

Research Article

Ceriporiopsis tianshanensis (Polyporales, Agaricomycetes) and *Sidera tianshanensis* (Hymenochaetales, Agaricomycetes), two new species of wood-inhabiting fungi from Xinjiang, Northwest China

Tai-Min Xu¹⁰, Yi-Fei Sun¹⁰, Shun Liu¹⁰, Chang-Ge Song¹⁰, Neng Gao²⁰, Dong-Mei Wu²⁰, Bao-Kai Cui¹⁰

1 Institute of Microbiology, School of Ecology and Nature Conservation, Beijing Forestry University, Beijing 100083, China

2 Xinjiang Production and Construction Group Key Laboratory of Crop Germplasm Enhancement and Gene Resources Utilization, Biotechnology Research Institute, Xinjiang Academy of Agricultural and Reclamation Sciences, Shihezi, Xinjiang 832000, China

Corresponding authors: Dong-Mei Wu (wdm0999123@sina.com); Bao-Kai Cui (cuibaokai@bjfu.edu.cn)

Abstract

Wood-inhabiting fungi are abundant in China, but their distribution is uneven, with more fungi in southwest China and fewer fungi in northwest China. During the investigation of wood-inhabiting fungi in Xinjiang, we collected a large number of specimens. Eight specimens growing on *Picea schrenkiana* were collected from Tianshan Mountains, and they were described as two new species in *Ceriporiopsis* and *Sidera* based on morphological characters and molecular evidence. *Ceriporiopsis tianshanensis* is characterized by a cream to salmon-buff pore surface, larger pores measuring 1–3 per mm, and broadly ellipsoid basidiospores $5-6.5 \times 3-4 \mu m$. *Sidera tianshanensis* is characterized by annual to perennial basidiocarps, measuring 15 mm thick, pores 5-7 per mm, cream to rosy buff pore surface, and allantoid basidiospores $3-3.5 \times 1-1.4 \mu m$. Detailed illustrations and descriptions of the novel species are provided.

Key words: macrofungi, phylogeny, polyporoid fungi, taxonomy, white-rot fungi

Introduction

China is rich in wood-inhabiting fungal resources, and more than 2000 species of the woody fungi have been reported (Dai 2010, 2012; Cui et al. 2019; Wu et al. 2022a, b). In the past ten years, many new species of the wood-inhabiting fungi have been discovered in China, and mainly distributed in the southwest and south areas, and few new species have been published from northwest China (Li et al. 2014; Chen et al. 2016; Shen et al. 2019; Wang et al. 2021; Yuan et al. 2021; Ji et al. 2022; Wu et al. 2022b; Liu et al. 2023a).

The Xinjiang Uygur Autonomous Region is located in northwestern China, and, as the largest province in China, it covers an area of 1,664,900 square kilometers. There is a typical temperate continental arid climate, with an extremely uneven distribution of water resources in time and space, more in the west and less in the east, more in the north and less in the south, more in the mountains and less in the plains (Wu et al. 2010; Hu et al. 2021). Due to severe climatic conditions, natural forests are mainly distributed in the Tianshan Mountains



Academic editor: Thorsten Lumbsch Received: 23 February 2023 Accepted: 2 May 2023 Published: 29 May 2023

Citation: Xu T-M, Sun Y-F, Liu S, Song C-G, Gao N, Wu D-M, Cui B-K (2023) *Ceriporiopsis tianshanensis* (Polyporales, Agaricomycetes) and *Sidera tianshanensis* (Hymenochaetales, Agaricomycetes), two new species of wood-inhabiting fungi from Xinjiang, Northwest China. MycoKeys 98: 1–18, https://doi. org/10.3897/mycokeys.98.102552

Copyright: © Tai-Min Xu et al. This is an open access article distributed under terms of the Creative Commons Attribution License (Attribution 4.0 International – CC BY 4.0). and Altai Mountains (Zhang and Zhang 2014; Huang et al. 2018). In the past, 592 species of macrofungi have been reported in Xinjiang, among which 243 are species of wood rot fungi, and most of them were distributed in the Tianshan Mountains and the Altai Mountains (Wang and Ayinuer 2004; Dai et al. 2007; Bau et al. 2008; Guli et al. 2015; Zhao 2022). In recent years, some new species of wood rot fungi have been discovered in Xinjiang: *Fomitopsis tianshanensis* B.K. Cui & Shun Liu, *Laetiporus xinjiangensis* J. Song, Y.C. Dai & B.K. Cui, *Porodaedalea schrenkianae* Y.C. Dai & F. Wu, and *Rhodonia tianshanensis* Yuan Yuan & L.L. Shen (Song et al. 2014; Yuan and Shen 2017; Liu et al. 2021a; Wu et al. 2022b).

During the investigation of wood rot fungi in Xinjiang, we collected a large number of specimens, including two belonging to Ceriporiopsis and six belonging to Sidera. The genus Ceriporiopsis Domański (Meruliaceae, Polyporales) was erected by Domański (1963) based on the morphological analyses to accommodate C. gilvescens (Bres.) Domański (type species), C. incarnata Domański, C. resinascens (Romell) Domański, C. aneirina (Sommerf.) Domańsk and C. placenta (Fr.) Domański. Currently, there are 41 species accepted in Ceriporiopsis, and eight species recorded in China: C. albonigrescens Núñez, Parmasto & Ryvarden, C. aurantitingens (Corner) T. Hatt., C. egula C.J. Yu & Y.C. Dai, C. lavendula B.K. Cui, C. micropora T.T. Chang & W.N. Chou, C. mucida (Pers.) Gilb. & Ryvarden, C. subrufa (Ellis & Dearn.) Ginns and C. subsphaerospora (A. David) M. Pieri & B. Rivoire (Zhao and Cui 2014; Zhao et al. 2015, 2023). The genus causes a white rot on angiosperms and gymnosperms (Niemelä 1985; Zhao and Cui 2014; Zhao et al. 2015; Spirin and Ryvarden 2016). It is characterized by annual, resupinate to effused-reflexed basidiocarps, a monomitic hyphal system with no action in Melzer's reagent or Cotton Blue, generative hyphae with clamp connections, and subcylindrical to ellipsoid basidiospores whih hyaline, thin walls (Gilbertson and Ryvarden 1987; Núñez and Ryvarden 2001; Ryvarden and Melo 2014; Zhao and Wu 2017). In phylogenetic analysis, Ceriporiopsis was polyphyletic and clustered into the phlebioid clade (Zhao and Cui 2014; Zhao et al. 2015; Zhao and Wu 2017). Zmitrovich (2018) transferred C. gilvescens and C. kunmingensis to the genus Mycoacia Donk (Zmitrovich 2018). Zhao et al. (2023) conducted a detailed phylogenetic analysis, and many species within Ceriporiopsis were placed in the genera Ceriporiopsoides C.L. Zhao, Hydnophlebia Parmasto, and Phlebicolorata C.L. Zhao. The remaining Ceriporiopsis species did not belong to the phlebioid clade but were grouped in the residual polyporoid clade and formed a relatively stable branch cluster. The genus Sidera Miettinen & K.H. Larss. (Rickenellaceae, Hymenochaetales) was established by Miettinen and Larsson (2011) based on phylogenetic and morphological analyses to accommodate S. lunata (Romell ex Bourdot & Galzin) K.H. Larss., S. lowei (Rajchenb.) Miettinen, S. lenis (P. Karst.) Miettinen (type species) and S. vulgaris (Fr.) Miettinen. To date, 18 species are accepted in Sidera, nine species were recorded in China: S. borealis Z.B. Liu & Yuan Yuan, S. inflata Z.B. Liu & Y.C. Dai, S. lenis, S. minutissima Y.C. Dai, F. Wu, G.M. Gates & Rui Du, S. parallela Y.C. Dai, F. Wu, G.M. Gates & Rui Du, S. punctata Z.B. Liu & Y.C. Dai, S. roseobubalina Z.B. Liu & Y.C. Dai, S. salmonea Z.B. Liu, Jian Yu & F. Wu, S. tibetica Z.B. Liu, Jian Yu & F. Wu (Liu et al. 2023b). The genus causes a white rot in the wood, and is characterized by resupinate basidiocarps that are white to cream or buff, mostly waxy when fresh, with a poroid or hydnoid hymenophore, monomitic or

dimitic hyphal system with generative hyphae bearing clamp connections, the presence of rosette-like crystals and allantoid to lunate basidiospores (Miettinen and Larsson 2011; Du et al. 2020; Liu et al. 2021b, 2022). In phylogenetic analysis, *Sidera* is a monophyletic genus and clustered into the *Rickenella* clade (Liu et al. 2021b, 2022, 2023b). In this study, two new species are described based on morphological and phylogenetic evidence.

Materials and methods

Morphological studies

The specimens used in this study were deposited at the herbarium of the Institute of Microbiology, Beijing Forestry University, China (BJFC). Macro-morphological descriptions were based on field notes and laboratory measurements. The microscopic routines used in this study followed Cui et al. (2019) and Liu et al. (2023a). Sections were studied at a magnification up to ×1000 using a Nikon E80i microscope and phase contrast illumination (Nikon, Tokyo, Japan). Line drawings were made with the aid of a drawing tube. Microscopic features, measurements and drawings were made from slide preparations of dried or fresh material stained with Cotton Blue and Melzer's reagent, as described by Dai (2010). To represent the variation in the size of the basidiospores, 5% of measurements were excluded from each end of the range and are given in parentheses. The following abbreviations were used: IKI = Melzer's reagent, IKI-= neither dextrinoid nor amyloid, KOH = 5% potassium hydroxide, CB = Cotton Blue, CB-= acyanophilous, L = mean spore length (arithmetic average of all spores), W = mean spore width (arithmetic average of all spores), Q = variation in the L/W ratios between the specimens studied, n = number of spores measured from a given number of specimens. Color terms followed Petersen (1996).

DNA extraction and sequencing

Total genomic DNA was extracted from dried specimens using a cetyltrime-thylammonium bromide (CTAB) Rapid Plant Genome Extraction Kit (Aidlab Biotech-nologies Company, Ltd., Beijing, China) according to the manufacturer's instructions with some modifications (Li et al. 2014; Ji et al. 2022). Two DNA gene fragments, ITS and nLSU, were amplified using the primer pairs ITS5/ITS4 and LR0R/LR7 (White et al. 1990). The PCR procedures for ITS and nLSU followed Song et al. (2022) and Sun et al. (2022) in the phylogenetic analyses. All PCR products were directly purified and sequenced at the Beijing Genomics Institute (BGI), China, with the same primers. Newly generated sequences were submitted to GenBank and are listed in Tables 1, 2.

Phylogenetic analysis

Phylogenetic analyses for *Ceriporiopsis* and *Sidera* were performed with maximum parsimony (MP), maximum likelihood (ML) and Bayesian inference (BI) analyses based on the combined ITS+nLSU dataset. New generated sequences were aligned with the additional sequences retrieved from GenBank (Tables 1, 2) using BioEdit 7.0.5.3 (Hall 1999) and ClustalX 1.83 (Thompson et al. 1997),

Species	Sample no.	Location	GenBank accession no.		Deference
			ITS	nLSU	Reference
Ceriporiopsis andreanae	CBS 279.92	USA, Montana	ALYI01000630	_	Cheng et al. 2022
C. herbicola	K 132752	UK, Oxfordshire	KX008364	KX081076	Zhao and Wu 2017
C. pseudogilvescens	Niemelä 7447	Finland	FJ496680	FJ496700	Tomšovský et al. 201
C. pseudogilvescens	TAA 168233	Estonia	FJ496673	FJ496702	Tomšovský et al. 201
C. pseudogilvescens	BRNM 686416	Slovakia	FJ496679	FJ496703	Tomšovský et al. 201
C. subrufa	BRNM 710164	Czech Republic	FJ496661	FJ496723	Tomšovský et al. 201
C. subrufa	BRNM 710172	Czech Republic	FJ496662	FJ496724	Tomšovský et al. 201
C. tianshanensis	Cui 19150	China, Xinjiang	OP920992	OP920984	Present study
C. tianshanensis	Cui 19151	China, Xinjiang	OP920993	OP920985	Present study
Ceriporiopsoides guidella	HUBO 7659	Italy	FJ496687	FJ496722	Tomšovský et al. 201
C. lagerheimii	58240	Ecuador, Napo	KX008365	KX081077	Zhao and Wu 2017
Hydnophlebia fimbriata	Dai 11672	China, Hunan	KJ698633	KJ698637	Zhao et al. 2015
H. fimbriata	Cui 1671	China, Jiangsu	KJ698634	KJ698638	Zhao et al. 2015
Mycoacia gilvescens	BRNM 710166	Czech Republic	FJ496684	FJ496720	Tomšovský et al. 201
M. gilvescens	Yuan 2752	China, Shaanxi	KF845953	KF845946	Zhao and Cui 2014
M. gilvescens	BRNM 667882	Czech Republic	FJ496685	FJ496719	Tomšovský et al. 201
M. kunmingensis	C.L. Zhao 152	China, Yunnan	KX081072	KX081074	Zhao and Wu 2017
M. kunmingensis	C.L. Zhao 153	China, Yunnan	KX081073	KX081075	Zhao and Wu 2017
Phlebicolorata alboaurantia	Cui 2877	China, Fujian	KF845954	KF845947	Zhao and Cui 2014
P. alboaurantia	Cui 4136	China, Fujian	KF845948	KF845955	Zhao and Cui 2014
P. pseudoplacenta	JV 050952	USA, Tennessee	JN592499	JN592506	Vlasák et al. 2012
P. pseudoplacenta	PRM 899297	USA	JN592497	JN592504	Vlasák et al. 2012
P. rosea	Dai 13573	China, Yunnan	KJ698635	KJ698639	Zhao et al. 2015
P. rosea	Dai 13584	China, Yunnan	KJ698636	KJ698640	Zhao et al. 2015
P. semisupina	Cui 10222	China, Zhejiang	KF845949	KF845956	Zhao and Cui 2014
P. semisupina	Cui 10189	China, Zhejiang	KF845958	KF845951	Zhao and Cui 2014
P. semisupina	Cui 7971	China, Yunnan	KF845950	KF845957	Zhao and Cui 2014
Raduliporus aneirinus	Dai 12657	Finland, Helsinki	KF845952	KF845945	Zhao and Cui 2014
Antrodia serpens	Dai 7465	Luxemburg	KR605813	KR605752	Liu et al. 2023a
Rhodofomes roseus	Cui 17046	China, Yunnan	ON417187	ON417238	Liu et al. 2023a

Table 1. List of species, specimens and GenBank accession numbers of sequences used in the phylogeny of Ceriporiopsis.

followed by manual adjustments. *Antrodia serpens* (Fr.) P. Karst. and *Rhodofomes roseus* (Alb. & Schwein.) Kotl. & Pouzar were used as outgroups in the phylogeny of *Ceriporiopsis* (Zhao and Wu 2017), while *Skvortzovia furfurella* (Bres.) Bononi & Hjortstam and *Skvortzovia furfuracea* (Bres.) G. Gruhn & Hallenberg were used as outgroups in the phylogeny of *Sidera* (Liu et al. 2022).

Maximum parsimony (MP) analysis was performed in PAUP* version 4.0b10 (Swofford 2002). The settings for phylogenetic analyses in this study followed the approach of Ji et al. (2022) and Zhu et al. (2019). All characters were equally weighted, and gaps were treated as missing data. Trees were inferred using the heuristic search option with TBR branch swapping and 1000 random sequence additions. Max trees were set to 5000, branches of

Species	Sample no.	Location	GenBank ad	ccession no.	Reference
			ITS	nLSU	
Sidera inflata	Cui 13610	China, Hainan	MW198480	-	Liu et al. 2021b
S. lenis	Miettinen 11036	Finland	FN907914	FN907914	Miettinen and Larsson 2011
S. lunata	JS 15063	Norway	DQ873593	DQ873593	Miettinen and Larsson 2011
S. malaysiana	Dai 18570	Malaysia	MW198481	MW192007	Liu et al. 2021b
S. minutipora	Gates FF257	Australia	FN907922	FN907922	Miettinen and Larsson 2011
S. minutipora	Cui 16720	Australia	MN621349	MN621348	Du et al. 2020
S. minutissima	Dai 18471A	China, Hainan	MW198482	MW192008	Liu et al. 2021b
S. minutissima	Dai 19529	Sri Lanka	MN621352	MN621350	Du et al. 2020
S. parallela	Cui 10346	China, Yunnan	MK346145	-	Du et al. 2020
S. parallela	Cui 10361	China, Yunnan	MK346144	-	Du et al. 2020
S. punctata	Dai 22119	China, Hainan	MW418438	MW418437	Liu et al. 2021b
S. roseo-bubalina	Dai 11277	China, Henan	MW198483	-	Liu et al. 2021b
S. salmonea	Dai 23354	China, Tibet	OM974250	OM974242	Liu et al. 2022
S. salmonea	Dai 23343	China, Tibet	OM974249	OM974241	Liu et al. 2022
S. salmonea	Dai 23428	China, Tibet	OM974251	OM974243	Liu et al. 2022
S. tianshanensis	Cui 19132	China, Xinjiang	OP920994	OP920986	Present study
S. tianshanensis	Cui 19143	China, Xinjiang	OP920995	OP920987	Present study
S. tianshanensis	Cui 19186	China, Xinjiang	OP920996	OP920988	Present study
S. tianshanensis	Cui 19192	China, Xinjiang	OP920997	OP920989	Present study
S. tianshanensis	Cui 19196	China, Xinjiang	OP920998	OP920990	Present study
S. tianshanensis	Cui 19251	China, Xinjiang	OP920999	OP920991	Present study
S. srilankensis	Dai 19654	Sri Lanka	MN621344	MN621346	Du et al. 2020
S. srilankensis	Dai 19581	Sri Lanka	MN621345	MN621347	Du et al. 2020
S. tenuis	Dai 18698	Australia	MK331866	MK331868	Du et al. 2020
S. tenuis	Dai 18697	Australia	MK331865	MK331867	Du et al. 2020
S. tibetica	Dai 23648	China, Tibet	OM974253	OM974245	Liu et al. 2022
S. tibetica	Dai 23407	China, Tibet	OM974252	OM974244	Liu et al. 2022
S. tibetica	Dai 21057	Belarus	MW198484	MW192009	Liu et al. 2021b
S. tibetica	Dai 22151	China, Guangxi	MW477794	MW474965	Liu et al. 2021b
S. vesiculosa	BJFC025367	Singapore	MH636565	MH636567	Du et al. 2020
S. vesiculosa	BJFC025377	Singapore	MH636564	MH636566	Du et al. 2020
S. borealis	Cui 11216	China, Shaanxi	MW198485	-	Liu et al. 2021b
S. vulgaris	Ryvarden 37198	New Zealand	FN907918	FN907918	Miettinen and Larsson 2011
S. lowei	Miettinen X419	Venezuela	FN907917	FN907917	Miettinen and Larsson 2011
S. lowei	Miettinen X426	New Zealand	FN907919	FN907919	Miettinen and Larsson 2011
Skvortzovia furfuraceum	KHL 11738	Finland	DQ873648	DQ873648	Liu et al. 2022
S. furfurella	KHL 10180	Puerto Rico	DQ873649	DQ873649	Liu et al. 2022

Table 2. List of species, specimens and GenBank accession numbers of sequences used in the phylogeny of Sidera.

zero length were collapsed, and all parsimonious trees were saved. Clade robustness was assessed using a bootstrap (BT) analysis with 1000 replicates (Felsenstein 1985). Descriptive tree statistics tree length (TL), consistency index (CI), retention index (RI), rescaled consistency index (RC) and homoplasy index (HI) were calculated for each generated Maximum Parsimonious Tree (MPT) (Page 1996).

Maximum likelihood (ML) analysis was conducted by RAxML-HPC252 through the CIPRES Science Gateway (www.phylo.org) and involved 100 ML searches. All model parameters were estimated by the program. Only the best maximum likelihood tree from all searches was retained. The maximum likelihood bootstrap values (ML-BS) were determined using rapid bootstrapping with 1000 replicates. The phylogenetic tree was visualized using Treeview (Page 1996).

Bayesian inference (BI) analysis was implemented in MrBayes 3.2.6 (Ronquist et al. 2012). There were two independent runs, each of which had four chains for 1,000,000 generations sampling from the posterior distribution every 1000th generation to check that the PSRF (potential scale reduction factors) were reasonably close to 1.0 for all parameters indicative of chain convergence. The first 25% of the sampled trees were discarded as burn-in, while the remaining trees were used to obtain the Bayesian posterior probabilities (BPPs) of the clades. A majority rule consensus tree of all remaining trees was calculated.

Branches that received bootstrap support for maximum parsimony (MP), maximum likelihood (ML) higher than or equal to 75% (MP and ML-BS) and Bayesian posterior probabilities (BPP) higher than or equal to 0.95 (BPP) were considered significantly supported. The best topologies from MP analyses are shown in this study, and the final alignments and the retrieved topologies were deposited in TreeBASE (http://www.treebase.org accessed on 28 April 2023), under accession ID: 29931.

Results

Molecular phylogeny

The phylogeny of *Ceriporiopsis*, based on a combined ITS and nLSU dataset, included 30 ITS sequences and 29 nLSU sequences from 30 fungal specimens, representing 17 species. The dataset had an aligned length of 2153 characters, of which 1399 characters were constant, 200 were variable and parsimony-uninformative and 554 were parsimony informative. Maximum parsimony analysis yielded one equally parsimonious tree (TL = 1902, CI = 0.601, RI = 0.763, RC = 0.459, HI = 0.399), and a strict consensus tree of these trees is shown in Fig. 1. The best model fit applied in the Bayesian inference analysis was GTR+I+G. Bayesian analysis and ML analysis resulted in a similar topology to MP analysis, with an average standard deviation of split frequencies of 0.006591 (BI).

The phylogeny of *Sidera*, based on a combined ITS and nLSU dataset, included 37 ITS sequences and 32 nLSU sequences from 37 fungal specimens, representing 19 species. The dataset had an aligned length of 2235 characters, of which 1453 characters were constant, 205 were variable and parsimony-uninformative and 577 were parsimony in-formative. Maximum parsimony analysis yielded one equally parsimonious tree (TL = 2233, CI = 0.583, RI = 0.760,

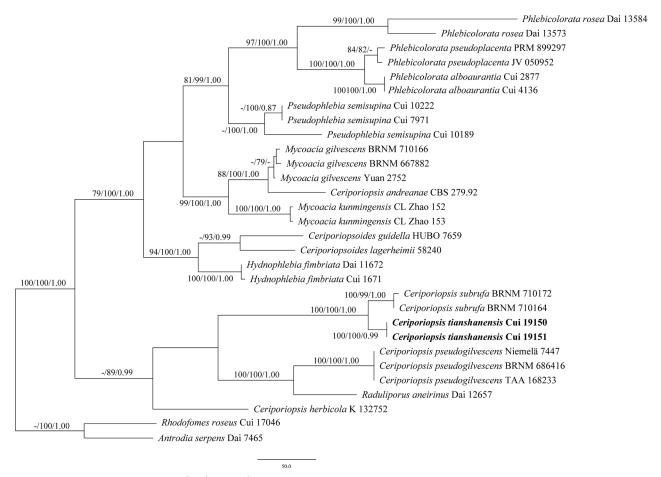


Figure 1. Maximum parsimony (MP) tree of *Ceriporiopsis* based on the combined ITS+nLSU dataset. Branches are labelled with maximum parsimony/maximum likelihood bootstrap values higher than 75% and Bayesian posterior probability values greater than 0.95. The new species is indicated in bold.

RC = 0.443, HI = 0.417), and a strict consensus tree of these trees is shown in Fig. 2. The best model fit applied in the Bayesian inference analysis was GTR+I+G. Bayesian analysis and ML analysis resulted in a similar topology as MP analysis, with an average standard deviation of split frequencies of 0.007516 (BI).

Within the phylogenetic tree of *Ceriporiopsis*, the new species *C. tianshanensis* was closely related to *C. subrufa* with high supports (100% ML, 100% MP, 1.00 BPP; Fig. 1). However, the ITS sequences of *Ceriporiopsis tianshanensis* and *C. subrufa* were significantly different, with 31 different nucleobases, and the similarity was 94.80% by nucleotide blast. The difference in the nLSU sequence was not significant; there were 4 different nucleobases, and the similarity was 99.29% by nucleotide blast.

In addition, the phylogenetic tree of *Sidera*, the new species *Sidera tianshanensis*, was closely related to *S. salmonea* with high support (100% ML, 93% MP, 1.00 BPP; Fig. 2). However, the ITS sequences of *Sidera tianshanensis* and *S. salmonea* were significantly different, with 40 different nucleobases, and the similarity was 94.29% by nucleotide blast. The difference in the nLSU sequence was not significant; there were 7 different nucleobases, and the similarity was 99.55% by nucleotide blast.

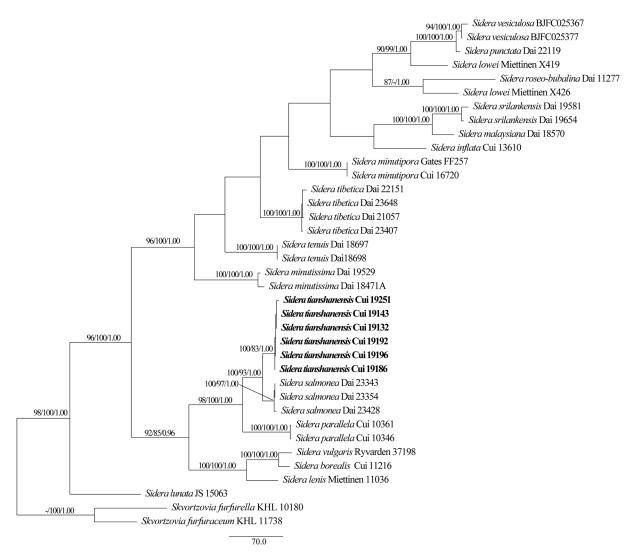


Figure 2. Maximum parsimony (MP) tree of *Sidera* based on the combined ITS+nLSU dataset. Branches are labelled with maximum parsimony/maximum likelihood bootstrap values higher than 75% and Bayesian posterior probabilities greater than 0.95.

Taxonomy

Ceriporiopsis tianshanensis B.K. Cui & T.M. Xu, sp. nov.

MycoBank No: 848610 Figs 3, 4

Diagnosis. Ceriporiopsis tianshanensis is characterized by a cream to salmon-buff pore surface when fresh, large pores measuring 1–3 per mm, broadly ellipsoid basidiospores measuring 5–6.5 × 3–4 μ m, and growth on the stump of *Picea schrenkiana* Fisch. et Mey.

Type. CHINA. Xinjiang Autonomous Region, Tekes County, Kosang Cave National Forest Park, on the stump of *Picea schrenkiana*, 19 September 2021, Cui 19150 (holotype).

Etymology. tianshanensis (Lat.): referring to the species occurrence in Tianshan.



Figure 3. Basidiocarps of Ceriporiopsis tianshanensis (Cui 19151). Scale bar: 1.0 cm.

Fruiting body. Basidiocarps annual, resupinate, adnate, not easily separated from the substrate, soft corky when fresh, fragile to hard fibrous when dry, up to 12 cm long, 3 cm wide, 2 mm thick. Pore surface white to cream or salmon-buff when fresh, becoming buff to vinaceous-buff or fawn when dry; pores irregular, 1-3 per mm; dissepiments thin, entire. Subiculum cream to buff and fibrous to soft corky when dry, up to 4 mm thick. Tubes concolorous with pore surface, corky, up to 4 mm long.

Hyphal structure. Hyphal system monomitic; generative hyphae with clamp connections, lack crystal, IKI–, CB–; tissues unchanged in KOH.

Subiculum. Generative hyphae hyaline, thin- to slightly thick-walled, often branched, interwoven, $3.5-5 \,\mu$ m in diameter.

Tubes. Generative hyphae hyaline, thin- to slightly thick-walled, occasionally branched, interwoven, $3-6 \mu m$ in diameter. Cystidia and other sterile hymenial elements absent. Basidia short clavate to barrel-shaped, bearing four sterigmata and a basal clamp connection, $12-22 \times 5-6 \mu m$; basidioles dominant, in shape similar to basidia, but smaller.

Spores. Basidiospores broadly ellipsoid, colorless, thin-walled, smooth, often with one guttule, IKI-, CB-, $5-6.5 \times 3-4 \mu m$, L = 5.9 μm , W = 3.5 μm , Q = 1.69-1.74 (n = 60/2).

Type of rot. White rot.

Additional specimen (paratype) examined. CHINA. Xinjiang Autonomous Region, Tekes County, Kosang Cave National Forest Park, on the stump of *Picea* schrenkiana, 19 September 2021, Cui 19151.

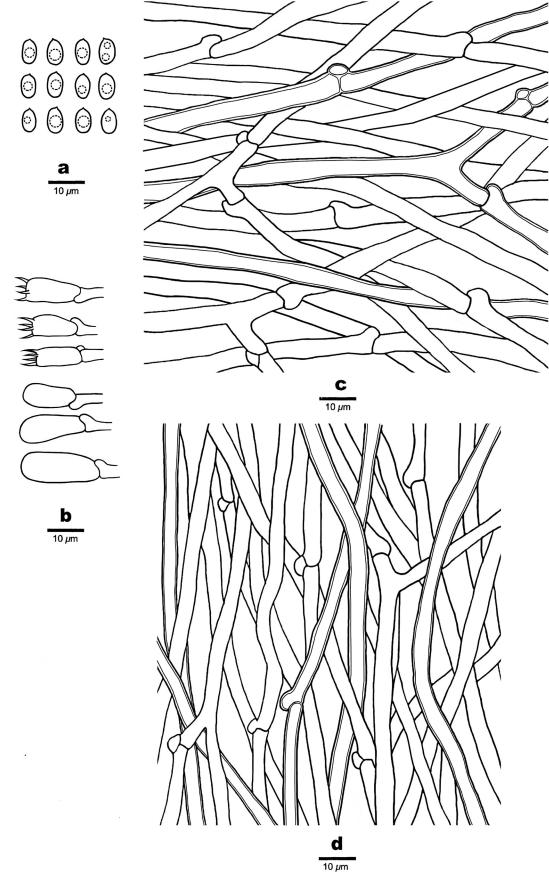


Figure 4. Microscopic structures of *Ceriporiopsis tianshanensis* (Cui 19150) **a** basidiospores **b** basidia and basidioles **c** hyphae from the subiculum **d** hyphae from trama.

Sidera tianshanensis B.K. Cui & T.M. Xu, sp. nov.

MycoBank No: 848611 Figs 5, 6

Diagnosis. Sidera tianshanensis is characterized by annual to perennial basidiocarps, measuring 15 mm thick, pores measuring 5–7 per mm, cream to rosy buff pore surface, allantoid basidiospores measuring 3–3.5 × 1–1.4 μ m, and growing on the stump or trunk of *Picea schrenkiana*.

Type. CHINA. Xinjiang Autonomous Region, Tekes County, Kosang Cave National Forest Park, on fallen trunk of *Picea schrenkiana*, 19 September 2021, Cui 19143 (holotype).

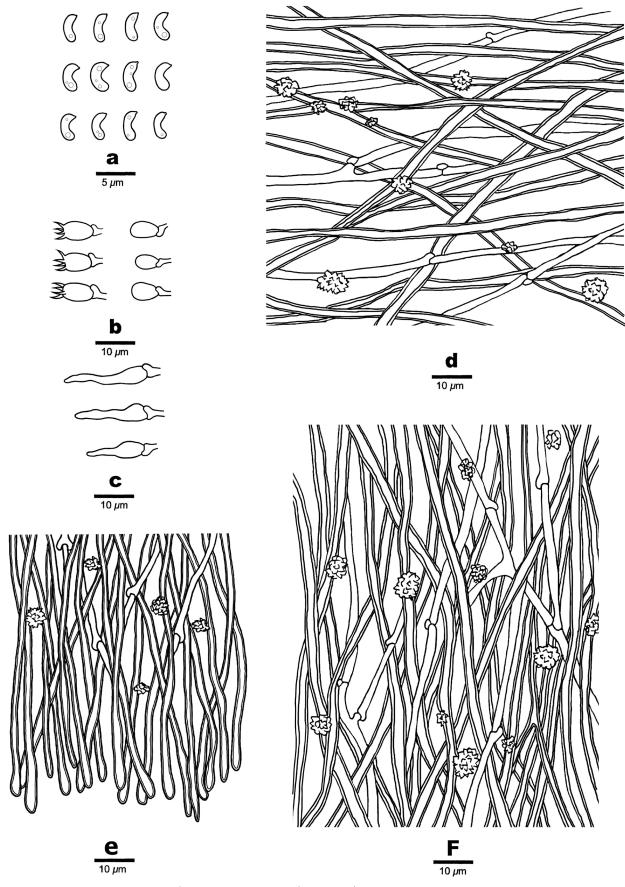
Etymology. tianshanensis (Lat.): referring to the species occurrence in Tianshan.

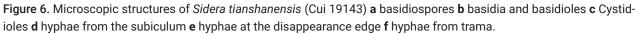
Fruiting body. Basidiocarps annual to perennial, resupinate, soft corky, up to 10 cm long, 5 cm wide, and 15 mm thick at the center; pore surface cream to buff yellow, uncracked; sterile margin indistinct, cottony, white, thinning out; pores angular, 5–7 per mm; dissepiments thin, entire; subiculum white, cottony and up to 0.1 mm thick; tubes concolorous with pore surface, up to 15 mm long.

Hyphal structure. Hyphal system dimitic; generative hyphae bearing clamp connections; rosette-like crystals frequently present; all hyphae IKI-, CB-; tissue unchanged in KOH.



Figure 5. Basidiocarp of Sidera tianshanensis (Cui 19143). Scale bar: 2.0 cm.





Subiculum. Generative hyphae infrequent, thin-walled, hyaline, occasionally branched, $2-2.5 \mu m$ in diameter; skeletal hyphae dominant, interwoven, unbranched, $2-3 \mu m$ diameter.

Tubes. Generative hyphae infrequent, thin-walled, hyaline, occasionally branched, $1.5-2.5 \mu m$ in diameter; skeletal hyphae dominant, thick-walled with a wide to medium lumen, hyaline, occasionally branched, interwoven, flexuous, $2-3 \mu m$ in diameter. Cystidia absent; cystidioles present, fusoid, thin-walled, hyaline, basally swollen, with hyphoid neck and sharp tip, $15-22 \times 3-4 \mu m$. Basidia barrel-shaped, hyaline, bearing four sterigmata and a basal clamp connection, $5.5-7 \times 3.5-4.5 \mu m$; basidioles pyriform, shorter than the basidia.

Spores. Basidiospores allantoid, hyaline, thin-walled, smooth, occasionally with one or more guttules, IKI-, CB-, $3-3.5 \times 1-1.4 \mu m$, L = $3.12 \mu m$, W = $1.18 \mu m$, Q = 2.6-2.7 (n = 150/5).

Type of rot. White rot.

Additional specimens (paratypes) examined. CHINA. Xinjiang Autonomous Region, Tekes County, Kosang Cave National Forest Park, on stump of *Picea schrenkiana*, 19 September 2021, Cui 19132; Tekes County, Karada Town, Qiongkushitai Village, on stump of *Picea schrenkiana*, 19 September 2021, Cui 19186, Cui 19192; on fallen trunk of *Picea schrenkiana*, 19 September 2021, Cui 19196, Cui 19251.

Discussion

In this study, phylogenetic trees of *Ceriporiopsis* and *Sidera* were constructed using combined ITS and nLSU sequences, respectively. The two newly proposed species formed separate branches on the phylogenetic trees with high support. In addition, both *Ceriporiopsis tianshanensis* and *Sidera tianshanensis* differ from other recorded species through their morphological characteristics.

According to our phylogenetic analyses of Ceriporiopsis based on the combined ITS+nLSU dataset, Ceriporiopsis tianshanensis was involved in Ceriporiopsis s.s. with strong support (100% ML, 100% MP, 1.00 BPPs) (Fig. 1). Ceriporiopsis subrufa was closely related to C. tianshanensis in the phylogenetic tree (Fig. 1), but there were obvious morphological differences between them. Ceriporiopsis subrufa is distinguished from C. tianshanensis by thicker basidiocarps (10 mm) and angular pores, and by growing on angiosperm trees (Ryvarden and Gilbertson 1993). Mophologically, Ceriporiopsis tianshanensis is similar to C. pseudoplacenta Vlasák & Ryvarden and C. aneirina (Sommerf.) Domański by white to cream to salmon-buff pore surface, and by broadly ellipsoid basidiospores of similar size (Ryvarden and Gilbertson 1993; Vlasák et al. 2012). However, Ceriporiopsis pseudoplacenta has thicker basidiocarps (6 mm), circular to angular pores and smaller basidiospores $(3.5-4.5 \times 2.2-3 \mu m)$ (Vlasák et al. 2012). The difference between Ceriporiopsis aneirina and C. tianshanensis is that the former has thicker basidiocarps (4 mm), generative hyphae with thin walls and crystals, and grows on angiosperm trees (Ryvarden and Gilbertson 1993).

The phylogenetic analysis of *Sidera* showed that *Sidera tianshanensis* was involved in *Sidera* s.s. with strong support (100% ML, 96% MP, 1.00 BPPs) (Fig. 2). In addition, *Sidera salmonea* was closely related to *S. tianshanensis* in the phylogenetic tree (Fig. 2). However, *Sidera salmonea* is distinguished from *S. tianshanensis* by its shape and size of the basidiospores (3–3.5 × 0.9–1.1 µm, Q = 3.03-3.21, lunate vs. $3-3.5 \times 1-1.4$ µm, Q = 2.6-2.7, allantoid), smaller

pores (7–9 per mm vs. 5–7 per mm), and surface of basidiocarps being salmon to slightly shiny, while that of the latter is cream to rosy buff (Liu et al. 2022). Morphologically, *Sidera tianshanensis* is similar to *S. parallela*, both presenting cream to buff yellow pore surfaces and the pore sizes were similar (Du et al. 2020). However, *Sidera parallela* differs from *S. tianshanensis* by its thinner basidiocarps (1.5 mm vs. 15 mm); in addition, *S. parallela* grows on angiosperm trees (Du et al. 2020), while *S. tianshanensis* grows on *Picea schrenkiana*.

Based on the records in previous literature and the introduction in this study, 42 species of *Ceriporiopsis* have been recorded in the world, among which 9 species are distributed in China (Binder et al. 2005; Zhao and Wu 2017; Ryvarden 2018, 2019, 2020; Zhao et al. 2023). *Ceriporiopsis* is widely distributed across five continents, with the exception of Antarctica and Oceania. The genus is most diverse in Africa, where it is represented by 17 species. South America has 10 species, North America has 7 species, Asia has 7 species, and Europe has 4 species. A total of 19 species of *Sidera* have been recorded worldwide, among which 10 species are distributed in China (Liu et al. 2023b). *Sidera* is currently a genus of fungi that has been relatively understudied. Among the discovered species that have been discovered so far, Asia has the highest number with 13 species, followed by Oceania with 4 species, Europe with 3 species, and North and South America with 1 species each. With the in-depth investigation of wood-inhabiting fungi in Xinjiang, an increasing number of new species of wood-inhabiting fungi will be discovered. The species diversity of wood-inhabiting fungi in China will also be richer.

Acknowledgments

We express our gratitude to Mr. Zheng-Xiang Qi (China) and Dr. Jun-Zhi Qiu (China) for their companionship during field collections.

Additional information

Conflict of interest

No conflict of interest was declared.

Ethical statement

No ethical statement was reported.

Funding

The research is supported by the Scientific and Technological Tackling Plan for the Key Fields of Xinjiang Production and Construction Corps (No. 2021AB004), the National Natural Science Foundation of China (Nos. U2003211, 32270010), and Beijing Forestry University Outstanding Young Talent Cultivation Project (No. 2019JQ03016).

Author contributions

Conceptualization, Y.-F.S.; and T.-M.X.; methodology, T.-M.X.; software, S.L.; validation, S.L.; C.-G.S.; formal analysis, T.-M.X.; investigation, T.-M.X.; Y.-F.S.; C.-G.S.; S.L.; N.G.; D.-M. W. and B.-K.C. resources, B.-K.C.; data curation, Y.-F.S.; and T.-M.X.; writing–original draft preparation, T.-M.X.; writing–review and editing, Y.-F.S. and B.-K.C.; visualization, T.-M.X.; supervision, B.-K.C.; project administration, B.-K.C. and D.-M.W.; funding acquisition, B.-K.C.; D.-M.W. and N.G.. All authors have read and agreed to the published version of the manuscript.

Author ORCIDs

Tai-Min Xu [©] https://orcid.org/0000-0002-5230-4140 Yi-Fei Sun [©] https://orcid.org/0000-0003-3997-3662 Shun Liu [©] https://orcid.org/0000-0001-9261-4365 Chang-Ge Song [©] https://orcid.org/0000-0001-5379-2353 Neng Gao [©] https://orcid.org/0009-0000-4745-987X Dong-Mei Wu [©] https://orcid.org/0009-0006-4126-0767 Bao-Kai Cui [©] https://orcid.org/0000-0003-3059-9344

Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

References

- Bau T, Hu JW, Zhou ZB, Xu B (2008) The new distribution of macrofungi in Xinjiang. Journal of Tarim University 20: 4.
- Binder M, Hibbett DS, Larsson KH, Larsson E, Langer E, Langer G (2005) The phylogenetic distribution of resupinate forms across the major clades of mushroom-forming fungi (Homobasidiomycetes). Systematics and Biodiversity 3(2): 113–157. https://doi. org/10.1017/S1477200005001623
- Chen JJ, Cui BK, Dai YC (2016) Global diversity and molecular systematics of *Wrightoporia* s.l. (Russulales, Basidiomycota). Persoonia 37(1): 21–36. https://doi. org/10.3767/003158516X689666
- Cheng T, Kolarik M, Quijada L, Stadler M (2022) A re-assessment of *Taxomyces andreanae*, the alleged taxol-producing fungus, using comparative genomics. IMA Fungus 13(1): 17. https://doi.org/10.1186/s43008-022-00103-4
- Cui BK, Li HJ, Ji X, Zhou JL, Song J, Si J, Yang ZL, Dai YC (2019) Species diversity, taxonomy and phylogeny of Polyporaceae (Basidiomycota) in China. Fungal Diversity 97(1): 137–392. https://doi.org/10.1007/s13225-019-00427-4
- Dai YC (2010) Hymenochaetaceae (Basidiomycota) in China. Fungal Diversity 45(1): 131-343. https://doi.org/10.1007/s13225-010-0066-9
- Dai YC (2012) Polypore diversity in China with an annotated checklist of Chinese polypores. Mycoscience 53(1): 49–80. https://doi.org/10.47371/s10267-011-0134-3
- Dai YC, Wei YL, Yuan HS, Huang MY, Penzina T (2007) Polypores from Altay and Tian Mts. in Xinjiang, northwest China. Cryptogamie. Mycologie 28: 269–279.
- Domański S (1963) Two new genera of fungi from group *Poria* Pers. Ex S.F. Gray. Acta Societatis Botanicorum Poloniae 32: 731–739. https://doi.org/10.5586/asbp.1963.044
- Du R, Wu F, Gate GM, Dai YC, Tian XM (2020) Taxonomy and phylogeny of Sidera (Hymenochaetales, Basidiomycota): Four new species and keys to species of the genus. MycoKeys 68: 115–135. https://doi.org/10.3897/mycokeys.68.53561
- Felsenstein J (1985) Confidence intervals on phylogenetics: An approach using the bootstrap. Evolution; International Journal of Organic Evolution 39(4): 783–791. https://doi.org/10.2307/2408678
- Gilbertson RL, Ryvarden L (1987) North American Polypores 2. *Megasporoporia Wrightoporia*. Fungiflora, Oslo.
- Guli A, Feng L, Qin XZ, Chen J, Marhaba, Yang XP, Liu AM, Wang ZH (2015) Investigation on wild fungi resources in Habahe Plain of Altai Prefecture (I). Xinjiang Nongye Kexue 52: 1707–1714.

- Hall TA (1999) Bioedit: A user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98. https://doi.org/10.1021/bk-1999-0734.ch008
- Hu W, Yao J, He Q, Chen J (2021) Changes in precipitation amounts and extremes across Xinjiang (northwest China) and their connection to climate indices. PeerJ 9: e10792. https://doi.org/10.7717/peerj.10792
- Huang J, Liu C, Guo Z, Ma K, Zang R, Ding Y, Lu X, Wang J, Yu R (2018) Seed plant features, distribution patterns, diversity hotspots, and conservation gaps in Xinjiang, China. Nature Conservation 27: 1–15. https://doi.org/10.3897/natureconservation.27.23728
- Ji X, Zhou JL, Song CG, Xu TM, Wu DM, Cui BK (2022) Taxonomy, phylogeny and divergence times of *Polyporus* (Basidiomycota) and related genera. Mycosphere: Journal of Fungal Biology 13(1): 1–52. https://doi.org/10.5943/mycosphere/13/1/1
- Li HJ, Cui BK, Dai YC (2014) Taxonomy and multi-gene phylogeny of *Datronia* (Polyporales, Basidiomycota).Persoonia32(1):170–182.https://doi.org/10.3767/003158514X681828
- Liu S, Han ML, Xu TM, Wang Y, Wu DM, Cui BK (2021a) Taxonomy and phylogeny of the *Fomitopsis pinicola* complex with descriptions of six new species from East Asia. Frontiers in Microbiology 12: 644979. https://doi.org/10.3389/fmicb.2021.644979
- Liu S, Chen YY, Sun YF, He XL, Song CG, Si J, Liu DM, Gates G, Cui BK (2023a) Systematic classification and phylogenetic relationships of the brown-rot fungi within the Polyporales. Fungal Diversity 118(1): 1–94. https://doi.org/10.1007/s13225-022-00511-2
- Liu ZB, Zhou HM, Liu HG, Yuan Y (2023b) Taxonomy and phylogeny of Sidera (Hymenochaetales, Rickenella clade) from China and North America revealing two new species. MycoKeys 96: 173–191. https://doi.org/10.3897/mycokeys.96.100743
- Liu ZB, Zhou M, Yuan Y, Dai YC (2021b) Global diversity and taxonomy of *Sidera* (Hymenochaetales, Basidiomycota): Four new species and keys to species of the genus. Journal of Fungi (Basel, Switzerland) 7(4): 251. https://doi.org/10.3390/jof7040251
- Liu ZB, Zhou M, Wu F, Yu J (2022) Two new species of *Sidera* (Hymenochaetales, Basidiomycota) from Southwest China. Journal of Fungi 8(4): 385. https://doi.org/10.3390/jof8040385
- Miettinen O, Larsson KH (2011) *Sidera*, a new genus in Hymenochaetales with poroid and hydnoid species. Mycological Progress 10(2): 131–141. https://doi.org/10.1007/ s11557-010-0682-5
- Niemelä T (1985) Mycoflora of Poste-de-la-Baleine, Northern Quebec. Polypores and the Hymenochaetales. Nature Cancer 112: 445–472.
- Núñez M, Ryvarden L (2001) East Asian polypores 2. Synopsis Fungorum 14: 165-522.
- Page RDM (1996) TreeView: Application to display phylogenetic trees on personal computers. Bioinformatics 12(4): 357–358. https://doi.org/10.1093/bioinformatics/12.4.357
- Petersen JH (1996) Farvekort. The Danish Mycological Society's color-chart. Foreningen til Svampekundskabens Fremme, Greve.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hőhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029
- Ryvarden L (2018) Studies in African Aphyllophorales 23 Some new species in *Ceriporiopsis* and *Diplomitoporus*. Synopsis Fungorum 38: 1–67.
- Ryvarden L (2019) Studies in African poroid fungi 30 Some poroid species from Uganda. Synopsis Fungorum 39: 41–45.

- Ryvarden L (2020) Studies in African Aphyllophorales 40 Some new polypores from Cameroon. Synopsis Fungorum 41: 1–49.
- Ryvarden L, Gilbertson RL (1993) European polypores. Part 1. Synopsis Fungorum 6: 1–387.
- Ryvarden L, Melo I (2014) Poroid fungi of Europe. Synopsis Fungorum 31: 1-455.
- Shen LL, Wang M, Zhou JL, Xing JH, Cui BK, Dai YC (2019) Taxonomy and phylogeny of Postia. Multi-gene phylogeny and taxonomy of the brown-rot fungi: Postia and its related genera. Persoonia 42: 101–126. https://doi.org/10.3767/persoonia.2019.42.05
- Song CG, Chen YY, Liu S, Xu TM, He XL, Wang D, Cui BK (2022) A phylogenetic and taxonomic study on *Phellodon* (Bankeraceae, Thelephorales) from China. Journal of Fungi 8(5): 429. https://doi.org/10.3390/jof8050429
- Song J, Chen YY, Cui BK, Liu HG, Wang YZ (2014) Morphological and molecular evidence for two new species of *Laetiporus* (Basidiomycota, Polyporales) from southwestern China. Mycologia 106(5): 1039–1050. https://doi.org/10.3852/13-402
- Spirin V, Ryvarden L (2016) Some basidiomycetes (Aphyllophorales) from Mexico. Synopsis Fungorum 35: 34–42.
- Sun YF, Xing JH, He XL, Wu DM, Song CG, Liu S, Vlasák J, Gates G, Gibertoni TB, Cui BK (2022) Species diversity, systematic revision and molecular phylogeny of Ganodermataceae (Polyporales, Basidiomycota) with an emphasis on Chinese collections. Studies in Mycology 101(1): 287–415. https://doi.org/10.3114/sim.2022.101.05
- Swofford DL (2002) PAUP*: phylogenetic analysis using parsimony (*and other methods), version 4.0b10. Sinauer Associates, Sunderland 56(9): 1776–1788. https://doi. org/10.1111/j.0014-3820.2002.tb00191.x
- Thompson JD, Gibson TJ, Plewniak F, Franois J, Higgins DG (1997) The CLUSTAL X windows interface: Flexible strategies for multiple sequence alignment aided by quality analysis tools. Nucleic Acids Research 25(24): 4876–4882. https://doi.org/10.1093/ nar/25.24.4876
- Tomšovský M, Menkis A, Vasaitis R (2010) Phylogenetic relationships in European *Ceriporiopsis* species inferred from nuclear and mitochondrial ribosomal DNA sequences. Fungal Biology 114(4): 350–358. https://doi.org/10.1016/j.funbio.2010.02.004
- Vlasák J, Vlasák Jr JJ, Ryvarden L (2012) Four new polypore species from the western United States. Mycotaxon 119(1): 217–231. https://doi.org/10.5248/119.217
- Wang JY, Ayinuer (2004) Macrofungi of Hanasi Lake Rigion from MT. Altal. Journal of Xinjiang University 21: 88–91. [Natural Science Edition]
- Wang XW, May TW, Liu SL, Zhou LW (2021) Towards a natural classification of *Hyphodontia* sensu lato and the trait evolution of basidiocarps within Hymenochaetales (Basidiomycota). Journal of Fungi 7(6): 478. https://doi.org/10.3390/jof7060478
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR protocols: a guide to methods and applications. Academic, San Diego, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1
- Wu F, Man XW, Tohtirjap A, Dai YC (2022a) A comparison of polypore funga and species composition in forest ecosystems of China, North America, and Europe. Forest Ecosystems 9: 100051. https://doi.org/10.1016/j.fecs.2022.100051
- Wu F, Zhou LW, Vlasák J, Dai YC (2022b) Global diversity and systematics of Hymenochaetaceae with poroid hymenophore. Fungal Diversity 113(1): 1–192. https://doi. org/10.1007/s13225-021-00496-4

- Wu ZT, Zhang HJ, Krause CM, Cobb NS (2010) Climate change and human activities: A case study in Xinjiang, China. Climatic Change 99(3–4): 457–472. https://doi. org/10.1007/s10584-009-9760-6
- Yuan Y, Shen LL (2017) Morphological characters and molecular data reveal a new species of *Rhodonia* (Polyporales, Basidiomycota) from China. Phytotaxa 328(2): 175– 182. https://doi.org/10.11646/phytotaxa.328.2.8
- Yuan Y, Chen JJ, Korhonen K, Martin F, Dai YC (2021) An updated global species diversity and phylogeny in the forest pathogenic genus *Heterobasidion* (Basidiomycota, Russulales). Frontiers in Microbiology 11: 596393. https://doi.org/10.3389/ fmicb.2020.596393
- Zhang HX, Zhang ML (2014) Insight into distribution patterns and conservation planning in relation to woody species diversity in Xinjiang, arid northwestern China. Biological Conservation 177: 165–173. https://doi.org/10.1016/j.biocon.2014.07.005
- Zhao ZX (2022) Diversity of macrofungi in Tomur National Nature Reserve and molecular systematics of important taxa. Master's Thesis, Tarim University, China, Xinjiang.
- Zhao CL, Cui BK (2014) Phylogeny and taxonomy of *Ceriporiopsis* (Polyporales) with descriptions of two new species from southern China. Phytotaxa 164(1): 17–28. https://doi.org/10.11646/phytotaxa.164.1.2
- Zhao CL, Wu F, Liu HX, Dai YC (2015) A phylogenetic and taxonomic study on *Ceriporiopsis* s. str. (Polyporales) in China. Nova Hedwigia 101(3–4): 403–417. https://doi.org/10.1127/nova_hedwigia/2015/0282
- Zhao CL, Wu ZQ (2017) *Ceriporiopsis kunmingensis* sp. nov. (Polyporales, Basidiomycota) evidenced by morphological characters and phylogenetic analysis. Mycological Progress 16(1): 93–100. https://doi.org/10.1007/s11557-016-1259-8
- Zhao CL, Qu MH, Huang RX, Karunarathna SC (2023) Multi-gene phylogeny and taxonomy of the wood-rotting fungal genus *Phlebia* sensu lato (Polyporales, Basidiomycota). Journal of Fungi 9(3): 320. https://doi.org/10.3390/jof9030320
- Zhu L, Ji X, Si J, Cui BK (2019) Morphological characters and phylogenetic analysis reveal a new species of *Phellinus* with hooked hymenial setae from Vietnam. Phytotaxa 1(1): 91–99. https://doi.org/10.11646/phytotaxa.356.1.8
- Zmitrovich IV (2018) Conspectus Systematis Polyporacearum v. 1.0. Folia Cryptogamica Petropolitana 6: 1–145.