

Habitat and distribution. Basidiomes were found on living trees of *Cleistanthus collinus* (Phyllanthaceae), distributed in Jawadhu Hills, Thiruvannamalai District, Tamil Nadu, India.



Figure 4. *Tropicoporus cleistanthicola* (MUBL1089 holotype) A contextual hyphae B tramal hyphae C hymenial setae D cystidioles E basidioles F basidia G basidiospores. Scale bars: 5 µm.

Additional material examined. INDIA, Tamil Nadu, Thiruvannamalai District, Jawadhu Hills; 12°51'20"N, 78°73'71"E; 15 Nov 2019; Sugantha Gunaseelan; on a living angiosperm tree (*Cleistanthus collinus*); SMK-PMP14 (MUBL1090, Paratype); GenBank: OR272291 (ITS); OR272336 (nLSU).

Notes. The present phylogenetic study indicated that T. cleistanthicola is sister to T. rudis with significant support (92% ML/0.9 BPP). However, T. rudis has applanate basidiomes with fulvous, velvety, concentrically zonate, matted, rimose pilear surface, whereas T. cleistanthicola has triquetrous basidiome and glabrous pilear surface with infrequent warts without cracks. Tropicoporus cleistanthicola and T. rudis are comparable only in mono-dimitic hyphal system and T. rudis lacks cystidioles and has larger basidiospores (4.9-6 × 4-4.8 µm) (Wu et al. 2022). Despite sharing pileate basidiomes, mono-dimitic hyphal system and presence of cystidioles in T. linteus, T. cleistanthicola differs by having effused-reflexed to pileate, narrowly zonate, meagrely warted pilear surface (Tian et al. 2013; Wu et al. 2022). Tropicoporus cleistanthicola resembles T. angustisulcatus, T. dependens, T. excentrodendri, T. substratificans and T. lineatus by sharing pileate, triquetrous basidiomes with concentrically zonation and presence of cystidioles, but T. cleistanthicola differs by having a mono-dimitic hyphal system and spore size $(4.7-5.4 \times 4.2-4.9 \ \mu m)$ (Zhou et al. 2015; Wu et al. 2022). Tropicoporus cleistanthicola and T. drechsleri are similar in having pileate basidiomes and a mono-dimitic hyphal system with the presence of cystidioles, but T. cleistanthicola differs by having smaller pores (5-7 pores/mm) and larger basidiospores (Salvador-Montoya et al. 2018). Except for sharing a mono-dimitic hyphal system and indistinctly stratified tube layers, T. cleistanthicola differs from T. flabellatus and T. guanacastensis in pores (size and shape), basidiospore shape and absence of cystidioles (Zhou et al. 2015; Lima et al. 2022).

Tropicoporus indicus S. Gunaseelan & M. Kaliyaperumal, sp. nov.

MycoBank No: 849482 Figs 5, 6

Etymology. The species epithet *"indicus"* (Lat.): referring to the species being collected from India.

Diagnosis. Tropicoporus indicus is characterised by applanate to meagrely triquetrous basidiome with concentrically zonate, sulcate, glabrous, deeply cracked to rimose pilear surface, homogenous context, acute margin, mono-dimitic hyphal system, presence of cystidioles, subglobose to broadly ellipsoid basidiospores measuring $5-6 \times 4.2-4.9 \mu m$.

Type. INDIA, Tamil Nadu, Kallakurichi District, Kalvarayan Hills; 11°91'30"N, 78°57'86"E; 29 Sep 2022; Sugantha Gunaseelan; on living angiosperm tree of *Albizia amara*; SMK- MK2 (MUBL1083, Holotype); GenBank: OR272293 (ITS); OR272338 (nLSU).

Description. *Basidiome* perennial, pileate, woody and without distinctive odour or taste when fresh, hard when dry. *Pilei* applanate to meagrely trique-trous, projecting up to 5.5 cm, 7 cm wide and 4 cm thick at the base; pileal surface concentrically zonate, sulcate, glabrous, deeply cracked to rimose near attachment, dark brown (7E4), greyish-brown (7F3). *Margin* velutinate, 2 mm thick, acute, brown (6E7). *Pore surface* light brown (5D5) to yellowish-brown (5E7); pores circular, 4–6 per mm. *Context* homogenous, up to 0.8 cm thick, brown (6E8) to dark brown (6F8). *Tubes* woody hard, up to 2.5 cm long, brown (6E7); tube layers stratified, each stratum up to 0.5 cm long.



Figure 5. *Tropicoporus indicus* (MUBL1083 holotype) A basidiomata (Holotype) B basidiome (SMK-MK2a- Isotype) C pore surface D cross-section of basidiome (arrows indicating stratified tube layers) E hymenial setae F cystidioles G–J basidiospores: G basidiospores in water H basidiospores in KOH I basidiospores in cotton blue J basidiospores in Melzer's reagent. Scale bars: 1 cm (A–D); 5 μ m (E–J).

Hyphal structures. Hyphal system monomitic in the context and dimitic in the trama, tissue darkening with KOH without swelling

Context. Generative hyphae, thin to thick-walled, hyaline to golden yellow, simple septate, rarely branched, $2-5 \,\mu m$ diam.

Trama. Generative hyphae dominant, thin to thick-walled, hyaline to pale yellow, septate, occasionally branched, $2-4.5 \mu m$ diam. Skeletal hyphae thick-walled with narrow to wide lumen, yellowish-brown, aseptate, unbranched, $2-4 \mu m$ diam.

Hymenium. Hymenial setae dark brown, thick-walled, ventricose to subulate with sharp to blunt tips, $7-28 \times 3-5 \mu m$. Cystidia absent. Cystidioles hyaline, thin-walled, ventricose to fusoid with elongated tapering apical portion, $5-21 \times 3-5 \mu m$. Basidia clavate to broadly clavate, $7-17 \times 3-6 \mu m$, with four sterigmata and a simple septum at the base. Basidioles clavate, $5-14 \times 3-5 \mu m$. Basidiospores broadly ellipsoid to subglobose, pale yellow in water, turning golden



Figure 6. *Tropicoporus indicus* (MUBL1083 holotype) **A** contextual hyphae **B** tramal hyphae **C** hymenial setae **D** cystidioles **E** basidioles **F** basidia **G** basidiospores. Scale bars: 5 µm.

yellow to brown in KOH, thick-walled, smooth, CB⁻, IKI⁻, (5–) 5.3–5.8 (– 6) × (4.2–) 4.7–4.9 μ m (n = 50/2), Q = 1.16 (Q range 1.05–1.3).

Habitat and distribution. Basidiomes were found on living trees of Fabaceae members (*Albizia amara* and *Prosopis cineraria*), distributed in Kalvarayan Hills, Kallakurichi District, Tamil Nadu, India.

Additional material examined. INDIA, Tamil Nadu, Kallakurichi District, Kalvarayan Hills; 11°90'39"N, 78°55'69"E; on a living angiosperm tree (*Prosopis cineraria*); 29 Sep 2022; Kezhocuyi Kezo; SMK-MK2a (MUBL1084, Paratype); GenBank: OR272294 (ITS); OR272339 (nLSU).

Notes. Phylogenetically, Tropicoporus indicus was recovered in the T. linteus clade. Tropicoporus indicus is similar to T. linteus by sharing, pileate, dimidiate basidiomes, concentrically sulcate pilear surface, zonate context, smaller pores (5–7/mm), a mono-dimitic hyphal system and presence of cystidioles. While varying in the nature of cracks, T. linteus has more or less cracked basidiomes, T. indicus has irregular deep cracks in basidiomes, with larger basidiospores (T. linteus $4.8-5.7 \times 3.8-4.8 \mu$ m and T. indicus $5-6 \times 4.2-4.9 \mu$ m) (Tian et al. 2013; Wu et al. 2022). Tropicoporus indicus and T. rudis share a mono-dimitic hyphal system, but T. indicus differs from T. rudis in having zonate, sulcate, deeply cracked to rimose basidiomes and larger pores (4-6/mm). In these regards, T. rudis is characterised by fulvous, velvety, concentrically zonate, matted, rimose basidiome and smaller pores (6-7/mm) (Wu et al. 2022). Tropicoporus indicus differs from T. angustisulcatus, T. excentrodendri, T. lineatus and T. substratificans in having mono-dimitic hyphal system and a cracked basidiome (Zhou et al. 2015; Wu et al. 2022). Tropicoporus indicus and T. flabellatus are similar in having a mono-dimitic hyphal system, but differ significantly by having concentrical zones, sulcate, glabrous, deeply-cracked to rimose pilear surface with larger pores (4-6/mm) and broadly ellipsoid to subglobose spores $(5-6 \times 4.2-4.9 \,\mu\text{m})$. Tropicoporus flabellatus, in contrast, has a velutinate pilear surface, uncracked basidiomes with smaller pores (7-9/mm) and smaller basidiospores (4.5-5 × 3.5-4 µm) (Lima et al. 2022). Tropicoporus indicus and T. guanacastensis are similar in having sulcate, cracked basidiome, stratified tube, mono-dimitic hyphal system and ventricose setae. However, the former differs in larger pores (4–6/mm) and larger basidiospores (T. indicus 5–6 \times 4.2-4.9 µm vs. T. guanacastensis 4.1-5 × 3.1-4 µm) (Zhou et al. 2015). Tropicoporus indicus and T. drechsleri share concentrically sulcate deeply-cracked pilei with mono-dimitic hyphal system, larger pores (< 6/mm) and presence of cystidioles, but the South American species differs in basidiospore size (4-5.5 × 3-4.5 µm) (Salvador-Montoya et al. 2018).

Tropicoporus pseudoindicus S. Gunaseelan & M. Kaliyaperumal, sp. nov. MycoBank No: 849483

Figs 7, 8

Etymology. The species *pseudoindicus* signifies the close morphological and phylogenetic relationships with the species *Tropicoporus indicus*.

Diagnosis. Tropicoporus pseudoindicus is characterised by applanate to meagrely ungulate to triquetrous basidiome with broadly zonate, distinctly cracked by radial fissures, sulcate pilear surface, duplex context with black line,



Figure 7. Tropicoporus pseudoindicus (MUBL1085 holotype) A basidiomata (Holotype) B pore surface C cross-section of basidiome (arrows indicating stratified tube layers and duplex context with black line) D hymenial setae E-H basidiospores: E basidiospores in water F basidiospores in KOH G basidiospores in cotton blue H basidiospores in Melzer's reagent. Scale bars: 1 cm (A-C); 5 µm (D-H).

acute to obtuse margin, pores 6–8/mm, mono-dimitic hyphal system, presence of cystidioles, subglobose to broadly ellipsoid basidiospores measuring 4–5.2 \times 3.7–4.7 $\mu m.$

Type. INDIA, Tamil Nadu, Kallakurichi District, Kalvarayan Hills; 11°86'98"N, 78°55'68"E; 29 Sep. 2022; Sugantha Gunaseelan; on a living angiosperm tree (*Albizia amara*); SMK-MK4 (MUBL1087, Holotype); GenBank: OR272295 (ITS); OR272340 (nLSU).

Description. *Basidiome* perennial, pileate, woody and without distinctive odour or taste when fresh, hard and light in weight when dry. *Pilei* applanate, meagrely ungulate to triquetrous, dimidiate, projecting up to 5 cm, 8 cm wide and 3.5 cm thick at base; pileal surface broadly zonate, distinctly cracked by radial fissures, sulcate, brown (6E8), dark brown (7F4) to greyish-brown (6F3). *Margin* acute to obtuse, up to 3 mm thick, light brown (6D5). *Pore surface* brown (6E6) to dark brown (7E6); sterile margin brown (6E6), up to 2 mm wide; pores circular, 6–8 per mm. *Context* duplex with black line, woody hard, up to 1.2 cm thick, several black lines present along context, brown (6E7) to dark brown (7F6). *Tubes* up to 1.5 cm long, annual layers distinct, each stratum up to 0.3 cm, brown (7E8) to dark brown (6F8).

Hyphal structures. Hyphal system monomitic in the context and dimitic in the trama, tissue darkening with KOH without swelling.

Context. Generative hyphae, thin to thick-walled, hyaline to golden yellow, simple septate, rarely branched, $2-5 \,\mu m$ diam.

Trama. Generative hyphae, dominant, thin to thick-walled, hyaline to pale yellow, septate, occasionally branched, $2-4 \mu m$ diam. Skeletal hyphae, thick-walled with narrow to wide lumen, yellowish-brown, aseptate, unbranched, $2-3.5 \mu m$ diam.

Hymenium. Hymenial setae dark brown, thick-walled, ventricose to subulate with sharp to blunt tips, rarely with lateral appendage, $5-18 \times 3-5.5 \,\mu$ m. Cystidia absent. Cystidioles hyaline, thin-walled, ventricose to fusoid with elongated tapering apical portion, $7-52 \times 2.5-5.2 \,\mu$ m. Basidia clavate to broadly clavate, $7-15 \times 2.7-6.2 \,\mu$ m, with four sterigmata and a simple septum at the base. Basidioles clavate, $5-13 \times 3.5-6 \,\mu$ m. Basidiospores broadly ellipsoid to subglobose, pale yellow in water, turning golden yellow to brown in KOH, thick-walled, smooth, CB⁻, IKI⁻, (4–) 4.2–5 (–5.2) × (3.7–) 4–4.5 (–4.7) μ m (n = 50/2) and Q = 1.14 (Q range 1.05–1.25).

Habitat and distribution. Basidiomes were found on living trees of Fabaceae members (*Albizia amara* and *Peltophorum pterocarpum*), distributed in Kalvarayan Hills, Kallakurichi District, Tamil Nadu, India.

Additional material examined. INDIA, Tamil Nadu, Kallakurichi District, Kalvarayan Hills; 11°87'33"N, 78°42'78"E; 29 Sep 2022; Kezhocuyi Kezo; on a living angiosperm tree (*Peltophorum pterocarpum*); SMK-MK4a (MUBL1088, Paratype); GenBank: OR272296 (ITS); OR272341 (nLSU).

Notes. Tropicoporus pseudoindicus and T. drechsleri, share similar characteristics, such as applanate basidiomes with mono-dimitic hyphal system and the presence of cystidioles; however, T. pseudoindicus differs in having smaller pores (6-8/mm) and larger basidiospores (*T. pseudoindicus* $4-5.2 \times 3.7-4.7 \mu m vs.$ T. drechsleri 4–5.5 × 3–4.5 µm) (Salvador-Montoya et al. 2018). Tropicoporus pseudoindicus resembles T. rudis in having a mono-dimitic hyphal system, presence of cystidioles, and basidiospore shape, but differs from T. rudis in having distinctly cracked, fissured to sulcate pilei, duplex context and smaller basidiospores (4–5.2 × 3.7–4.7 μm) (Vlasák et al. 2013). Tropicoporus pseudoindicus differs from T. guanacastensis in having cystidioles and subglobose to broadly ellipsoidal spores (T. pseudoindicus 4–5.2 × 3.7–4.7 µm vs. T. guanacastensis 4.1–5.0 × 3.1–4.0 μm) (Tian et al. 2013). Tropicoporus pseudoindicus and T. linteus share similar pileate basidiomes with sulcate pilear surface, smaller pores (5–7/mm), mono-dimitic hyphal system and presence of cystidioles. However, the former differs in basidiospore size (T. linteus $4.8-5.7 \times 3.8-4.8 \mu m$ vs. T. pseudoindicus 4-5.2 × 3.7-4.7 µm) (Tian et al. 2013; Wu et al. 2022). Tropicoporus pseudoindicus and T. flabellatus are similar in their mono-dimitic hyphal system, but differ significantly in pilear characteristics, absence of cystidioles and size and shape of basidiospores (T. flabellatus 4.5-5 × 3.5-4 µm vs. T. pseudoindicus (4-) 4.2-5 (-5.2) × (3.7-) 4-4.5 (-4.7) μm) (Lima et al. 2022). Tropicoporus pseudoindicus differs from some other reported Tropicoporus species (namely T. angustisulcatus, T. excentrodendri, T. lineatus and T. substratificans) in having concentrically zonate, glabrous, distinctly cracked pileal surface and a mono-dimitic hyphal system (Zhou et al. 2015; Wu et al. 2022).



Figure 8. *Tropicoporus pseudoindicus* (MUBL1085 holotype) A contextual hyphae B tramal hyphae C hymenial setae D cystidioles E basidioles F basidia G basidiospores. Scale bars: 5 µm.

Tropicoporus tamilnaduensis M. Kaliyaperumal & S. Gunaseelan, sp. nov.

MycoBank No: 849481 Figs 9, 10

Etymology. The species epithet *tamilnaduensis* refers to the locality of the type specimen (Tamil Nadu).

Diagnosis. Tropicoporus tamilnaduensis is characterised by applanate to meagrely ungulate basidiome with glabrous, broadly zonate, sulcate and deeply irregularly cracked pilear surface, homogenous context, obtuse margin, pores 4-5/mm, mono-dimitic hyphal system, presence of cystidioles, subglobose to broadly ellipsoid basidiospores measuring $4.5-5.7 \times 3.5-4.7 \mu$ m.

Type. INDIA, Tamil Nadu, Cuddalore District, Thaiyalkunampattinam, Kanni Tamil Nadu; 11°59'18"N, 79°60'17"E; 31 Dec 2022; Malarvizhi Kaliyaperumal; on a living angiosperm tree (*Madhuca longifolia*); MKDM02 (MUBL1085, holotype); GenBank: OR272297 (ITS); OR272343 (nLSU).

Description. *Basidiome* perennial, pileate, without distinctive odour or taste when fresh, woody hard and light in weight when dry. *Pilei* applanate to meagrely ungulate, projecting up to 5 cm, 8 cm wide and 4 cm thick at base; pileal surface glabrous, broadly zonate, sulcate, deeply irregularly cracked near attachment, brown (6E7), yellowish-brown (5F4) to golden brown (7F7) turning greyish-brown (5F3). *Margin* obtuse, 4 mm thick, light brown (6D5). *Pore surface* brown (6E6), sterile margin yellowish-brown (5E6), up to 2 mm wide; pores circular, 4–5 per mm; dissepiments thick, entire. *Context* homogenous, zonate, brown (6D7) to dark brown (6F8), woody hard, up to 2 cm thick. *Tubes* brown (6E6), up to 2 cm long, annual layers distinct, each stratum up to 0.3 cm long.

Hyphal structures. Hyphal system monomitic in the context and dimitic in the trama, tissue darkening with KOH without hyphal swelling.

Context. Generative hyphae, thin to thick-walled, hyaline to golden yellow, simple septate, rarely branched, $2-5 \,\mu m$ diam.

Trama. Generative hyphae, dominant, thin to thick-walled, hyaline to pale yellow, septate, occasionally branched, $2-4 \mu m$ diam. Skeletal hyphae, thick-walled with narrow to wide lumen, yellowish-brown, aseptate, unbranched, $2-3.5 \mu m$ diam.

Hymenium. Hymenial setae dark brown, thick-walled, ventricose to subulate with sharp to blunt tips, $6-19 \times 3.8-5 \mu m$. Cystidia absent. Cystidioles hyaline, thin walled, ventricose to fusoid with elongated tapering apical portion, $10-45 \times 2-5 \mu m$. Basidia clavate to broadly clavate, $7-15 \times 2.7-6.2 \mu m$, with four sterigmata and a simple septum at the base. Basidioles clavate, $5-13 \times 3.5-6 \mu m$. Basidiospores broadly ellipsoid to subglobose, pale yellow in water, turning golden yellow to brown in KOH, thick-walled, smooth, CB⁻, IKI⁻, (4.5-) 4.7-5.5 (-5.7) \times (3.5-) 3.7-4.5 (-4.7) μm (n = 50/2), Q = 1.13 (Q range 1.05-1.25).

Habitat and distribution. Basidiomes are found on living trees of *Madhuca lon*gifolia and *Prosopis cineraria*, distributed in Cuddalore District, Tamil Nadu, India.

Additional material examined. INDIA, Tamil Nadu, Cuddalore District, Thaiyalkunampattinam, Kanni Tamil Nadu; 11°50'14"N, 79°54'14"E; 31 Dec 2022; Malarvizhi Kaliyaperumal; on a living angiosperm tree (*Prosopis cineraria*); MKDM02a (MUBL1086, Paratype); GenBank: OR272344 (nLSU).

Notes. *Tropicoporus tamilnaduensis* and *T. linteus* are similar in having pileate sulcate basidiomes, zonate context and a mono-dimitic hyphal system



Figure 9. Tropicoporus tamilnaduensis (MUBL1085 holotype) **A** basidiomata (Holotype) **B** pore surface **C** cross-section of basidiome (arrows indicating stratified tube layers) **D** hymenial setae **E**–**H** basidiospores: **E** basidiospore in water **F** basid-iospores in KOH **G** basidiospores in cotton blue **H** basidiospores in Melzer's reagent. Scale bars: 1 cm (**A**–**C**); 5 μm (**D**–**H**).

with cystidioles. However, T. tamilnaduensis differs from T. linteus in deeply-cracked basidiomes and smaller basidiospores (Tian et al. 2013; Wu et al. 2022). Tropicoporus tamilnaduensis and T. rudis share a homogenous context, a mono-dimitic hyphal system and subglobose to broadly ellipsoid basidiospores, but T. tamilnaduensis differs in having zonate, sulcate, deeply irregularlycracked basidiome and larger pores (4–5/mm) (Wu et al. 2022). Tropicoporus tamilnaduensis differs from T. angustisulcatus, T. lineatus and T. substratificans in having sulcate and deeply cracked basidiomes and a mono-dimitic hyphal system, while T. angustisulcatus, T. lineatus and T. substratificans have basidiomes with velutinate to glabrous, uncracked pilear surface and a dimitic hyphal system (Zhou et al. 2015; Wu et al. 2022). Tropicoporus tamilnaduensis resembles T. excentrodendri, T. dependens and T. sideroxylicola by sharing concentrically zonate, sulcate, cracked to rimose pilear surface; however, the former differs by having a mono-dimitic hyphal system (Zhou et al. 2015; Salvador-Montoya et al. 2018). Tropicoporus tamilnaduensis and T. guanacastensis are similar in having a mono-dimitic hyphal system; however, T. tamilnaduensis has ungulate, sulcate, deeply irregularly cracked basidiomes with larger pores (4–7/mm) and broadly ellipsoid to subglobose spores $(4.5-5.7 \times 3.5-4.7 \mu m)$ (Zhou et al. 2015). Although sharing a mono-dimitic hyphal system, T. tamilnaduensis differs from T. flabellatus by having sulcate, deeply irregularly cracked basidiomes, cystidioles and larger pores (4-7/mm) with homogenous context (Lima et al. 2022). Tropicoporus tamilnaduensis



Figure 10. *Tropicoporus tamilnaduensis* (MUBL1085 holotype) **A** tramal hyphae **B** contextual hyphae **C** hymenial setae **D** cystidioles **E** basidioles **F** basidia **G** Basidiospores. Scale bars: 5 µm.

varies from *T. drechsleri* by having sulcate, deeply irregularly cracked basidiomes and broadly ellipsoid to ellipsoid basidiospores $(4.5-5.7 \times 3.5-4.7 \mu m)$ (Salvador-Montoya et al. 2018).

Discussion

Recently, the *Inonotus linteus* complex has gained attention because of its medicinal values and as an emerging potential pathogen in plants (Dai et al. 2009; Dai 2010), humans (Sutton et al. 2005; Haidar et al. 2017; Gupta et al. 2022) and dogs (Hevia et al. 2019). Zhou et al. (2015) segregated the *I. linteus* complex into two new genera viz. *Sanghuangporus* Sheng H. Wu, L.W. Zhou & Y.C. Dai and *Tropicoporus* L.W. Zhou, Y.C. Dai & Shen. H. Wu. Since then, many new species/combinations from tropical countries, especially from China followed by the Americas were introduced (Salvador-Montoya et al. 2018; Brown et al. 2019; Wu et al. 2022). *Tropicoporus* is characterised by its annual to perennial, resupinate, effused-reflexed to pileate basidiomes with mono-dimitic, dimitic hyphal system, ellipsoid to subglobose basidiospores. To date, twenty-three legitimate species are accepted under *Tropicoporus*, of which eleven were from tropical American countries, seven were from East Asian countries and one each from Africa, Costa Rica, Cuba and French Guiana.

The Bayesian phylogram illustrated in the present study is consistent with the previous studies (Coelho et al. 2016; Salvador-Montoya et al. 2018; Brown et al. 2019; Lima et al. 2022). The four new *Tropicoporus* species from Tamil Nadu, India, fit well within the *Tropicoporus* clade but formed a unique, distinct lineage that was the sister clade to *T. rudis* (earlier treated as *Xanthochrous rudis*). The *T. rudis* clade consists of strictly African collections (92% BS, 1.00 BPP) in the phylogeny (Fig. 1). This clade, in turn, forms the sister clade to a clade composed of *T. stratificans*, *T. substratificans* and *T. linteus* with 62% BS, 0.96 BPP.

The Eastern Ghats has discontinuous mountain ranges with hills ranging from 1,100 to 1,600 m with luxuriant vegetation of tropical evergreen to deciduous, thorn forest or scrub jungle that harbours diverse groups of wood rot fungi. This is the first report of the genus *Tropicoporus* from the Eastern Ghats of Tamil Nadu with three novel species, viz. *T. cleistanthicola*, *T. indicus* and *T. pseudoindicus*.

Tropicoporus cleistanthicola, T. tamilnaduensis, T. indicus and T. pseudoindicus are characterised by their perennial, pileate basidiomes with mono-dimitic hyphal system, presence of cystidioles and hymenial setae, smooth, thickwalled, coloured and inamyloid basidiospores (Table 2). However, there is significant variation in their basidiome characteristics, pore (shape and size) and basidiospore (shape and size). *Tropicoporus cleistanthicola* differs from the other three species in having effused-reflexed to pileate, sulcate, and narrowly zonate basidiome with infrequent warts. *Tropicoporus tamilnaduensis* differs in irregularly cracked basidiome with glabrous, sulcate and irregularly cracked, ungulate basidiome and smaller basidiospores ($4.5-5.4 \times 3.5-4.7 \mu m$), while *T. indicus* has regularly cracked and concentrical zonate basidiome, glancing pore surface and larger basidiospores ($5-6 \times 4.2-4.9 \mu m$). Morphologically, *T. pseudoindicus* is similar to *T. indicus* in sharing concentrically zonate, glabrous and rimose with maturity, mono-dimitic hyphal system and presence of cystidioles,

Species	Basidiomata	Context	Margin	Pores /mm	Hyphal system	Setae (in µm)	Cystidioles (in µm)	Basidiospores (in µm)	Q value (Q range)
T. cleistanthicola	Effused-reflexed to pileate, applanate to triquetrous basidiome with narrowly zonate, glabrous, meagrely warted pilear surface	Homogenous	Acute	5–7	Mono-Di	5-32 × 4-5.5	7-45 × 2-5	(4.7-) 4.9-5.2 (-5.4) × (4.2-) 4.5-4.7 (-4.9)	1.1 (1.05–1.2)
T. indicus	Applanate to meagrely triquetrous basidiome with concentrically zonate, sulcate, glabrous, deeply cracked to rimose pilear surface	Homogenous	Acute	4-6	Mono-Di	7-28 × 3-5	5-21 × 3-5	(5-) 5.3-5.8 (- 6) × (4.2-) 4.7-4.9	1.16 (1.05–1.3)
T. pseudoindicus	Applanate, meagrely ungulate to triquetrous basidiome with broadly zonate, distinctly cracked by radial fissures, sulcate pilear surface	Duplex with blackline	Acute to obtuse	6-8	Mono-Di	5–18 × 3–5.5	7-52 × 2.5-5.2	(4-) 4.2-5 (-5.2) × (3.7-) 4-4.5 (-4.7)	1.14 (1.05–1.25)
T. tamilnaduensis	Applanate to meagrely ungulate basidiome with glabrous, broadly zonate, sulcate and deeply irregularly cracked pilear surface	Homogenous	Obtuse	4-5	Mono-Di	6-19 × 3.8-5	10-45 × 2-5	(4.5-) 4.7-5.4 (-5.7) × (3.5-) 3.7-4.5 (-4.7)	1.13 (1.05–1.25)

Table 2. Synoptic comparison of characteristics amongst species of the newly-reported Tropicoporus from India.

while *T. pseudoindicus* differs by having sulcate deeply cracked, radially fissured basidiome, homogenous context and smaller basidiospores. Our Indian *Tropicoporus* species (*Tropicoporus cleistanthicola*, *T. tamilnaduensis*, *T. indicus* and *T. pseudoindicus*) could be easily distinguished by its pileate basidiomes and mono-dimitic hyphal system from the other *Tropicoporus* resupinate species (*T. boehmeriae*, *T. hainanicus*, *T. minus*, *T. ravidus*, *T. stratificans*, *T. tenuis* and *T. texanus*) (Wu et al. 2015; Coelho et al. 2016; Brown et al. 2019; Wu et al. 2022).

Key to species of Tropicoporus in the Afro-Asian region

carps resupinate to effused-reflexed2	1
carps distinctly pileate7	-
carps annual to biennial3	2
carps perennial6	-
spores cyanophilic T. tenuis	3
spores acyanophilic4	-
carp resupinate to effused reflexed, pileal surface tomentose to	4
pasidiospores > 3 μm in length Τ. excentrodendri	
carp resupinate, basidiospores < 3 μm in length 5	_
ments lacerate, context layer present between tube layers	5
ments entire, context layer absent between tub layers	-
carp resupinate, cystidioles present, pores 10–12/mm T. minus	6
carp cushion-shaped, cystidioles absent, pores 8–10/mm	-
system strictly dimitic T. lineatus	7
system mono-dimitic, dimitic in trama8	-

8	Context homogenous9
-	Context duplex with black line
9	Effused reflexed to pileate, uncracked basidiomeT. cleistanthicola
-	Applanate to ungulate or triquetrous, cracked pilear surface10
10	Pores > 6/mm, cyanophilic basidiospores T. rudis
-	Pores < 6/mm, acyanophilic basidiospores11
11	Applanate to triquetrous basidiome with acute velutinate margin, regularly
	cracked pilear surface
-	Applanate to meagrely ungulate basidiome with obtuse margin and deep- ly irregularly cracked pilear surface

Acknowledgements

Malarvizhi Kaliyaperumal and Sugantha Gunaseelan thank EMR-SERB, DST (EMR/2016/003078), Government of India for the financial assistance. MK and GS are grateful to 'The PCCF' of Tamil Nadu Forest Department for providing permission (E2/20458/2017), assistance and support during the field visits in Eastern Ghats of Tamil Nadu. MK, SG and KK thank Prof N. Mathivanan, The Director, Centre for Advanced Studies in Botany, University of Madras, Chennai, for providing the laboratory facilities. Samantha Chandranath Karunarathna thanks the National Natural Science Foundation of China (Numbers NSFC 32260004) and the High-Level Talent Recruitment Plan of Yunnan Provinces ("High-End Foreign Experts" programme) for their support. The authors extend their appreciation to the Researchers Supporting Project number (RSP2024R56), King Saud University, Riyadh, Saudi Arabia.

Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

Funding

EMR-SERB, DST (EMR/2016/003078), Government of India; National Natural Science Foundation of China (Numbers NSFC 32260004). High-Level Talent Recruitment Plan of Yunnan Provinces ("High-End Foreign Experts" programme) and Researchers Supporting Project number (RSP2024R56), King Saud University, Riyadh, Saudi Arabia.

Author contributions

Conceptualisation: MK, SG, KK; Data Curation: MK, SG, KK, SK,EY, CZ, AME, ST; Formal analysis: MK, EY, SK, CZ, AME, ST; Funding acquisition: SG, MK, SK, EY, CZ, AME, ST; Investigation: MK, SG, KK; Methodology: MK, SG, KK; Project administration: MK; Resources: MK, SG, KK; Software: MK, SK, EY, ST; Supervision: MK, SK; Validation MK, SG, KK, SK; Visualisation: MK; Writing – original draft MK, SG, KK; Writing – review & editing MK, SK, EY, CZ, AME, ST.

Author ORCIDs

Sugantha Gunaseelan D https://orcid.org/0000-0001-7089-2292

Data availability

All holotype and paratype collections of the new species are deposited at Madras University Botany Laboratory (MUBL), Centre for Advanced Studies in Botany, University of Madras, Chennai-600 025, Tamil Nadu, India. The sequences generated during this study are deposited in NCBI GenBank. The ITS and nLSU alignment is deposited in TreeBase.

References

- Brown AA, Lawrence DP, Baumgartner K (2019) Role of basidiomycete fungi in the grapevine trunk disease esca. Plant Pathology 69(2): 205–220. https://doi.org/10.1111/ ppa.13116
- Bruen TC, Philippe H, Bryant D (2006) A simple and robust statistical test for detecting the presence of recombination. Genetics 172(4): 2665–2681. https://doi.org/10.1534/genetics.105.048975
- Chen W, Tan H, Liu Q, Zheng X, Zhang H, Liu Y, Xu L (2019) A review: The bioactivities and pharmacological applications of *Phellinus linteus*. Molecules 24(10): e1888. https://doi.org/10.3390/molecules24101888
- Coelho G, Silveira ADO, Antoniolli ZI, Yurchenko E (2016) *Tropicoporus stratificans* sp. nov. (Hymenochaetales, Basidiomycota) from southern Brazil. Phytotaxa 245(2): 144–152. https://doi.org/10.11646/phytotaxa.245.2.5
- Dai YC (2010) Hymenochaetaceae (Basidiomycota) in China. Fungal Diversity 45(1): 131–343. https://doi.org/10.1007/s13225-010-0066-9
- Dai YC, Yang ZL, Cui BK, Yu CJ, Zhou LW (2009) Species diversity and utilization of medicinal mushrooms and fungi in China. International Journal of Medicinal Mushrooms 11(3): 287–302. https://doi.org/10.1615/IntJMedMushr.v11.i3.80 [Review]

Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: More models, new heuristics and parallel computing. Nature Methods 9(8): e772. https://doi.org/10.1038/nmeth.2109

- Doyle JJ, Doyle JL (1987) A rapid isolation procedure for small quantities of fresh tissue. Phytochemical Bulletin. Botanical Society of America 19: 11–15.
- Edler D, Klein J, Antonelli A, Silvestro D (2020) raxmlGUI 2.0: A graphical interface and toolkit for phylogenetic analyses using RAxML. Methods in Ecology and Evolution 12(2): 373–377. https://doi.org/10.1111/2041-210X.13512
- Góes-Neto A, Loguercio-Leite C, Guerrero RT (2005) DNA extraction from frozen field-collected and dehydrated herbarium fungal basidiome: Performance of SDS and CTAB-based methods. Biotemas 18: 19–32. https://periodicos.ufsc.br/index.php/biotemas/article/download/21410/19377/68437
- Guindon S, Gascuel O (2003) A simple, fast and accurate algorithm to estimate large phylogenies by maximum likelihood. Systematic Biology 52(5): 696–704. https://doi. org/10.1080/10635150390235520
- Gupta P, Kaur H, Dwivedi S, Agnihotri S, Rudramurthy SM (2022) First case of *Tropicoporus tropicalis* keratitis in an immunocompetent host from India and review of the

literature. Journal de Mycologie Médicale 32(1): e101205. https://doi.org/10.1016/j. mycmed.2021.101205

- Haidar G, Zerbe CS, Cheng M, Zelazny AM, Holland SM, Sheridan KR (2017) *Phellinus* species: An emerging cause of refractory fungal infections in patients with X-linked chronic granulomatous disease. Mycoses 60(3): 155–160. https://doi.org/10.1111/ myc.12573
- Hevia A, Iachini R, Fernández J, Lazzari J, Suárez-Alvarez R, Abrantes R, Toranzo A, Refojo N, Canteros C (2019) Mycosis due to *Tropicoporus tropicalis* (= *Inonotus tropicalis*) in a domestic dog. Mycopathologia 184(5): 701–706. https://doi.org/10.1007/s11046-019-00368-1
- Huson DH, Bryant D (2006) Application of phylogenetic networks in evolutionary studies. Molecular Biology and Evolution 23(2): 254–267. https://doi.org/10.1093/molbev/msj030
- Kaur R, Kaur M, Singh AP, Ghuman NK, Dhingra GS (2022) Diversity of some colourful poroid and non-poroid Agaricomycetous fungi. In: Rajpal VR, Singh I, Navi SS (Eds) Fungal Diversity, Ecology and Control Management Fungal Biology. Springer, Singapore, 199–254. https://doi.org/10.1007/978-981-16-8877-5_11
- Kornerup A, Wanscher JH (1978) Methuen Handbook of Colour (3rd edn.). Eyre Methuen, London.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular evolutionary genetics analysis across computing platforms. Molecular Biology and Evolution 35(6): 1547–1549. https://doi.org/10.1093/molbev/msy096
- Lima VX, de Oliveira VRT, de Lima-Júnior NC, Oliveira-Filho JRC, Santos C, Lima N, Gibertoni TB (2022) Taxonomy and phylogenetic analysis reveal one new genus and three new species in *Inonotus s.l.* (Hymenochaetaceae) from Brazil. Cryptogamie. Mycologie 43(1): 1–21. https://doi.org/10.5252/cryptogamie-mycologie2022v43a1
- Natarajan K, Kolandavelu K (1998) Resupinate Aphyllophorales of Tamil Nadu, India. CAS in Botany, University of Madras. https://koeltz.com/en/resupinate-aphyllophoralesof-tamilnadu-india-1998-133-p
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029
- Salvador-Montoya CA, Costa-Rezende DH, Ferreira-Lopes V, Borba-Silva MA, Popoff OF (2018) Tropicoporus drechsleri (Hymenochaetales, Basidiomycota), a new species in the "Inonotus linteus" complex from Northern Argentina. Phytotaxa 338: 075–089. https://doi.org/10.11646/phytotaxa.338.1.6
- Sayers ER, Bolton EE, Brister JR, Canese K, Chan J, Comeau DC, Farrell CM, Feldgarden M, Fine AM, Funk K, Hatcher E, Kannan S, Kelly C, Kim S, Klimke W, Landrum MJ, Lathrop S, Lu Z, Madden TM, Malheiro A, Marchler-Bauer A, Murphy TD, Phan L, Pujar S, Rangwala SH, Schneider VA, Tse T, Wang J, Ye J, Trawick BW, Kim D, Pruitt KD, Sherry ST (2023) Database resources of the National Centre for Biotechnology Information in 2023. Nucleic Acids Research 51(D1): D29–D38. https://doi.org/10.1093/ nar/gkac1032
- Sutton DA, Thompson EH, Rinaldi MG, Iwen PC, Nakasone KK, Jung HS, Rosenblatt HM, Paul ME (2005) Identification and first report of *Inonotus (Phellinus) tropicalis* as an etiologic agent in a patient with chronic granulomatous disease. Journal of Clinical Microbiology 43(2): 982–987. https://doi.org/10.1128/JCM.43.2.982-987.2005

- Tian XM, Yu HY, Zhou LW, Decock C, Vlasák J, Dai YC (2013) Phylogeny and taxonomy of the *Inonotus linteus* complex. Fungal Diversity 58(1): 159–169. https://doi. org/10.1007/s13225-012-0202-9
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. Journal of Bacteriology 172(8): 4238–4246. https://doi.org/10.1128/jb.172.8.4238-4246.1990
- Vlasák J, Li HJ, Zhou LW, Dai YC (2013) A further study on *Inonotus linteus* complex (Hymenochaetales, Basidiomycota) in tropical America. Phytotaxa 124(1): 25–36. https://doi.org/10.11646/phytotaxa.124.1.3
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR Protocols: A Guide to Methods and Applications. Academic Press, San Diego, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1
- Wu F, Qin WM, Euatrakool O, Zhou LW (2015) Tropicoporus boehmeriae sp. nov. (Hymenochaetaceae, Basidiomycota) from Thailand, a new member of the Inonotus linteus complex. Phytotaxa 231(1): 73–80. https://doi.org/10.11646/phytotaxa.231.1.7
- Wu F, Zhou LW, Vlasák J, Dai YC (2022) Global diversity and systematics of Hymenochaetaceae with poroid hymenophore. Fungal Diversity 113(1): 1–192. https://doi. org/10.1007/s13225-021-00496-4
- Zhou LW, Vlasák J, Decock C, Assefa A, Stenlid J, Abate D, Wu SH, Dai YC (2015) Global diversity and taxonomy of the *Inonotus linteus* complex (Hymenochaetales, Basidiomycota): Sanghuangporus gen. nov., *Tropicoporus excentrodendri* and *T. guanacastensis* gen. et spp. nov., and 17 new combinations. Fungal Diversity 77(1): 335–347. https://doi.org/10.1007/s13225-015-0335-8

Supplementary material 1

Pairwise distance matrix, based on nucleotide sequences of four new *Tropicoporus* spp. and its related species

- Authors: Sugantha Gunaseelan, Kezhocuyi Kezo, Samantha C. Karunarathna, Erfu Yang, Changlin Zhao, Abdallah M. Elgorban, Saowaluck Tibpromma, Malarvizhi Kaliyaperumal Data type: xls
- Explanation note: Pairwise distance matrix, based on nucleotide sequences of four new *Tropicoporus* spp. and its related species (Pairwise distances calculations were accomplished using MEGA X v.10.0.2. Distances and standard errors are respectively displayed in the lower-left matrix and the upper-right matrix).
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/mycokeys.102.117067.suppl1

Supplementary material 2

Molecular Phylogeny of four new Indian *Tropicoporus* species inferred from ITS sequences

Authors: Sugantha Gunaseelan, Kezhocuyi Kezo, Samantha C. Karunarathna, Erfu Yang, Changlin Zhao, Abdallah M. Elgorban, Saowaluck Tibpromma, Malarvizhi Kaliyaperumal Data type: jpg

- Explanation note: The topology is from Bayesian analysis. Bootstrap values and Bayesian posterior probabilities, equal to or above 60% and 0.90, respectively, are labelled at the nodes. The newly-generated sequences are coloured and bold, and the type specimens are in bold.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/mycokeys.102.117067.suppl2

Supplementary material 3

Molecular Phylogeny of four new Indian *Tropicoporus* species inferred from nLSU sequences

Authors: Sugantha Gunaseelan, Kezhocuyi Kezo, Samantha C. Karunarathna, Erfu Yang, Changlin Zhao, Abdallah M. Elgorban, Saowaluck Tibpromma, Malarvizhi Kaliyaperumal Data type: jpg

- Explanation note: The topology is from Bayesian analysis. Bootstrap values and Bayesian posterior probabilities, equal to or above 60% and 0.90, respectively, are labelled at the nodes. The newly-generated sequences are coloured and bold and the type specimens are in bold.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/mycokeys.102.117067.suppl3